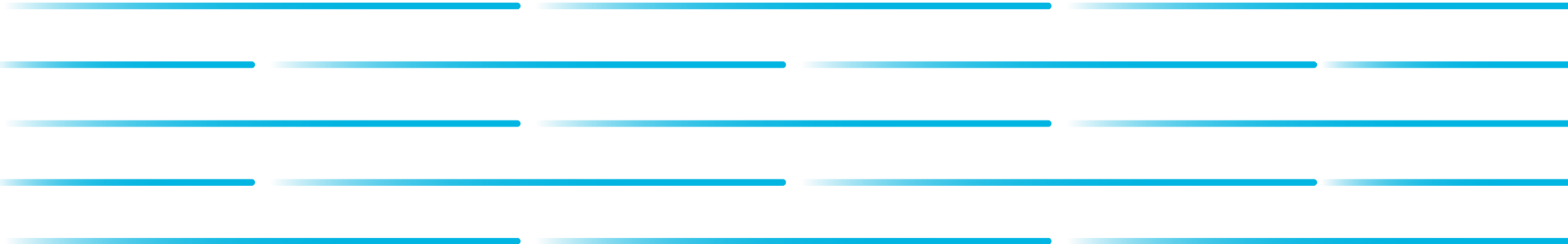




The future of cell based vaccine production

Mats Lundgren, Customer Applications Director, GE Healthcare



Vaccine production technology trends



Vaccine bioprocess technology

Platform technologies applied where possible (e.g., cell expansion on microcarriers and purification by chromatography)

Single-use technologies and automated solutions

Updated cell substrates—from eggs and diploid cells to continuous cell lines

Live viral vector production—need for efficient platforms

Process economy modelling implemented early in process development

Focus on analytical technologies driven by increased regulatory requirements



Vaccine production today

Processes developed decades ago

Old cell substrates or eggs
Limited purification
Significant expertise required

Processes difficult to scale up

Centrifugation
Fixed installations
Roller bottles

Unfavorable process economy

Low yields
Long process times
Labor-intense processes
Dedicated facilities

Increased regulatory requirements

Open handling
Batch variability
Serum supplementation



Vaccine production tomorrow

Processes developed decades ago

Platform cell lines

Efficient purification
based on
chromatography

Processes difficult to scale up

Scalable technologies
enabled by, e.g.,
single-use
technologies

Unfavorable process economy

Efficient and rational
process design

Flexible facilities

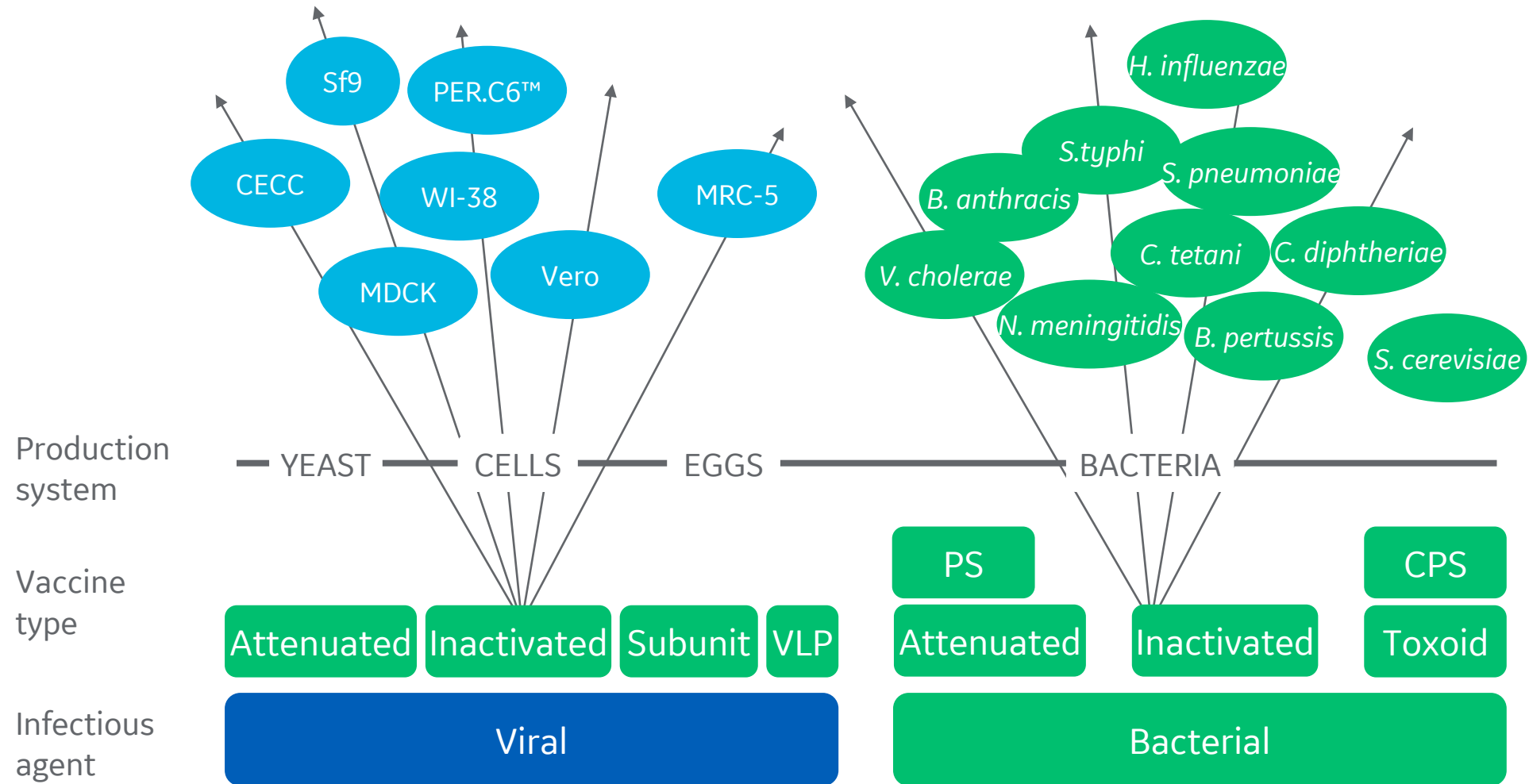
Increased regulatory requirements

Closed handling
Quality by design (QbD)
Chemically defined cell
culture media



Cell culture and virus propagation

Cell substrates for vaccines

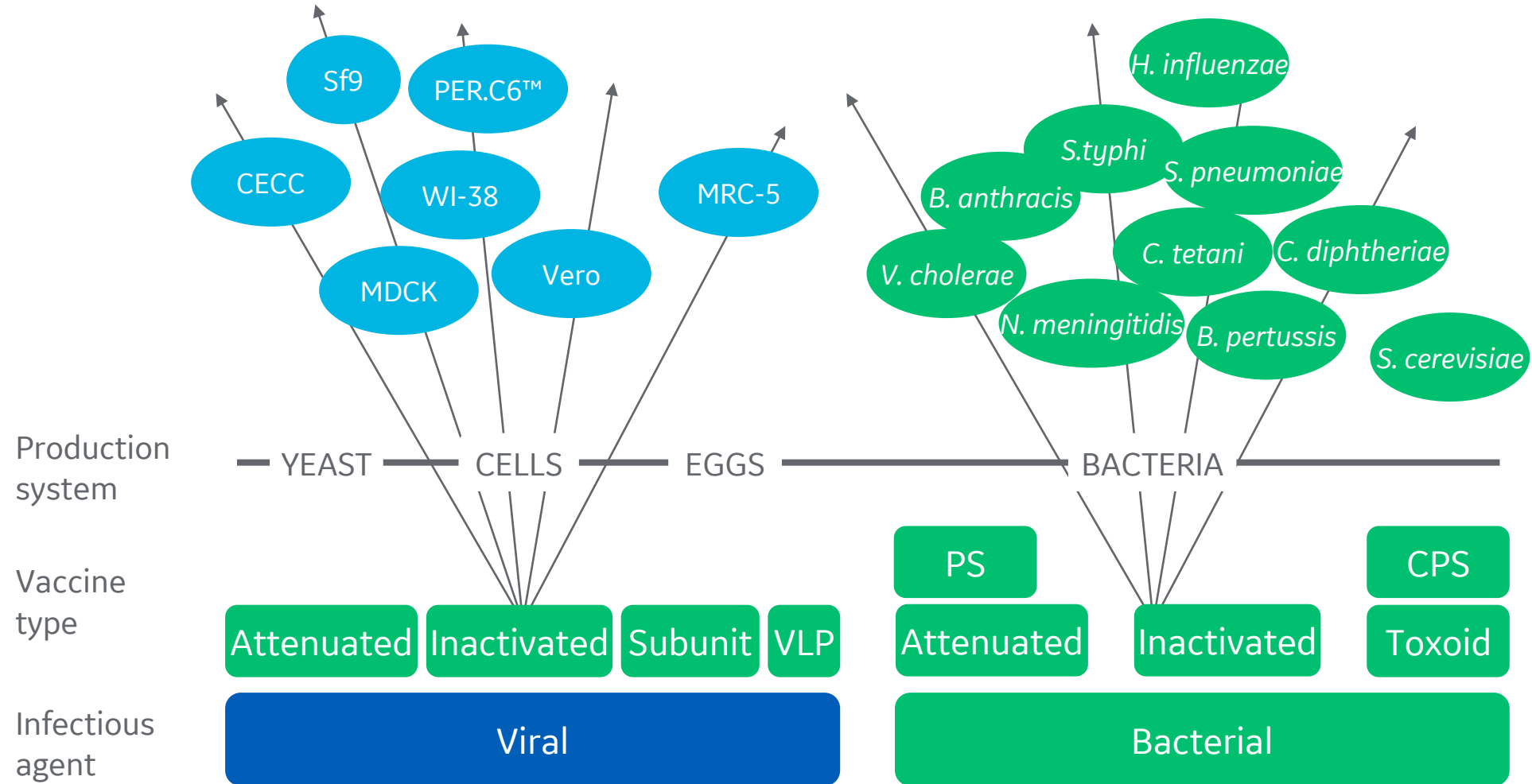


CPS = conjugated polysaccharides, PS = polysaccharides, VLP = virus-like particle



40 vaccines still to be developed

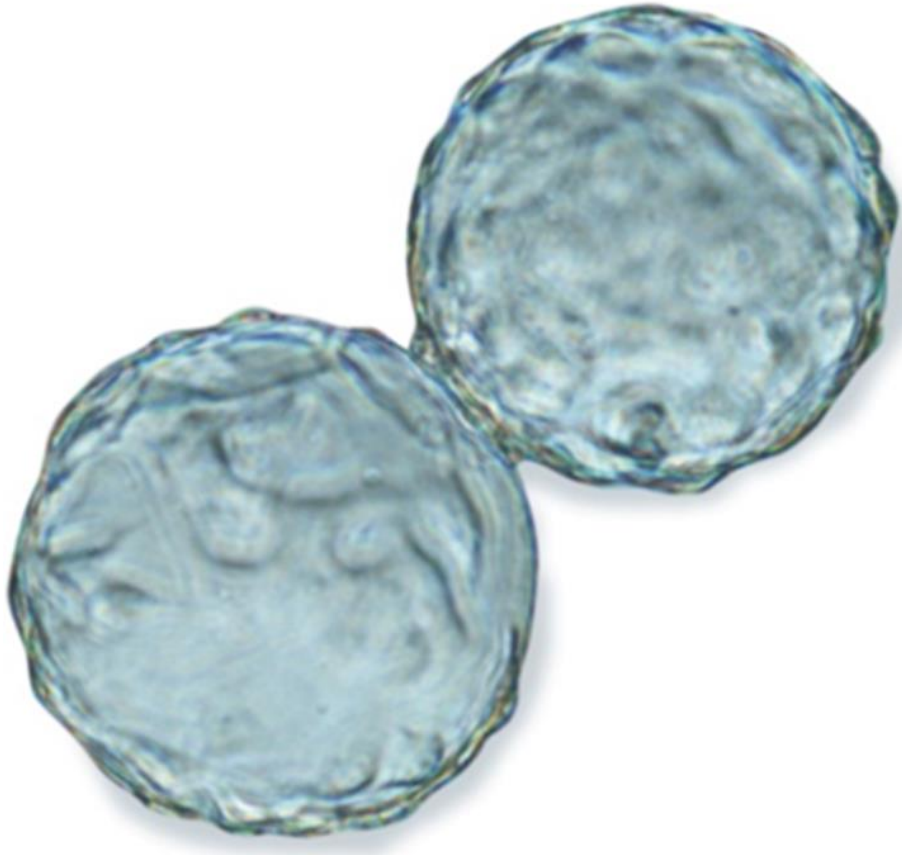
Where would this trend lead?



CPS = conjugated polysaccharides, PS = polysaccharides, VLP = virus-like particle



Selecting a cell line for virus production



Cell substrate evolution from primary to diploid to continuous cell lines

Modern options: Vero, MDCK, EBx, AGE, PER.C6™...

Requirements

- Suitable for GMP production
- Good safety track record
- Animal-origin free media preferred
- Good virus propagation
- Broadly and highly permissive
- Scalable to high-volume production



Cell culture medium and serum



Serum—ensure quality, traceability, and origin

Classical medium

Animal-origin free media

Complex media containing hydrolysates

Chemically defined media



Scale-up of adherent and suspension cells

Adherent cells

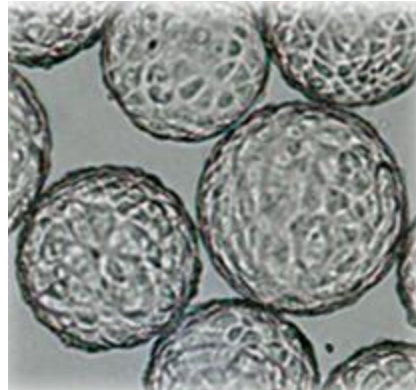
Cell growth is limited by surface area

Need enzymatic passaging

More complex scale-up

Higher virus production/cell

Microcarriers increase volumetric output by maximizing the surface to volume ratio for adherent cells



Suspension cells

Cell growth is limited by cell concentration in medium

Easier passage and scale-up

Lower virus production/cell



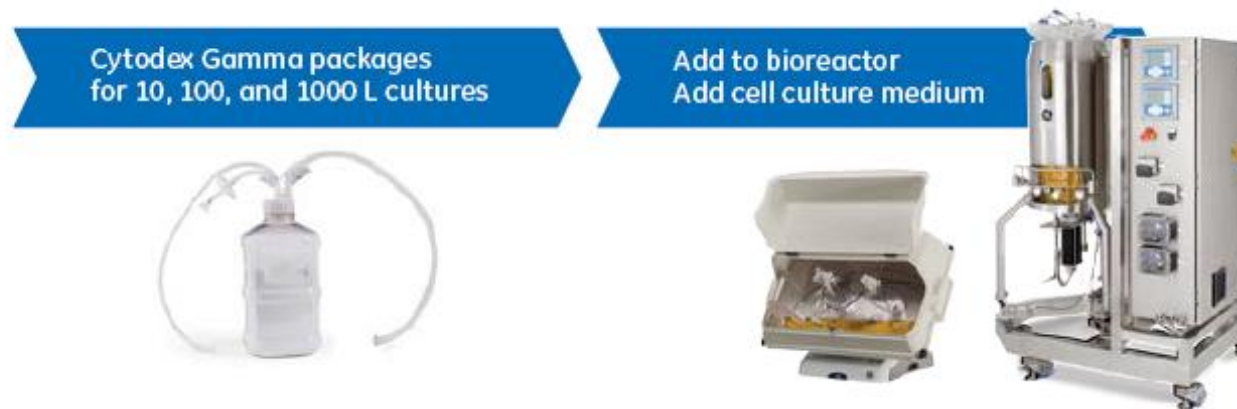
Introduction to Cytodex™ 1 and 3 Gamma microcarriers

Delivered gamma-sterilized and ready to use. Supplied dry to save storage space and facilitate transportation.

Conventional process:

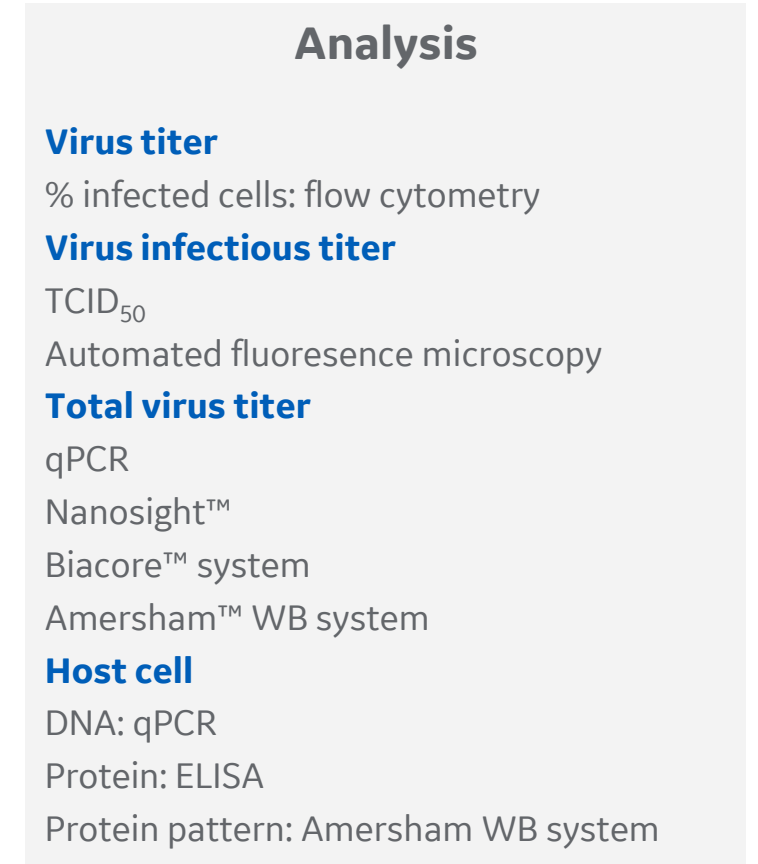
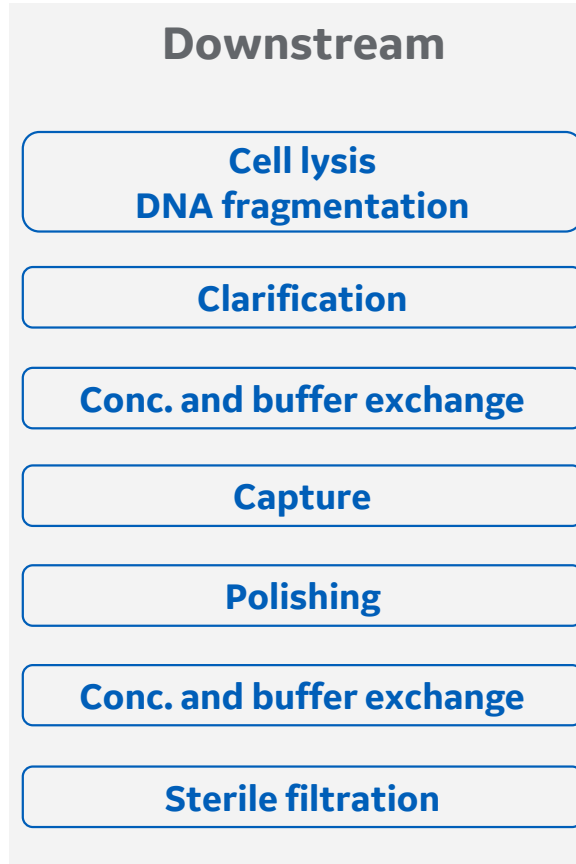
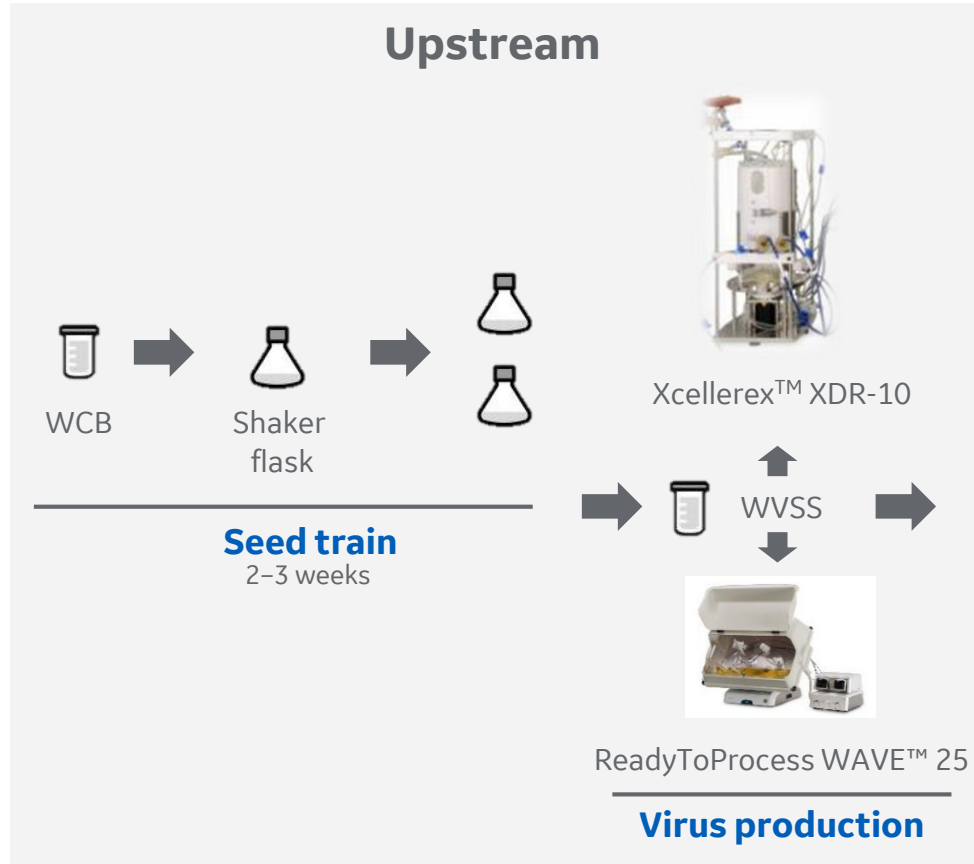


Simplified process:



Adenovirus vector

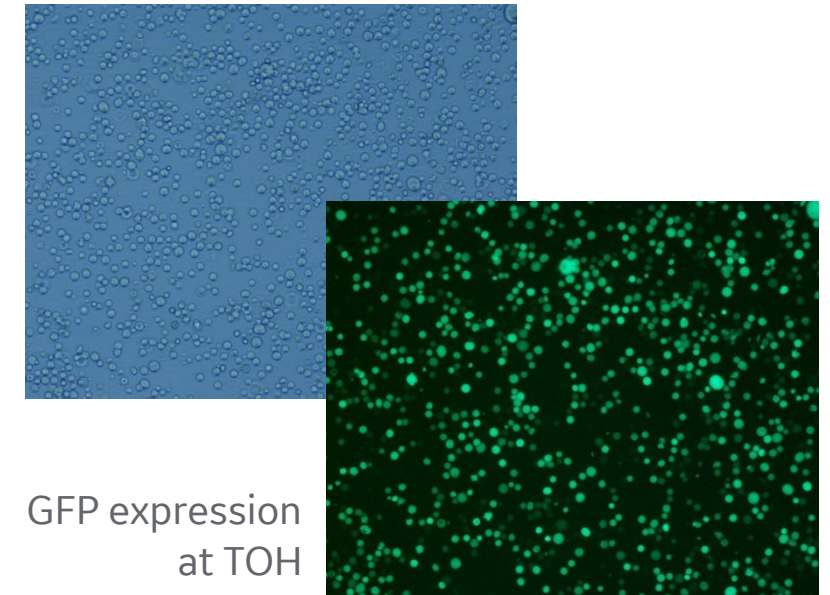
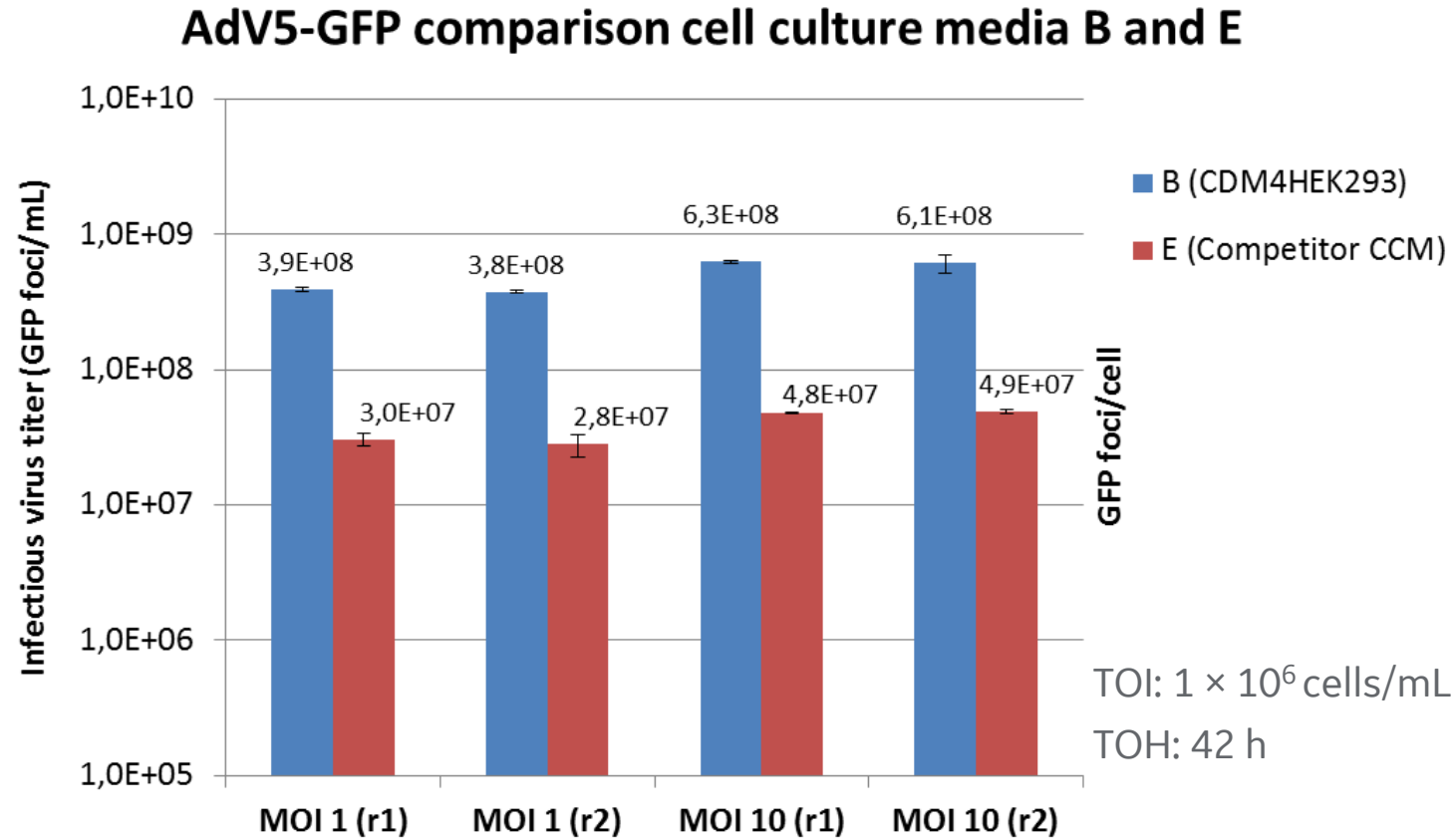
AV vaccine production process



TCID₅₀ = 50% tissue culture infective dose, WCB = working cell bank, WVSS = working viral seed stock



AdV productivity in CCM B (CDM4HEK293) vs E (competitor)



GFP expression
at TOH

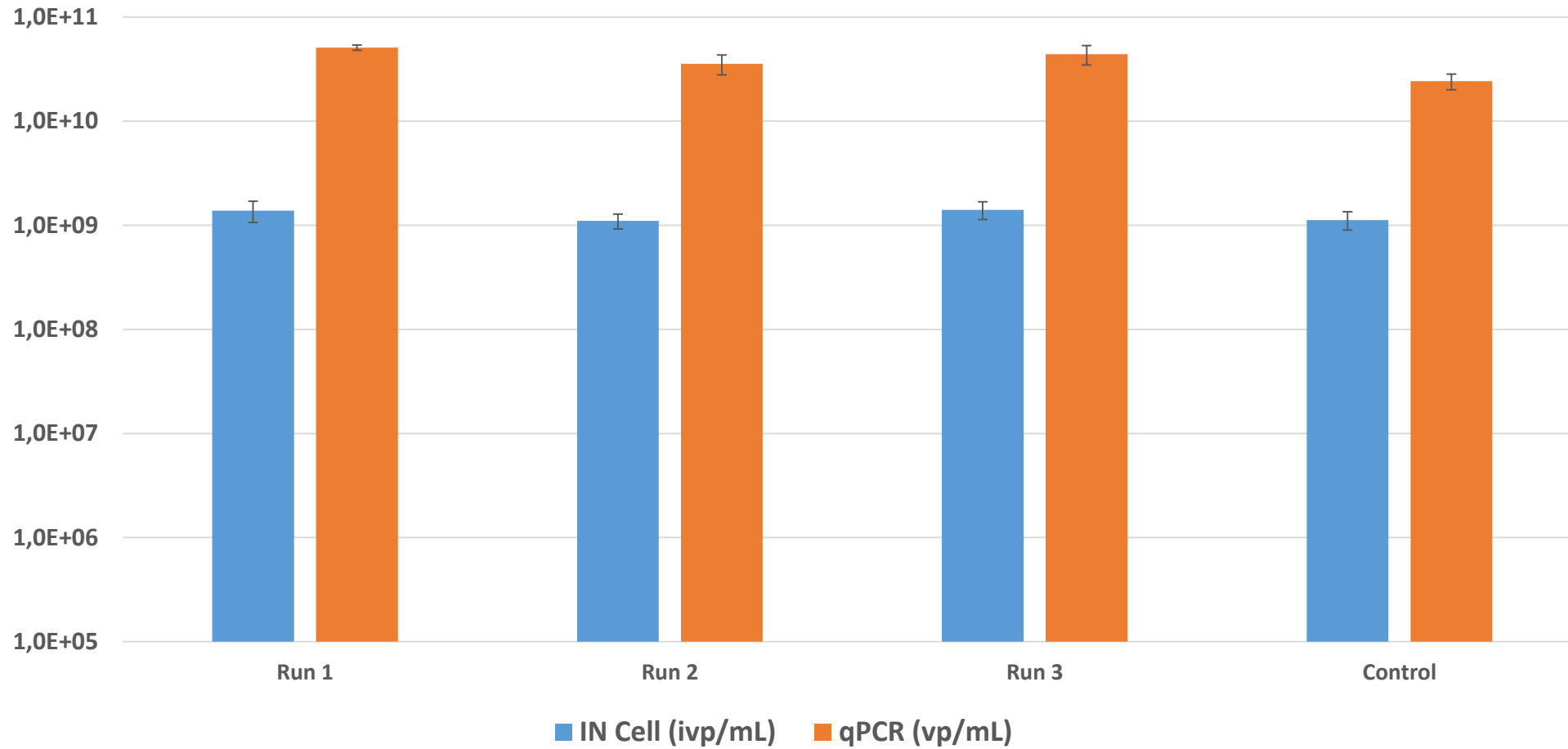


HyClone™ CDM4HEK293 media

AdV = adenovirus
CCM = cell culture medium
GFP = green fluorescent protein
MOI = multiplicity of infection
TOI = time of infection
TOH = time of harvest



Consistent adenovirus production in single-use Xcellerex™ XDR-10 bioreactor system

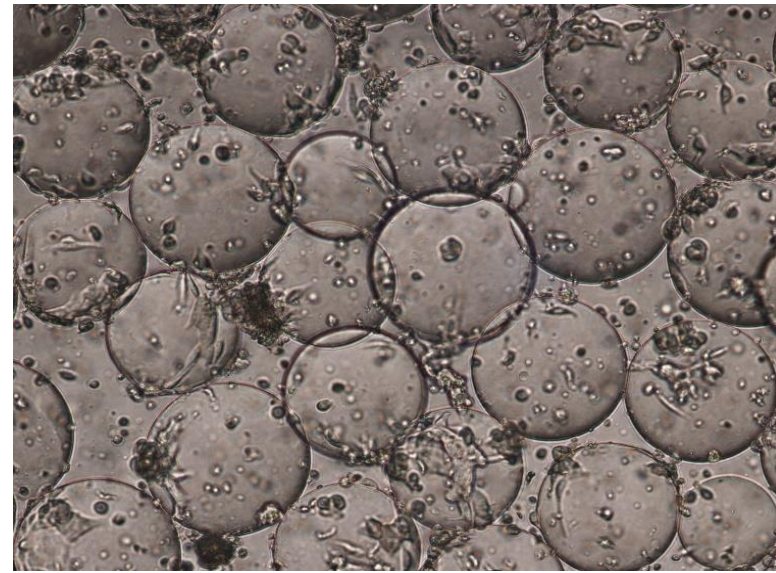
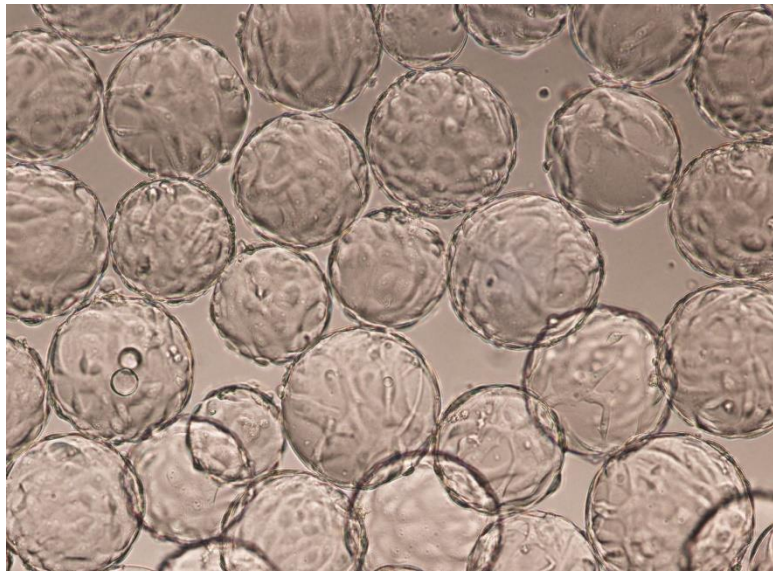


Rotavirus

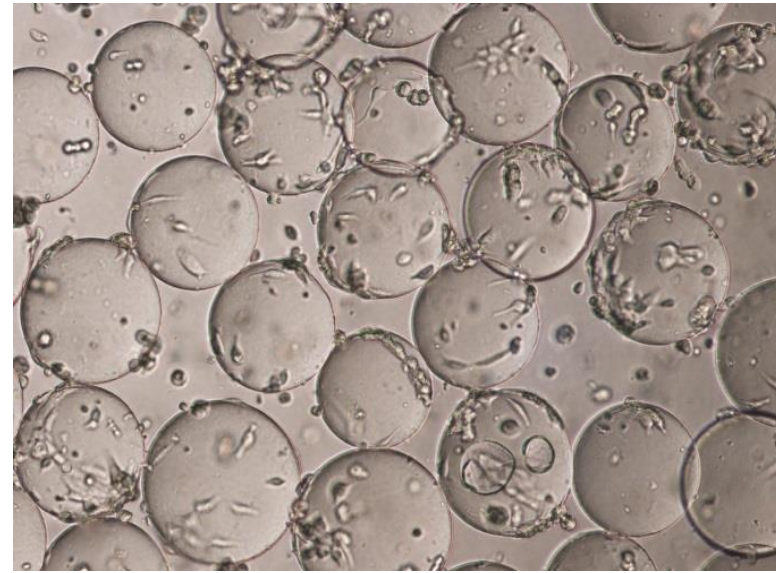
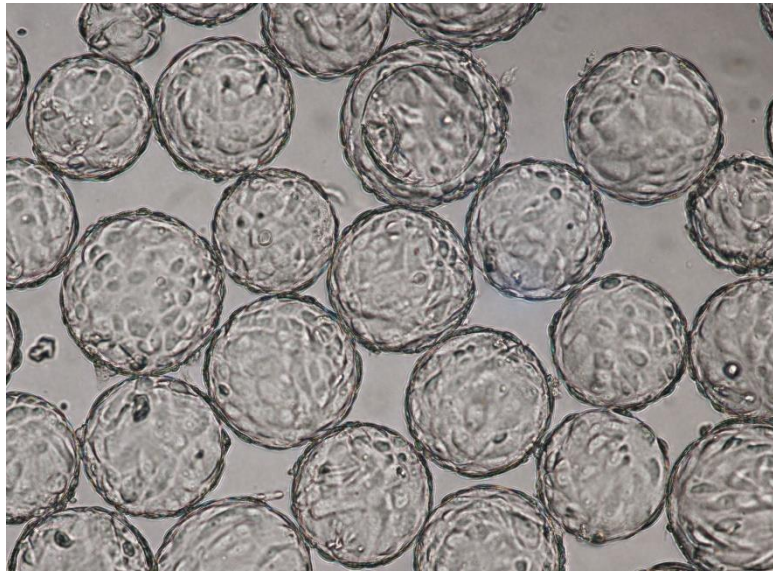
Rotavirus vaccines

- Common cause of diarrheal disease in young children
- 200 000 deaths in children under 5 years of age annually, majority in Africa and Asia (data estimated from 2013)
- Vaccines on the market: eg. Merck, GSK, Bharat and Lanzhou
- Limited efficacy in developing countries
- Live attenuated oral vaccines produced in Vero cells
- Vaccines produced by old technology in T-flasks / Roller bottles using animal derived components (serum and trypsin)





Competitor
medium



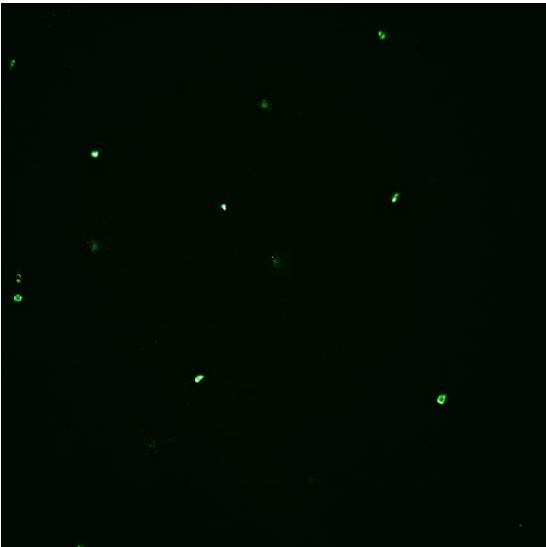
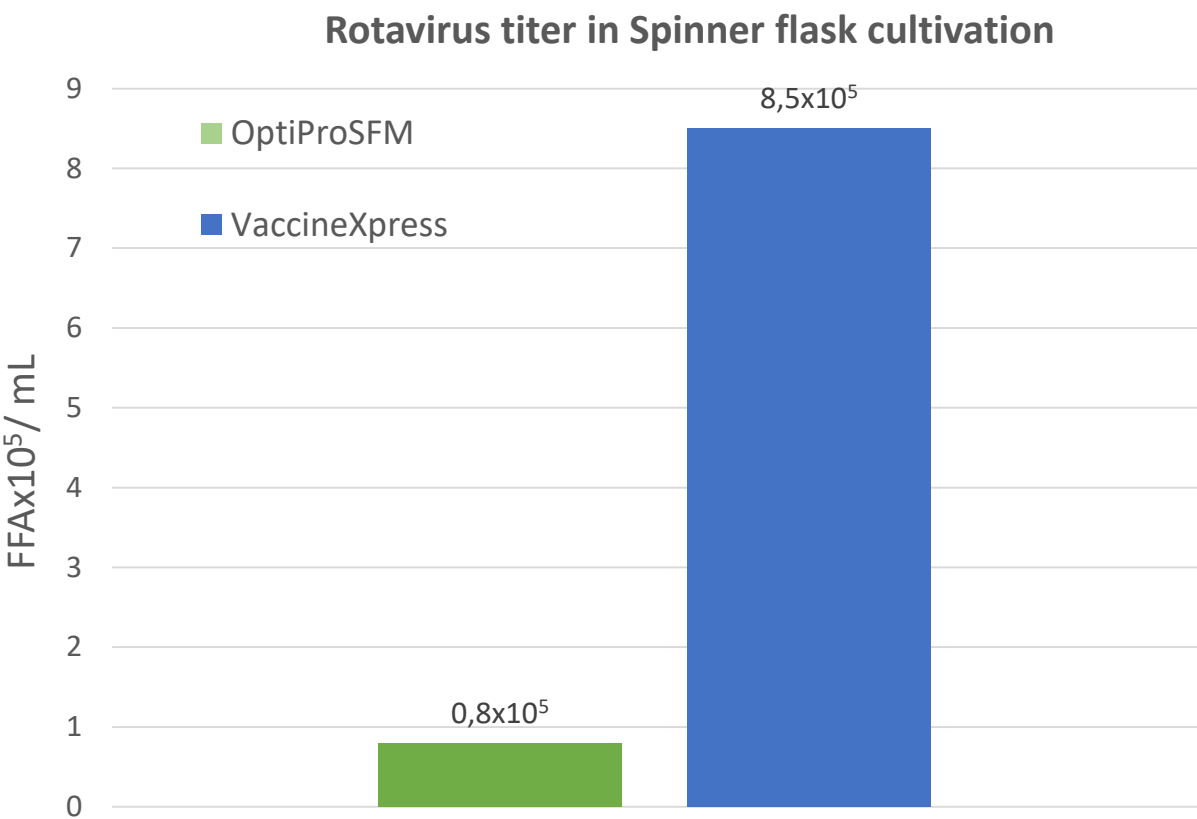
VaccineXpress
medium

72 h post Cytodex
inoculation

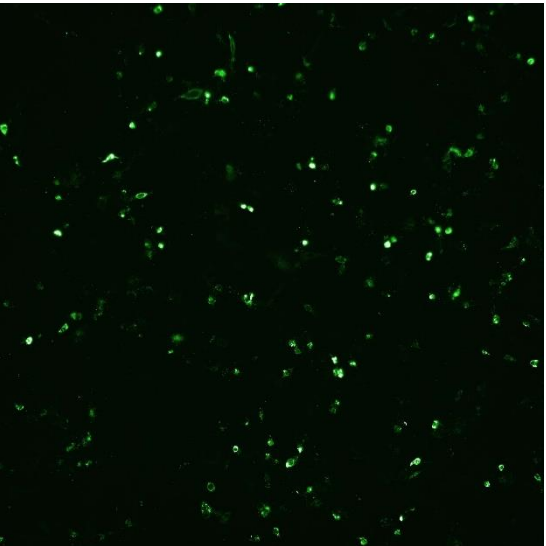
Time of
harvest



Rotavirus can be propagated on Cytodex 1 Gamma using VaccineXpress



Competitor medium



VaccineXpress medium

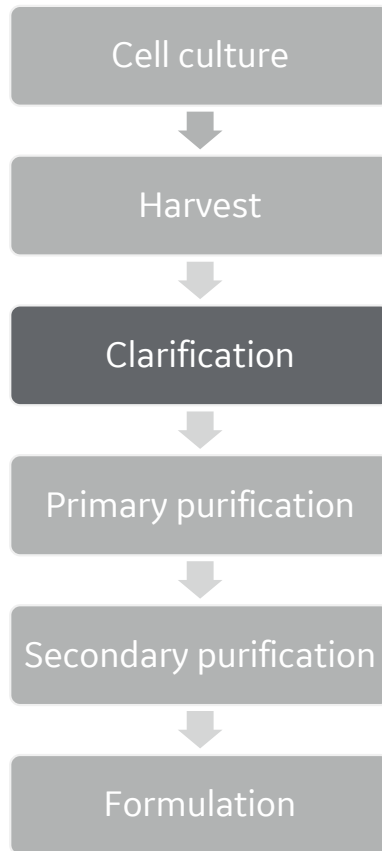
Rotavirus expression (IN Cell)



Virus purification

Clarification

Process flow



Available techniques

Filtration

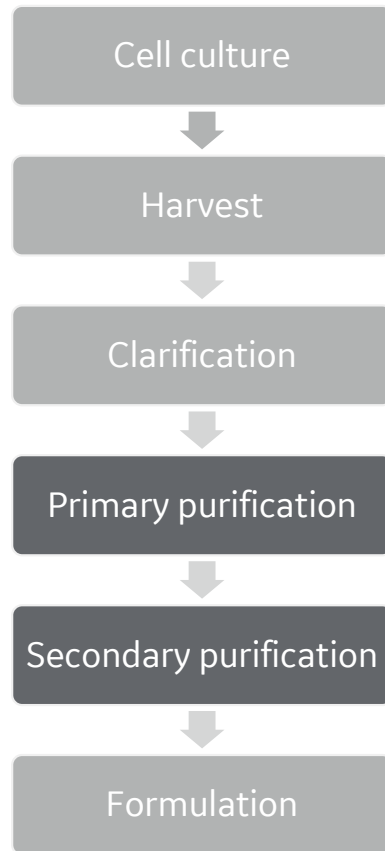
- Normal flow
- Tangential flow (TFF)—hollow fiber filters

Centrifugation



Purification

Process flow



Available techniques

TFF—hollow fiber filters

Density gradient centrifugation

Selective precipitation

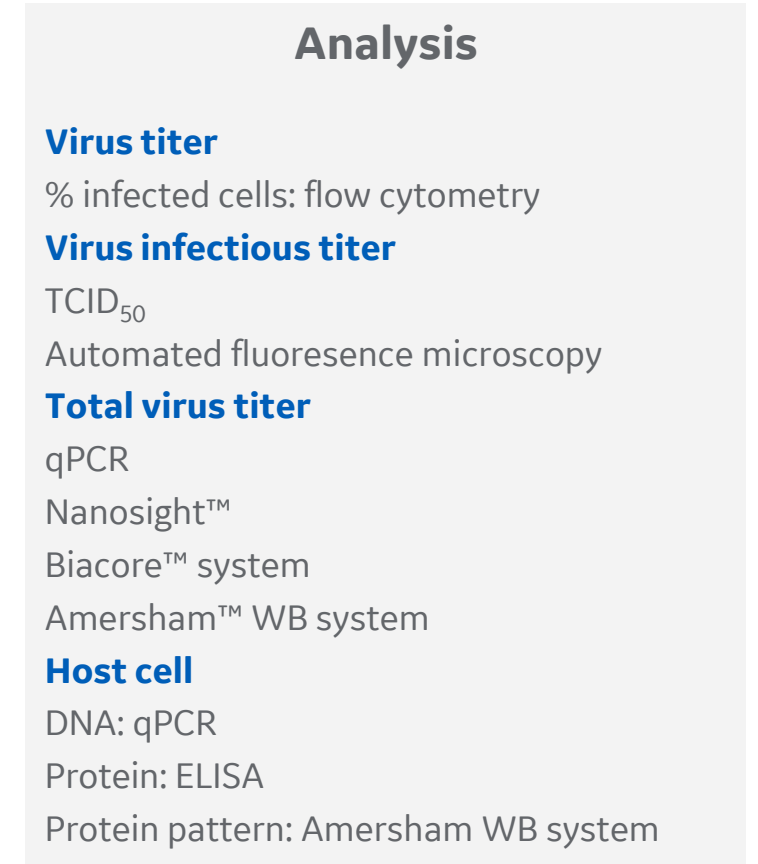
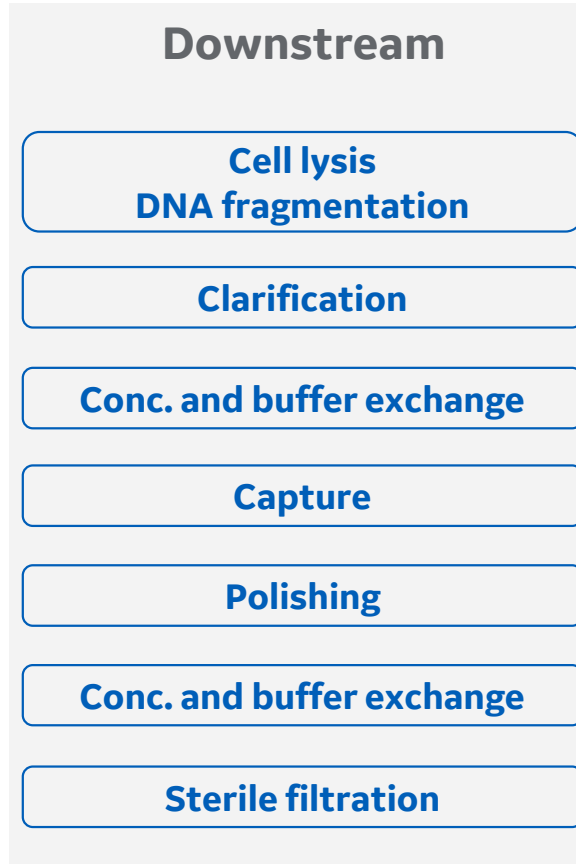
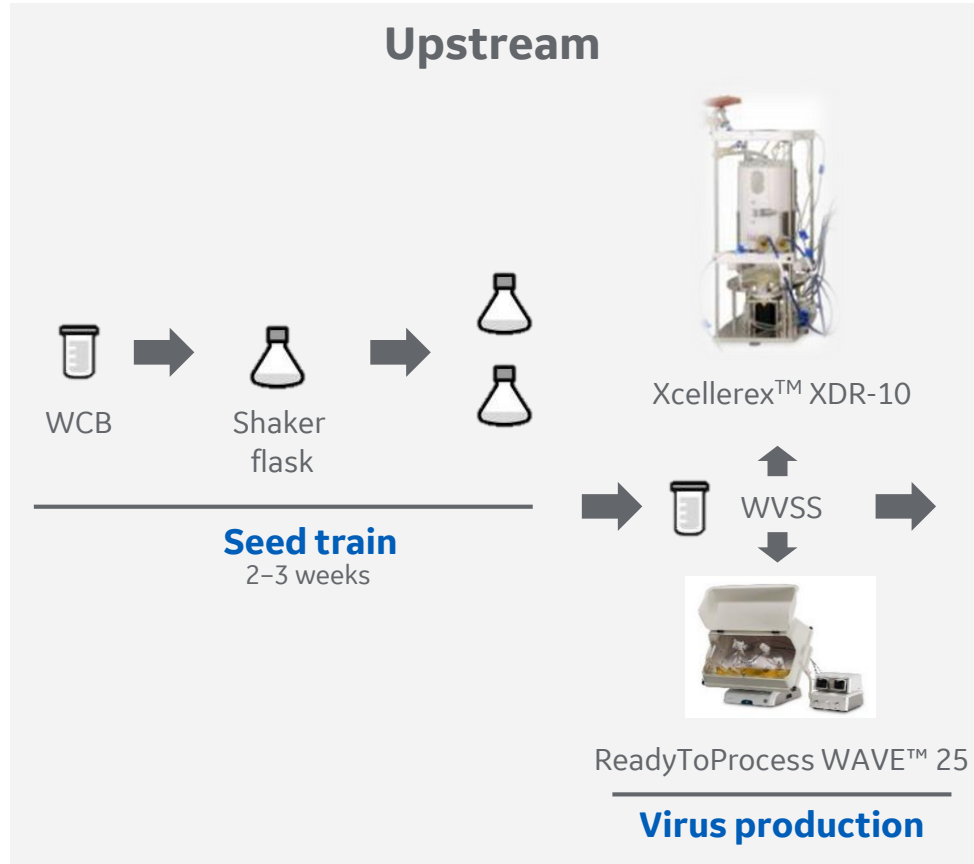
Chromatography

- IEX, MM, AC, HIC, SEC
- Resin format (packed bed)
- Membrane format (capsule), ReadyToProcess™ Adsorber Q

AC = affinity chromatography, HIC = hydrophobic interaction chromatography, IEX = ion exchange chromatography, MM = multimodal chromatography, SEC = size exclusion chromatography



AV vaccine production process



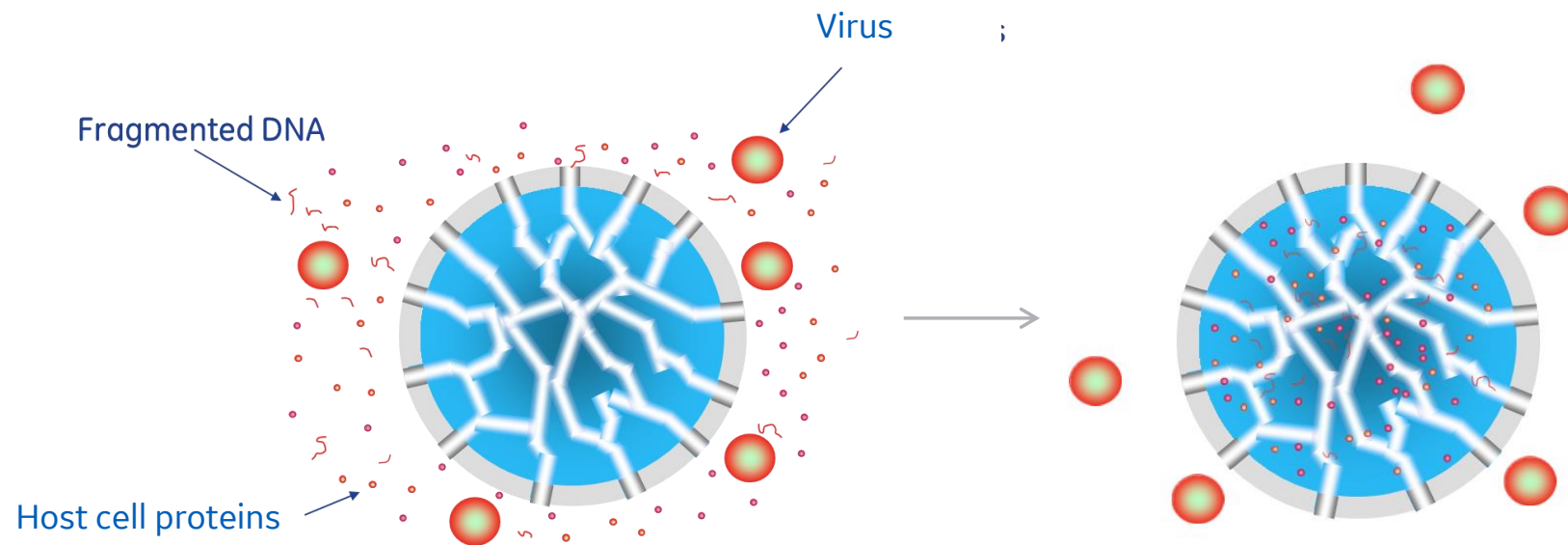
TCID₅₀ = 50% tissue culture infective dose, WCB = working cell bank, WVSS = working viral seed stock



Core bead chromatography: host cell proteins and DNA fragments bind to the core and viruses stay in the void

Modern alternative to SEC

Easily scalable and suitable for single-use chromatography



SEC = size exclusion chromatography



Application examples core beads

Influenza

- Egg-based
- Cell-based

Dengue, Zika, and other
flaviviruses

Lentivirus

Adenovirus

Cytomegalavirus

Respiratory syncytial virus

Poxvirus vectors

Polysaccharide conjugates

VLPs, etc., dependent on size

VLPs = virus-like particles



Cost breakdown of process steps
and cost simulation of process
alternatives

Economical considerations in early development

Litterature search

- Find unit operations for AV purification
- Define suitable running conditions

Process modeling in Biosolve™

- Set up different process alternatives
- Investigate different production scales

Evaluation of results

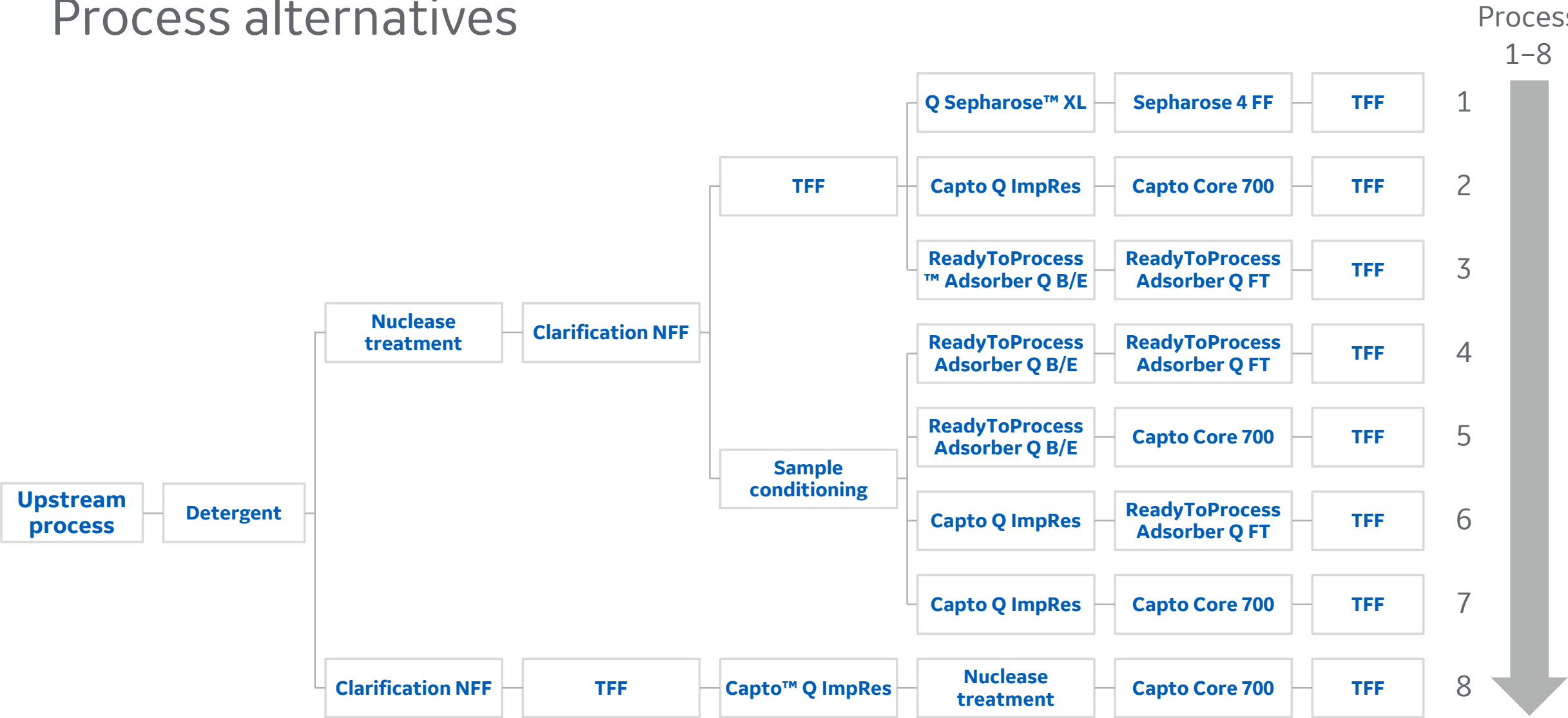
- Identify economically feasible unit operations to evaluate experimentally

Process development

- Start to experimentally evaluate low cost alternatives
- Evaluate only high cost alternatives if needed for required purity



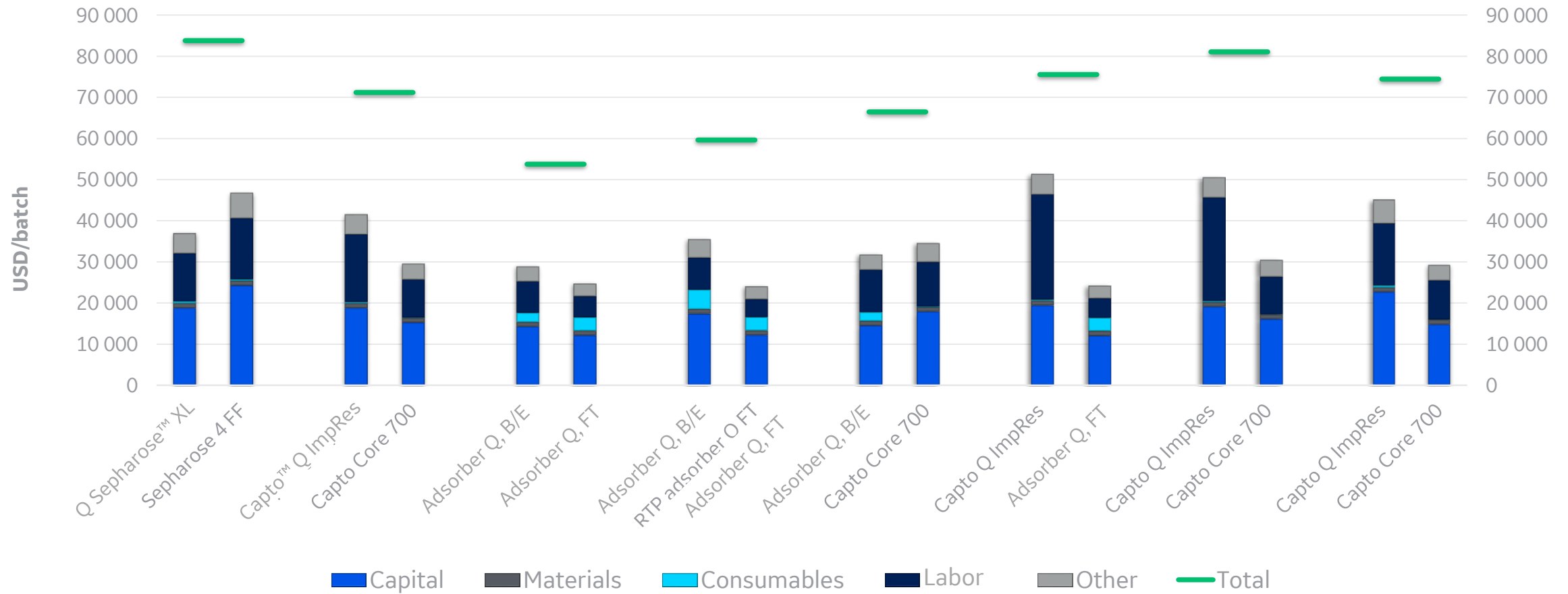
Process alternatives



B/E = bind-elute mode, FT = flow-through mode, NFF = normal flow filtration,

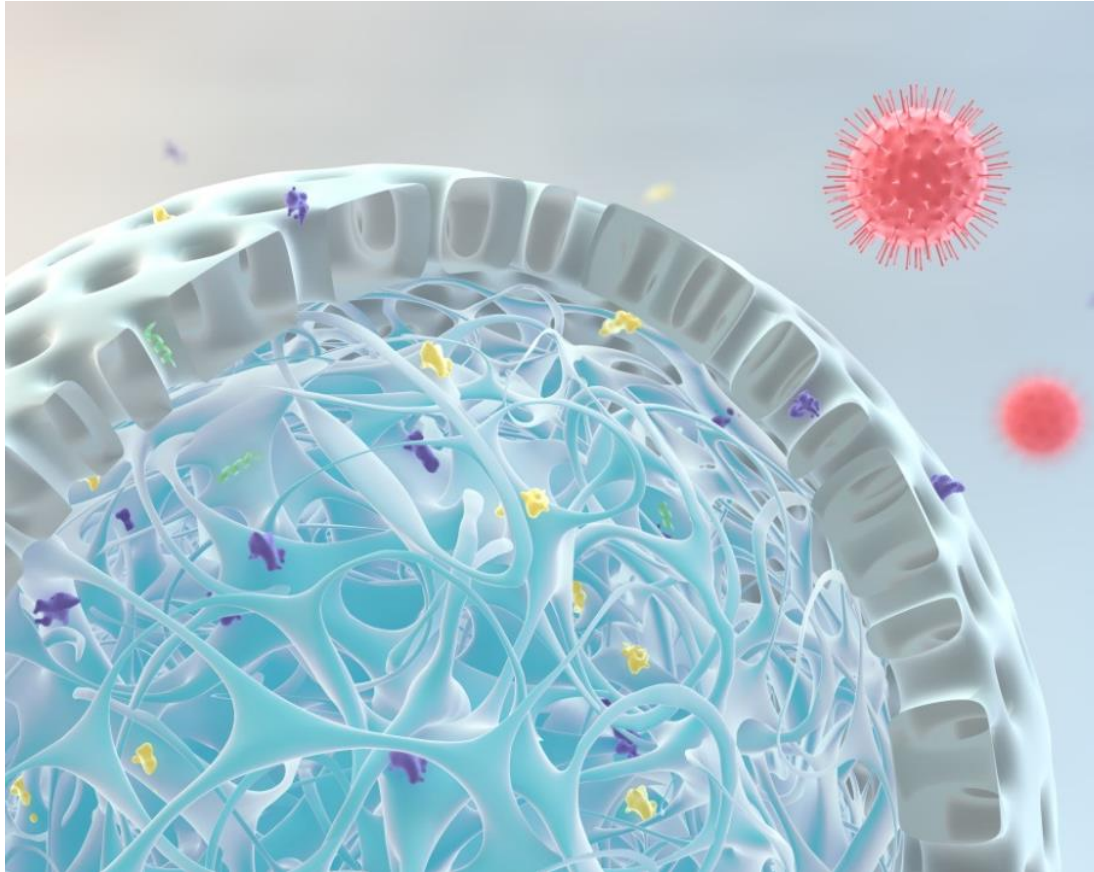


Contributing cost factors



Evaluation of productivity for
modernizing a vaccine process with
a different purification technique

Study objectives

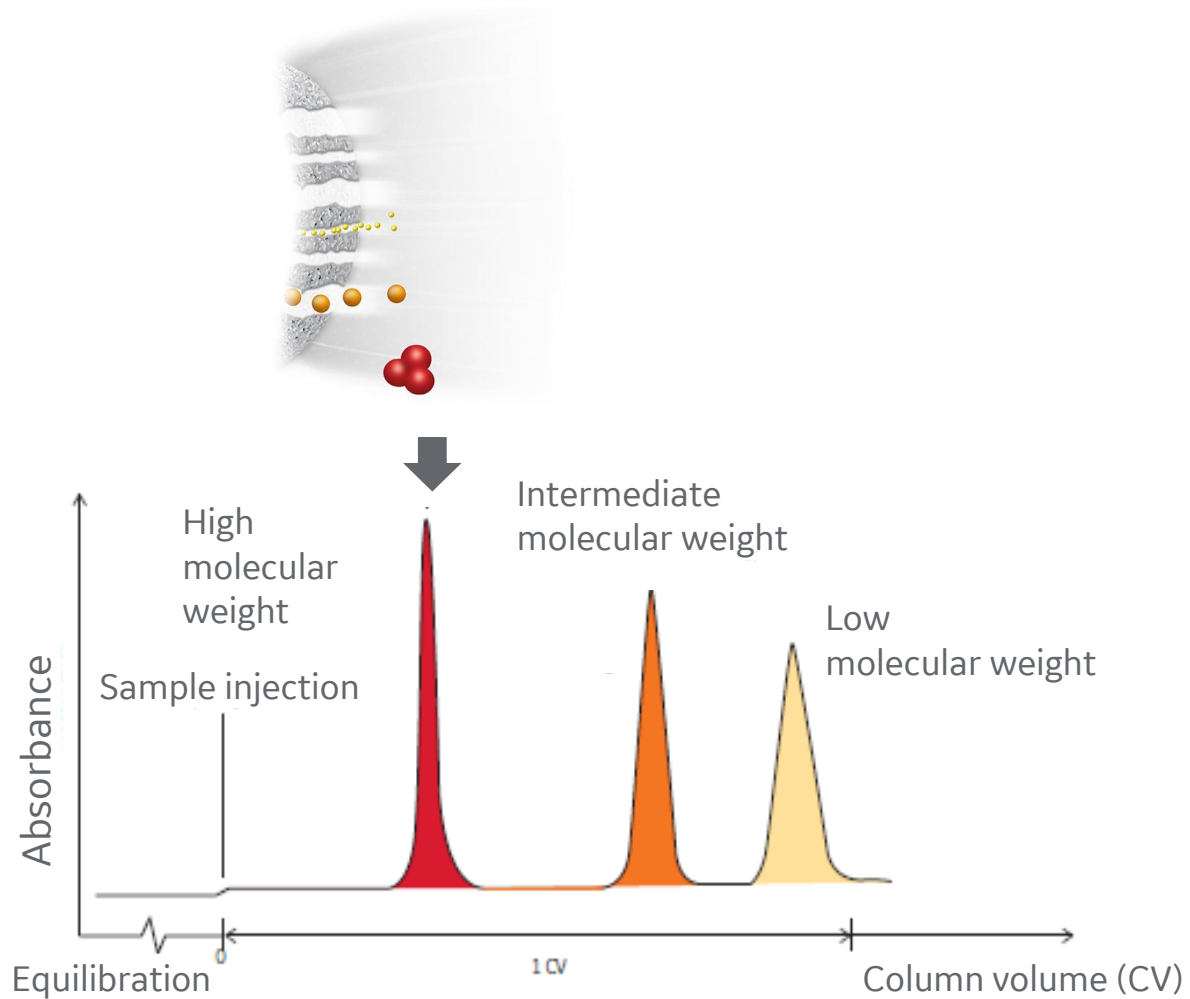


Evaluate the effect on productivity
by replacing a SEC step with a core bead
chromatography step in a vaccine
process at different production scales

SEC = size exclusion chromatography



Principle of SEC



Excluded from pores



Enter a fraction of the pores



Enter all pores

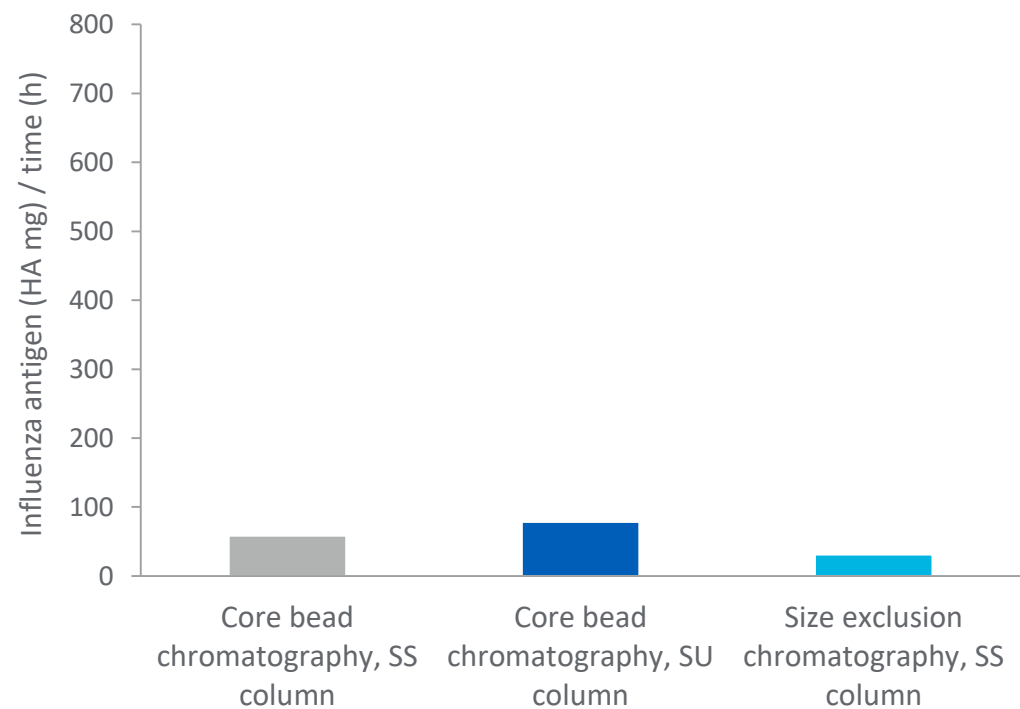


SEC = size exclusion chromatography

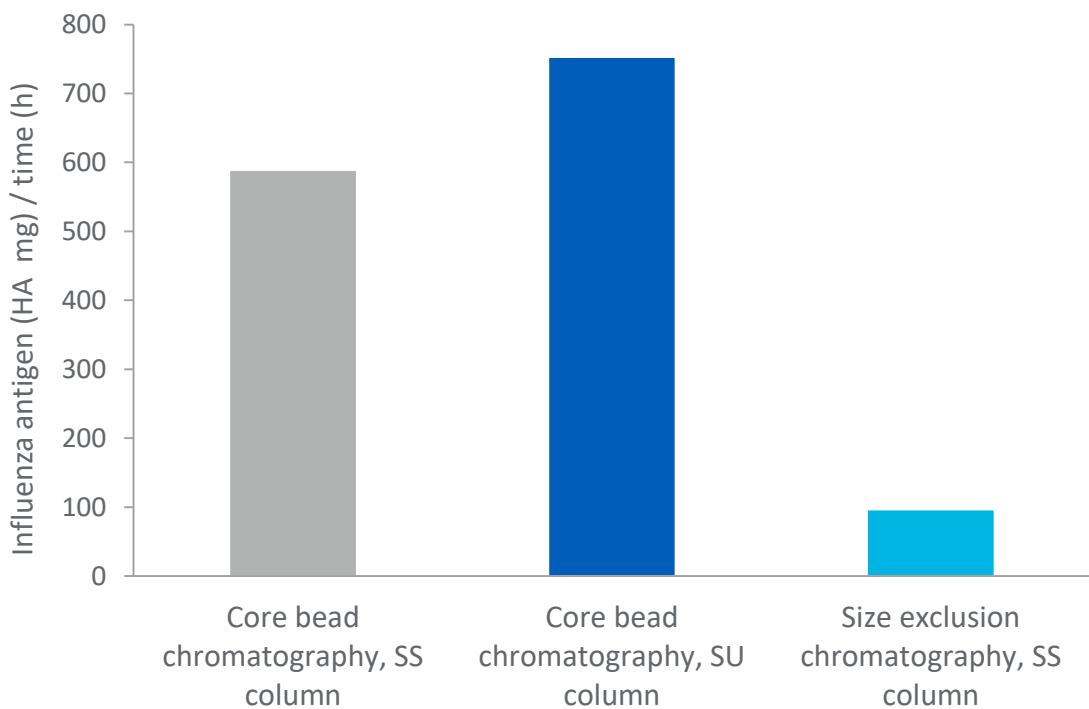


Productivity for SEC and core bead chromatography

200-L scale



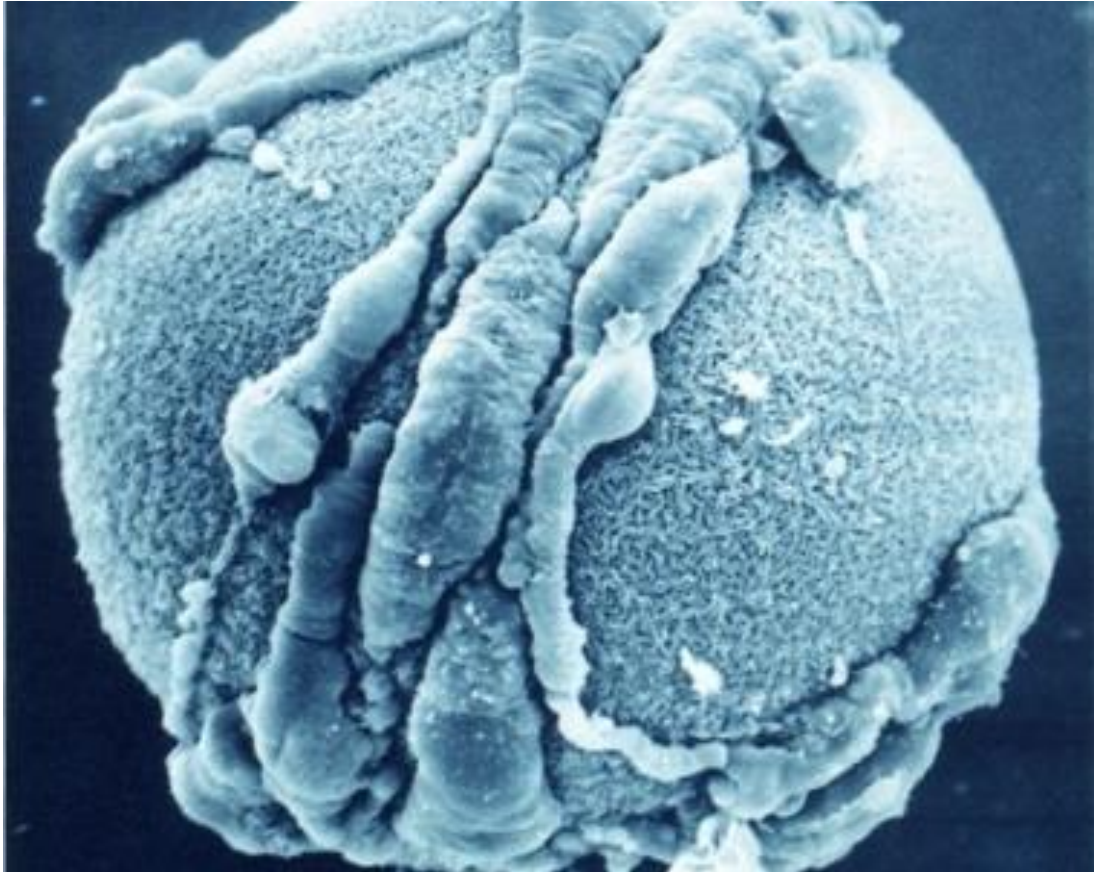
2000-L scale



HA = hemagglutinin, SEC = size exclusion chromatography, SS = stainless steel, SU = single-use



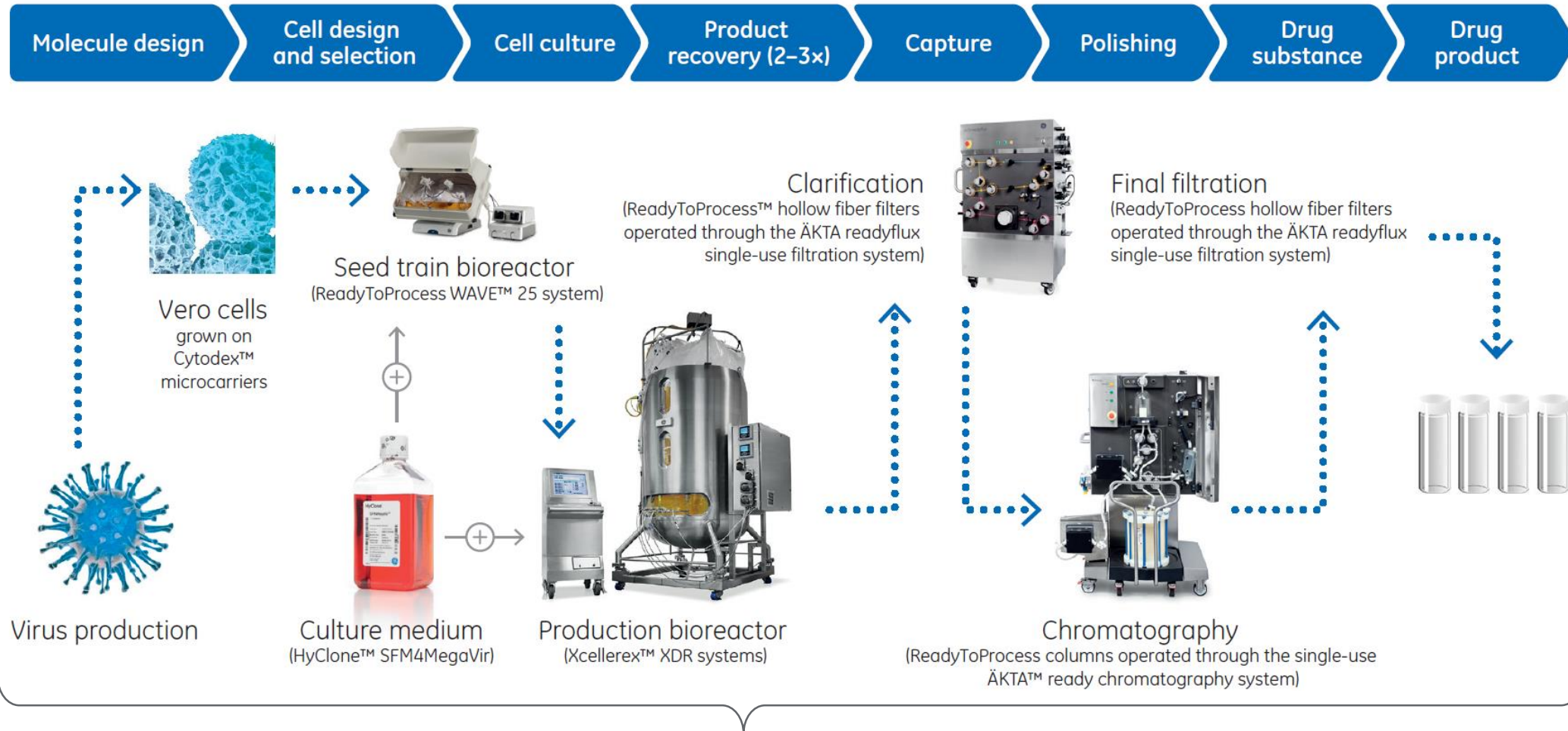
Conclusion



- Paradigm shift for vaccine production—from lab bench process to rational design incorporating process economy calculations early
- A combination of single-use membrane and resin technologies seems to yield beneficial economy overall
- Core bead technology can increase productivity as compared to SEC



End-to-end vaccine manufacturing solutions



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