versatile rechnologies for vaccine Manufacturing

DCVMN 17th Annual General Meeting 24-27 October 2016 Buenos Aires, Argentina

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Developing Countries Vaccine Manufacturers Network

Merck

## The Economist Intelligence Unit Report The Changing Biopharma Risk Equation



- Biopharma top growth strategies
- Top regions / countries of interest for expansion
- Types of products in development and which will be most disruptive
- ROI expectations on growth strategies
- Top of mind risks associated with growth strategies
- Approaches to managing risks
- Confidence in ability to execute



#### Classification: Public The Economist Intelligence Unit Survey Demographics



### LATAM Plans to Expand Beyond its Borders...



Most important strategies for growth over the next five years:

**2%** Expansion into new products

Expansion into new geographical markets

**64%** intend to add production and/or development capacity or grow market share in EMEA & Asia



### while the rest of the world moves to LATAM...



Most important strategies for growth over the next five years:

Expansion into new products

Expansion into new types of therapeutic categories

**52%** intend to add production and/or development capacity or grow market share in LATAM & Middle East/Africa

### Vaccine Development & Manufacturing Strategy in LATAM

**36%** Biotechs are already developing or plan to develop **vaccines** 

## 40%

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LATAM Biotechs plan to partner with other pharmaceutical companies to manufacture new drug and therapy products



## 30%

Global Biotechs plan to invest in production facilities in emerging markets to manufacture new drug and therapy products



Vs.

### LATAM: The Changing Biopharma Risk Equation

Manufacturing and Development of New Drug and Therapy Products

## **Risks will increase** somewhat or significantly over the next five years:

### LATAM

- **52%** controlling costs in development and production
- **44%** maintaining IP protection
- **44%** scaling up and supplying market demand



- **41%** controlling costs in development and production
- **40%** maintaining IP protection
- **36%** maintaining regulatory compliance



### Get a Copy of the Report www.gobeyondbiopharma.com



Are you ready to go beyond today's biopharma?

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- Download the EIU report
- Explore the survey results through interactive infographics
- Stay engaged with updated blog content

## production & purification of virus-Like particle (VLP)-Based vaccine

- Case study and testbed for modern vaccine scale-up and optimization
- > VLPs

Powerful platform for creation of new vaccines

- Processes are currently quite diverse
- Standardized platforms needed to accelerate vaccine development!



### Production & Purification of VLP-Based Vaccine Work carried out in collaboration with iBET



iBET: Instituto de Biologia Experimental e Tecnológica, Oeiras, Portugal

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### Production & Purification of VI P-Based Vaccine Insect Cell Expression System for VLP

VLP produced in Sf9 insect cells co-infected with MLV-GAG and HCV-E1E2 using baculovirus



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#### **ANTIGENS**

E1 and E2 envelope glycoproteins from Hepatitis C virus

#### **STRUCTURE**

Capsid and envelope from retrovirus (murine leukemia



### Hepatitis C

- 170 million people infected, over 350,000 deaths/year
- Causes cirrhosis and liver cancer
- Current therapies only partially effective, costly and poorly tolerated
- No vaccine currently exists



### Production & Purification of VLP-Based Vaccine Process Challenges





### Production & Purification of VLP-Based Vaccine Insect Cell / Baculovirus VLP Production Platform





### Production & Purification of VLP-Based Vaccine Process Step Optimization

## Strategy

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### Production & Purification of VLP-Based Vaccine Establish Scalable, Single-Use Bioreactor Production





## Production & Purification of VLP-Based Vaccine Establish Scalable, Single-Use Bioreactor Production

### **Reasons for increasing Disposables**

#### (% Indicating Attribute is "Very Important" or "Important")



Source: 8th Annual Report and Survey of Biopharmaceutical Manufacturing Capacity and Production, April 2011

Versatile Technologies for Vaccine Production | DCVMN Annual Meeting | Buenos Aires, Argentina | 24-27 Oct 2017 | Damon Asher





## Production & Purification of VLP-Based Vaccine Establish Scalable, Single-Use Bioreactor Production



Ratio	3 L	50 L	200 L	1000 L	2000 L
Working Volume : Total Volume	0.8	0.8	0.8	0.8	0.8
Impeller Diameter : Vessel Diameter	0.6	0.3	0.3	0.3	0.3
Vessel Height : Vessel Diameter	1.8:1	2.0:1	2.0:1	2.0:1	2.0:1
Liquid Height : Vessel Diameter	1.4:1	1.7:1	1.6:1	1.6:1	1.6:1
Internal Baffle	No	Single (Pa	ddle-Type)	Single	(X-Type)
Dual sparger (open pipe/ microsparger)	YES	YES	YES	YES	YES
Min – Max Working Volume (L)	1 - 2.4	10 - 50	40 - 200	200 - 1000	400 - 2000



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## Transition from Glass to Scalable Single-Use Bioreactor Initial challenges







### CR Conditions:

27 °C,  $pO_2 = 30\%$ Agitation rate: 70–200 rpm Impeller: 1 marine Aeration rate: 0.01 vvm Working volume = 2 L

### Sf9 cells in CR







### STR Conditions:

27 °C,  $pO_2 = 30\%$ Agitation = 70-250 rpm Impeller: 2 six-blade rushton Aeration rate: 0.01 vvm Working volume = 2 L



Sf9 cells in STR

Observation of cell aggregates for unoptimized CR culture Long lag-phase observed for CR culture



# Transition from Glass to Scalable Single-Use Bioreactor **Process Optimization**

Table 1. Shear stressin the two bioreactors

	Agitation Rate	Shear stress (N/m <sup>2</sup> )	
	(rpm)	STR	CR
→	70	0.14	0.05
	90	0.20	0.07
	110	0.27	0.09
	130	0.34	0.12
	150	0.43	0.15
	170	0.51	0.18



New CR Conditions: Agitation rate: 150 rpm

	μ <sub>max</sub> (h <sup>-1</sup> )	T <sub>d</sub> (h)
CR	0.20	35
STR control	0.20	34

CR match STR hydrodynamic parameters for agitation rates above 150 rpm

 Enhanced cell growth in CR
 CR and STR show similar growth profiles . . . . .

# Transition from Glass to Scalable Single-Use Bioreactor **Optimized Conditions**



Mobius<sup>®</sup> 3L Single-Use Bioreactor

### **Optimized conditions**

- Increased agitation rate
- Increased cell density of inoculation
- Replaced micro sparger with an open-pipe sparger



STR=Glass stirred tank



. . . .

## Transition from Glass to Scalable Single-Use Bioreactor Sf9 Growth Profiles





Images of Sf9 cell during culture in CR or STR



STR

CR

No formation of cell aggregates

Metabolite concentration in culture supernatants



Comparable metabolite profiles in STR and CR

## Transition from Glass to Scalable Single-Use Bioreactor **VLP-HCV Productivity in the Two Bioreactors**

. . . . STR=Glass stirred tank CR=Single-use bioreactor



WB analysis of VLP-HCV production kinetics



Similar GAG and HCV-E1E2 expression kinetics

Gag-MLV titer (P30) in the bioreactor harvested bulk



Equivalent productivity in STR and CR culture bulks

Baculovirus replication kinetics



Comparable baculovirus replication kinetics



## **Production & Purification of VLP-Based Vaccine** Single-Use Bioreactor, Scale-Up to 50L





#### **VLP-HCV** Production

Growth kinetics of Sf9 cells in the Mobius<sup>®</sup> 50 L and 3 L bioreactors. The black arrow indicates the infection of the bioreactors and control shake-flask.



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### Production & Purification of VLP-Based Vaccine Insect Cell / Baculovirus VLP Production Platform





## Production & Purification of VLP-Based Vaccine Clarification Optimization



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### **Disposable capsule filters**

Eliminates the time and expense associated with assembling, cleaning, and validating stainless steel housings.

Replacement for centrifugation

Polygard<sup>®</sup> CN, nominal pore sizes of **10, 5, 0.6 and 0.3 μm** 

Pleated, all-polypropylene depth filters Filter area: 17 cm<sup>2</sup>; Inlet flux: 988 LMH









# Production & Purification of VLP-Based Vaccine Clarification Optimization





### Clarification

- Filter-only clarification train can be used without compromising recovery yield of VLPs.
- Filter cascade composed of a Polygard<sup>®</sup> CN 5 µm filter followed by a 0.3 µm depth filter showed the highest recovery of HCV-VLP, improving on centrifugation/2° depth filtration
- Moderate DNA removal with depth filtration was seen

### Unlike centrifugation (CFG), depth filtration resulted in ~70% DNA clearance

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### Production & Purification of VLP-Based Vaccine Insect Cell / Baculovirus VLP Production Platform





## Optimization of TFF for Concentration and Purification Membrane Choices



	Cellulosic Membrane (e.g. Ultracel® membrane)	PES Membrane (e.g. Biomax® membrane)	
Membrane	Composite regenerated cellulose (void free structure)	Hydrophilic modified polyethersulfone (void free structure)	
Typical Available NMWL	3, 5, 10, 30, 100, 300, 1000 kD (new 1 kD in PD)	5, 8, 10, 30, 50, 100, 300, 500, 1000 kD	
Relative protein binding	Ultra low (~0.1 g/m2) Use with any protein concentration (good with dilute)	Low –medium (~0.2 g/m2) Use with solutions > 0.1 mg/mL	
pH Stability	2-13	1-14	
Comments	<ul> <li>Organic solvent resistance</li> <li>High yield</li> <li>Very hydrophilic – use with solutions containing hydrophobic components (i.e. antifoams or detergents)</li> </ul>	<ul> <li>Resistant to rigorous cleaning regimes (strong bases and acids)</li> <li>High flux</li> </ul>	







## **Optimization of TFF for Concentration and Purification Purification Results**





#### **UF/DF**

Pellicon<sup>®</sup> cassette with **300** kD regenerated cellulose membrane offered the best combination of recovery and purification

4-5X concentration achieved

Better removal of baculovirus, DNA, and host-cell protein!

Both membranes were fully retentive of the VLP



### Production & Purification of VLP-Based Vaccine Insect Cell / Baculovirus VLP Production Platform



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## Purification strategy **Resin Selection**







BVLRV

## Anion Exchange Chromatography (AEX) for VLP Purification **Resin Selection**







Tentacle morphology increases surface area and virus binding capacity



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## Anion Exchange Chromatography (AEX) for VLP Purification Separation of VLP from Baculovirus

Inputs: [NaCI] (100/200/300 mM) and flow rate (100/200/400 cm/hr) Responses: % VLP recovery and Baculovirus LRV



Fractogel<sup>®</sup> TMAE Anion Exchange Resin



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## VLP-Based Vaccine Process Optimal Purification Balance

- Successfully purified VLPs using Fractogel<sup>®</sup> TMAE commercial resins
- Yield of >60% with ~2 LRV baculovirus can be achieved with a flow-through/wash purification strategy
- Options to increase recovery or purification depending on product value by varying process conditions









### **VLP-Based Vaccine Process**

### Pre-Packed Chromatography Columns Add Flexibility



- Elimination of labor-intensive packing
- No capital investment
- No hardware to clean
- Fast setup and qualification



Chromabolt<sup>®</sup> Prepacked Chromatography Columns 10cm, 20cm, 32cm



Column types (prepacked vs. manually packed)

## Time Savings



### **Summary**



Successfully used Mobius<sup>®</sup> 3L **disposable bioreactor** for production of VLP-based vaccine in insect cell culture system



Optimized downstream processing using Polygard<sup>®</sup> CN 5.0→0.3 µm depth filters followed by UF/DF using Pellicon<sup>®</sup> cassette with Ultracel<sup>®</sup> 300 kD membrane



Purified VLP by using Fractogel<sup>®</sup> commercial AEX resins



Integrated all the above components to create a fully scalable process that meets recovery and impurity clearance requirements.



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Thank you

