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Biosafety Considerations for Cell Based Viral Vaccines

Margaret Temple SGS Vitrology Site Director Glasgow - UK





Presentation Outline:

- Sources of adventitious contamination in biological material
- Safety concerns relating to virus based therapeutics
- Why validated test methods are of interest to the Regulatory Authorities
- Cell bank and virus vaccine characterisation tests and methods



An MHRA accredited contract testing company providing GMP and /or GLP drug development services to the global biotechnology industry, to comply with the current requirements of the international regulatory authorities.

Testing data supports products in clinical evaluation and marketed under commercial licence



APPROVAL OF A CELL BASED VACCINE

For Marketing Approval

- Safety
- Efficacy
- Consistency

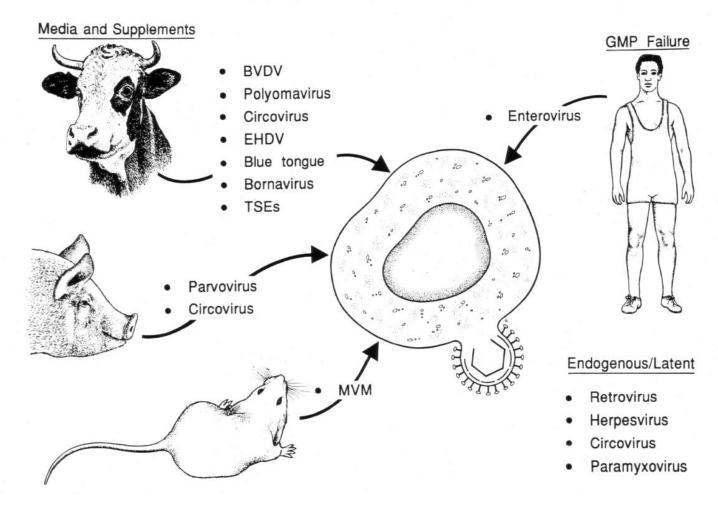
Investigational Medicinal Product (IMP) for clinical trials

- Safety
- Efficacy Toxicological
 Consistency Biological



ADVENTITIOUS AGENTS: SGS ADVENTITIOUS AGENTS: CELL CULTURE DERIVED PRODUCTS

SOURCE OF ADVENTITIOUS AGENTS





Biological Safety

Microbiological

Identity

Genetic Stability

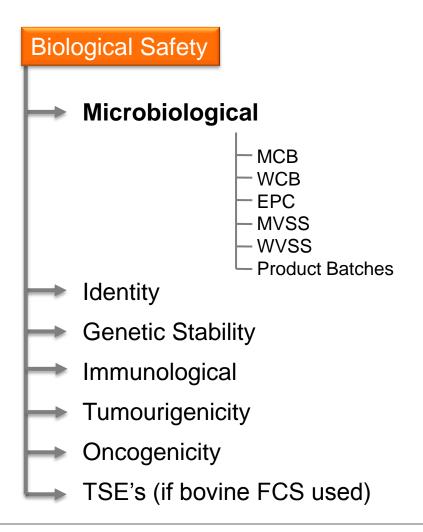
Immunological

Tumourigenicity

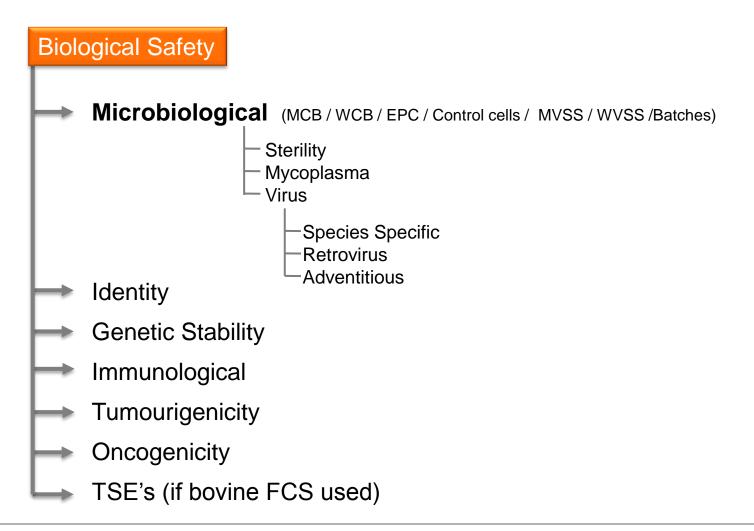
Oncogenicity

TSE's (history & if bovine FCS used)

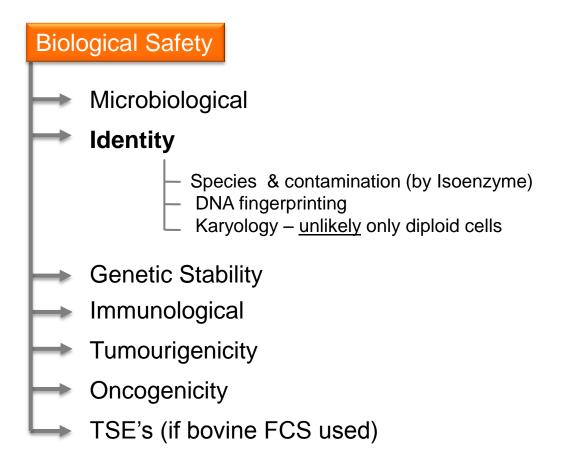




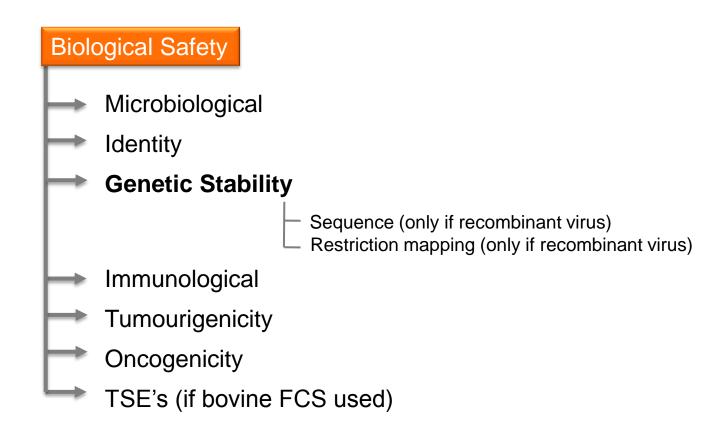




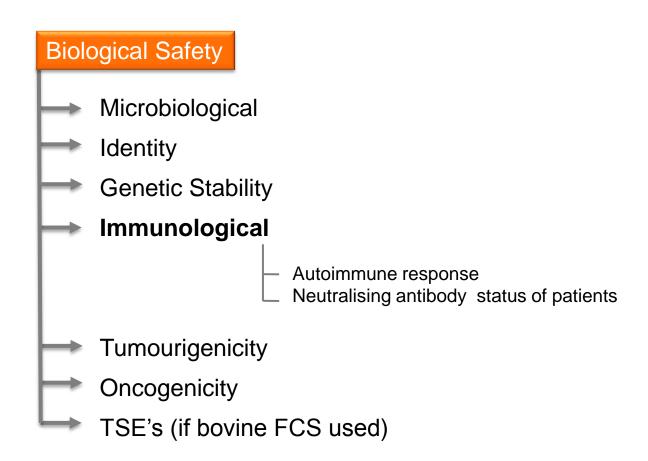




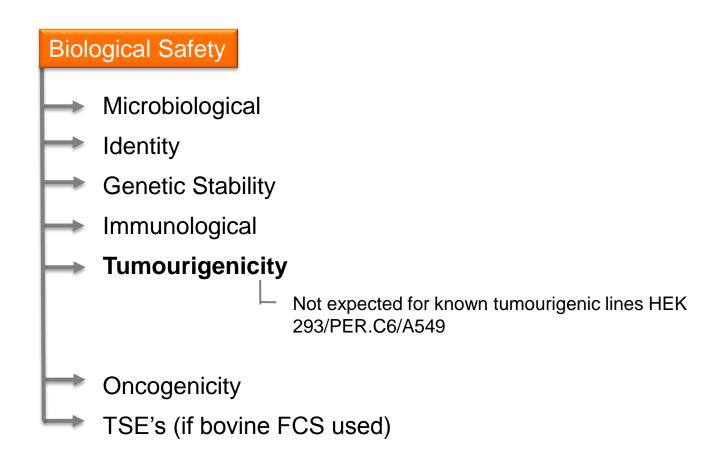














Biological Safety Microbiological Identity **Genetic Stability** Immunological **Tumourigenicity Oncogenicity** Define acceptable levels of host cell DNA & proteins **HCD Size** TSE's (if bovine FCS used)



Biological Safety

- Microbiological
- Identity
- Genetic Stability
- → Immunological
- Tumourigenicity
- Oncogenicity
 - TSE's (history)

Adapt to serum free cultivation where possible
Sourcing of FCS, if essential



Testing of

- Cell banks and raw materials
- Cell derived harvests and final product
- Viral seeds and harvests
- Animal tissue and cell derived products

For manufacture of

- Monoclonal antibodies
- Recombinant protein
- Viral vaccine
- Cell therapies
- Gene therapies



Assay Systems Require:

Protocol (Development, Validation, Study)

Method SOP(s) (Development, Validation, Study)

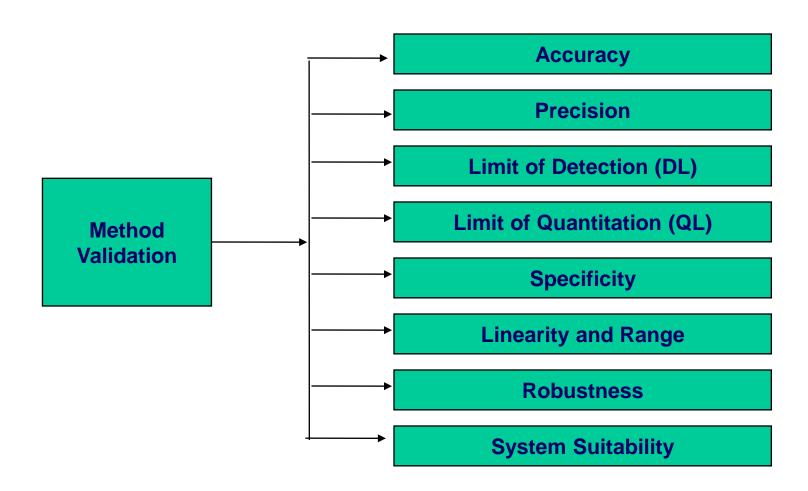
Data Record

■ Final Report or (Development, Validation, Study)

Summary Report/C of A (Development, Validation, Study)



Validation of methods to meet ICH guidelines Q2(R1)





Importance of Assay Validation Documents

- □ FDA Guidance for Industry: Analytical Procedures and Methods Validation; Chemistry, Manufacturing and Controls Documentation
 - Validated procedures should be used for in-process, release, acceptance, and stability testing
- EMEA Guideline on virus safety evaluation of biotechnological IMP
 - For Phase I/II clinical trials, the suitability of the analytical methods used for viral testing should be demonstrated.
 - Results of values found for ICH validation should be available (eg. specificity, linearity, range, accuracy, precision, quantification and detection limit)
 - It is not necessary to provide a full validation report.
 - Phar. Eur. viral tests are not normally required to be re-validated.
 - Note: For Phase III studies a full validation report should be available



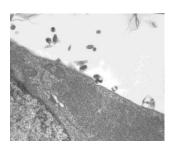
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Methods used for the Characterization



In vivo assays

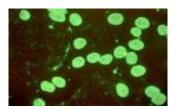


Transmission Electron Microscopy

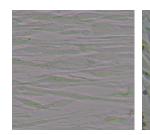
Sterility



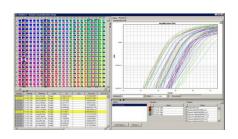
Cell Bank & Viral Vaccine Characterisation



Mycoplasma



In vitro adventitious agent assays



Q-PCR for specific viruses, RT activity



Cell Bank Characterisation (eg. Human) FDA Vaccine Guidance 2010 Phar. Eur. 5.2.3 & ICH Q5A

Concern	Assay	MCB	WCB	CAL	CC
Identity	DNA fingerprint	+	+	+	
	Isoenzyme	+	+	+	
Microbial	Sterility	+	+	+	<u>+</u>
Microbial	Mycoplasma	+	+	<u>#</u>	<u>+</u>
	Mycobacteria	(+)	(+)	(+)	(+)
Adventitious viruses	In vitro	+	(+)	+	(+)
	In vivo (including GPigs)	+		+	(+)
Tumourigenicity Karyology	Nude mice G-banding	+	+	+ +	

^{1 Note} If production culture conditions are changed the status of the unprocessed bulk harvest should be confirmed by an *In vivo* assay.

Requirement for Phar. Eur.

Master cell bank (MCB)
Working cell bank (WCB)
Cells at limit (CAL) (EOP)
Bulk Harvest (BH)
Control cells (CC)
Alternative stage (+)



Testing of Human Cell Bank continued....

Assay	МСВ	WCB	CAL	CC
TEM PERT ¹ HEK 293 Co-cult	+ + (+)		+ + (+)	(+) (+)
In vitro	(+)	(+)	(+)	(+)
PCR	(+)	(+)	(+)	(+)
PCR	(+)	(+)	(+)	
iined	Master cell bank Working cell bank Cells at limit Bulk Harvest Control cells Alternative stage		(MCB) (WCB) (CAL) (EG) (BH) (CC)	OP)
	TEM PERT 1HEK 293 Co-cult In vitro PCR PCR	TEM + PERT + HEK 293 Co-cult (+) In vitro (+) PCR (+) PCR (+) Master Workin Cells a Bulk H Contro	TEM + PERT + 1HEK 293 Co-cult (+) In vitro (+) (+) PCR (+) (+) PCR (+) (+) Master cell bank Working cell bank Cells at limit Bulk Harvest	TEM + + + + + + + + + + + + + + + + + + +



Bovine viruses of concern that may go undetected by 9CFR

Table 3

Viruses with human and bovine host range that are not predicted to be detected by 9CFR test.

Viruses to Consider for Additional Evaluation	
Anelloviridae (proposed family) Torque teno virus TTV ^a	Parvoviridae
Bornaviridae Borna disease virus BDV [134]	Bovine adeno-associated virus BAAV ^a Bovine hokovirus BHoV ^a
Bunyaviridae Cache valley virus CVV [2] Puumala virus [135] Caliciviridae	Picornaviridae Bovine enterovirus BEV-1 ^a , BEV-2 ^a Seneca valley virus SVV [138] Polyomaviridae Bovine polyomavirus BPyV [71, 139]
Norovirus [formerly Norwalk agent] [136] Circoviridae Bovine circovirus bovCV ² Coronaviridae Bovine torovirus BtoV ²	Poxviridae Aracatuba virus ^a Cantagalo virus ^a Cowpox virus [140] Pseudocowpox virus PCPV [141, 142]
Flaviviridae Louping ill virus ^a Saint Louis encephalitis virus SLEV ^a Wesselsbron virus [137]	Reoviridae Banna virus BAV [82] Epizootic haemorrhagic disease virus EHDV [143] Rotavirus [144]
Hepeviridae Hepatitis E virus HEV ^a	Retroviridae Bovine foamy virus BFV [145]
Papillomaviridae Bovine papilloma virus BPV ^a	Bovine leukemia virus BLV ^a Togaviridae Ross River virus RRV [99]

^a No information found to allow assessment of ability to be detected by 9CFR, hence considered as potential risk.



Porcine viruses of concern that may go undetected by 9CFR

Table 4

Viruses with human and porcine host range that are not predicted to be detected by 9CFR test.

Viruses to Consider for Additional Evaluation	
Anelloviridae (proposed family) Torque teno virus TTV ²	Paramyxoviridae Menangle virus MENV [119]
Caliciviridae Norovirus [formerly Norwalk agent] [136] Sapovirus ^a	Parvoviridae Porcine hokovirus PHoV [62]
Circoviridae Porcine circovirus PCV-1 & PCV-2 [146](B.Potts per	Picornaviridae Porcine enterovirus PEV-9 PEV-10 [129] Seneca valley virus SVV [138]
Flaviviridae Louping ill virus[133] Powassan virus [147] Wesselsbron virus [137]	Reoviridae Banna virus BAV [82] Rota virus[144] Retroviridae
Hepeviridae Hepatitis E virus HEV ^a	Porcine endogenous retrovirus PERV ^a
Herpesviridae	Togaviridae Ross River virus RRV [99]
Porcine cytomegalovirus PCMV ^a	

Note: These are not exhaustive lists, and other viruses related to those listed above may also be of concern.

a No information found to allow assessment of ability to be detected by 9CFR, hence considered as potential risk.



PCR available at Vitrology => already being used for large vaccine clients



VITROLOGY

Origin, Source and History of Cell Line

- Source
 - species, strain, tissue or organ of origin
 - state of health, medical history if human donor
- Cultivation history
 - procedures used in culturing cells
 - procedures used to establish cell line
 - description of genetic manipulation or selection
 - identity, testing for endogenous and adventitious agents
 - constituents of culture medium
- Procedures used
 - cell fusion
 - transfection
 - Selection & colony isolation
 - cloning
 - gene amplification
 - adaptation to culture conditions and media



Methods for cell bank identity testing

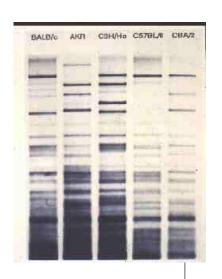
- Microscopic observation
 - Cell monolayers can show "general" characteristic morphology
 - Does not provide definitive results on the identity
- Isoenzyme analysis
 - Provides identification of the species of origin
 - Technique is referenced in the regulatory guidelines. AuthetiKit™ system (Innovative Chemisty Inc.)
 - Defines a set of enzymes with a characteristic "mobility pattern" for each species

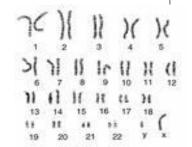
Potential Cell Mixture	Distinguishing Enzymes	
NSO and CHO	PEP B	
Human and Vero	AST or MDH	
CHO and BHK	MDH	
Human and CHO	LDH	
Human and NS0	LDH	
Insect and Mammal	NP or G6PD	



Methods for cell bank identity testing

- DNA fingerprinting or Random Amplified Polymorphic DNA (RAPD)
- Vaccine cell substrate guideline requirement (Ph. Eur. 5.2.3)
- The technique can be used to distinguish between individuals of the same species
- When applied to cell lines, the method is sensitive enough to identify a specific cell line
- For example, can distinguish between human cell lines HEK 293 and A549.
- Karyotypic analysis
 - In general is required when product may contain live cells (eg vaccine)
 - Modal chromosome number and characteristic chromosome markers
 - Useful for the evaluation of novel cell lines.
 - Diploid cells should be shown to be diploid
 - Diploid cells have two homologous copies of each chromosome
 - MRC-5, WI-38, FRhL-2 are well characterised diploid cell lines and do not require further qualification
 - Karyology may not be required for highly purified products





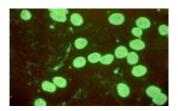


Sterility

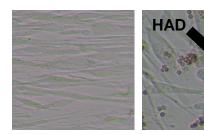
In vivo assays

Transmission Electron Microscopy

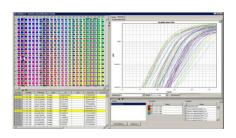
Purity of Cell Banks



Mycoplasma



In vitro adventitious agent assays



Q-PCR for specific viruses, RT activity



Sterility of cell banks (USP/Phar.Eur/JP)

- Cell and virus banks manufactured under cGMP conditions require sterility testing
- Methods are outlined in the US and EU Pharmacopeias and the 21 CFR 610.12.
- For cell banks test 1% or minimum of 2 containers for MCB and WCB (ICH Q5D recommendation)
- Direct broth inoculation or membrane filtration
- Aerobic and anaerobic incubations, 14 days
- Observation for turbidity (microbial growth)
- Qualification of Test Item by spiking
- Alternative Rapid Microbiological Methods (RMM) if validated may be used on cell therapy/gene therapy products.



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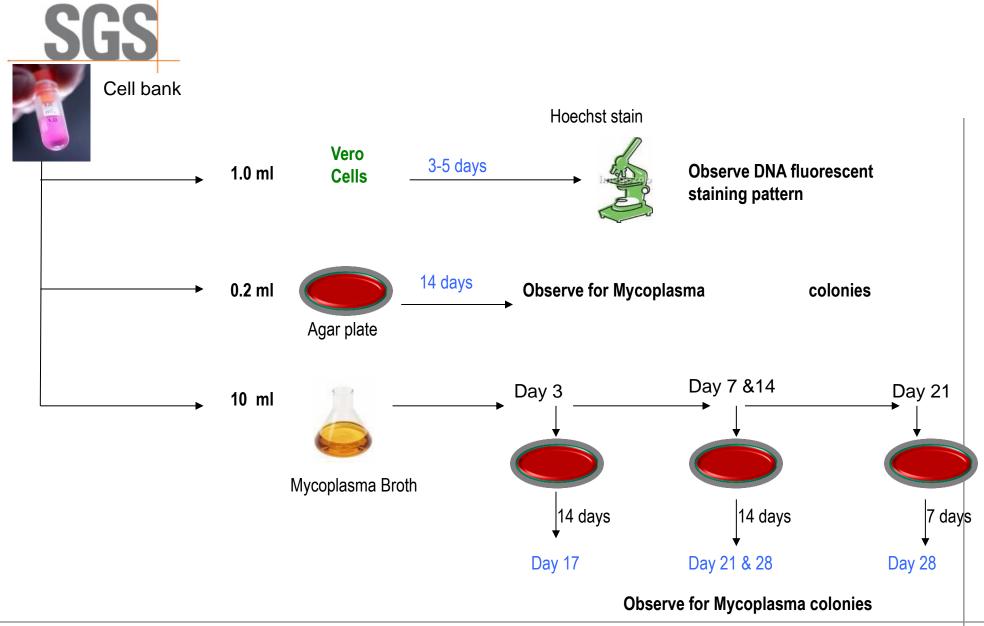
Mycoplasma contamination

- Small extracellular parasite- prokaryote
- Infect a wide variety of hosts (mammals, birds, reptiles, fish, insects, plane
- 5 species cause 95% of the contaminations of cell cultures
- Mycoplasma presence decreases quality or quantity of product
- Mycoplasma alters:
 - Cell function, growth & metabolism
 - Virus propagation and yield
 - Can cause chromosomal aberrations
- Testing according to guidelines
 - USP 63 and Eur Phar 2.6.7 Mycoplasma
 - Important to check for assay inhibitory substances



Mycoplasma colonies on agar Fried egg apearance

Mycoplasma Detection - Indicator cell, Broth & Agar

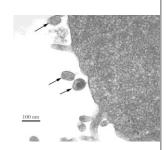


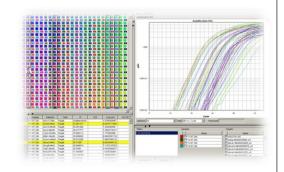


Purity- Adventitious Agent Assays

- Requirement to utilize a number of different assays to detect and wide range of possible contaminants
- Broad specificity assays
 - in vitro and in vivo virus assays,
 - transmission electron microscopy (TEM)
- Assays to detect contaminants associated with specific species
 - rodent, human, bovine, porcine viruses
- Assays to detect retroviruses
 - infectivity assays
 - molecular biology assays (PCR)
 - biochemical assays (reverse transcriptase)
 - morphological assays (electron microscopy)

the "unknown"

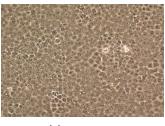


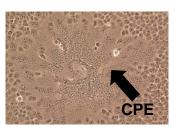




Broad specificity virus detection methods in cells

- Cytopathic Effect (CPE)
 - CPE is a degenerative change in cells
 - CPE may lead to plaque formation
 - not all viruses produce CPE





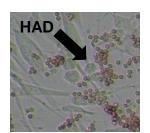
Vero

Vero + Measles virus

- Haemadsorption (HAD)
 - HAD is a visual method for detecting viruses based on adherence of red blood cells (RBC) to infected cells.
 - Mix of bloods- Guinea pig, Human O, and Chick

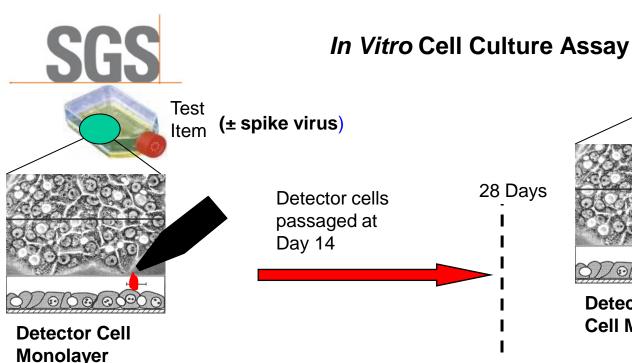
MRC-5

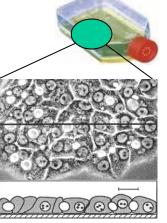




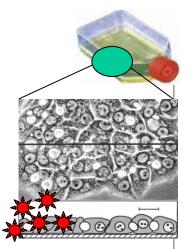
MRC-5 + Parainfluenza virus

CPE and HAD are critical methods within the *In vitro* virus detection assay





Detector Cell Monolayer



Infected Detector Cell Monolayer

Common Detector Cells

- MRC-5 (Human)
- Vero (Simian)
- Same Species and Tissue as Cell Bank
 SP2/0, BHK, CHO, NS0 etc.
- Include Rabbit kidney cells:- for simian cell substrates used in vaccine production (detects Herpes B virus)
- In vitro assays to detect bovine and porcine viruses described in:
 - ■Title 9 Code Federal Regulations 113.47/51/53

End Point Detection Methods

- Cytopathic Effect CPE
- Haemadsorption HAD
- Others
 - Immunofluorescence
 - RTase / PERT assay
 - Electron Microscopy
 - PCR

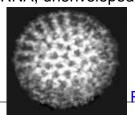


VITROLOGY



- detects intracellular virus particles
- rapid morphological detection
- used to indentify "unknown" contaminants
- a test with broad specificity
- may detect viruses that are not detected by *In vitro* or *In vivo* assays
- used to measure retrovirus load in bulk harvest prior to downstream purification

Double stranded RNA, unenveloped



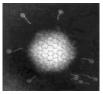
Rotavirus

Double stranded DNA, enveloped



Poxvirus

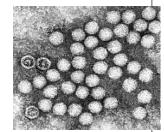
Double stranded DNA, unenveloped



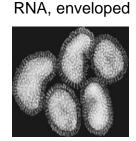
Adenovirus

Single stranded

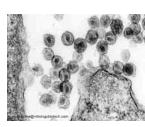
Single stranded RNA, unenveloped



Enterovirus



myxoviruses



HIV



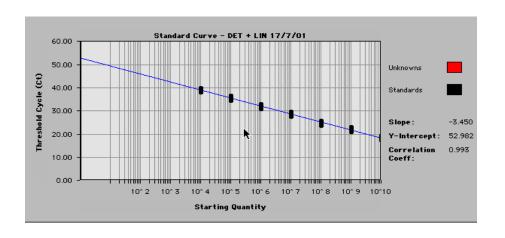
Rabies

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The Product Enhanced Reverse Transcriptase (PERT) Assay – detection and quantitation of Retroviral RT

Brome Mosiac virus (BMV) RNA template is converted to cDNA by retroviral RT enzyme if present in a "test article"



Q-PERT Assay is Quantitative

F-PERT Assay is Qualitative

Measurement of RT Enzyme Activity

F-PERT used for testing vaccines and for end point in retroviral infectivity assays

QPERT used in virus vaccine bulk harvest retroviral load monitoring (CEF).



RT Assays (F-PERT) Retrovirus RTase detection by PCR

- Regulatory Guidance
 - Letters to Industry, CBER FDA, 1998 (Vaccines)
 - ICH Q5A (cell therapy, recombinant proteins etc)
 - FDA Guidance Vaccines (2010)
 - Phar. Eur. 5.2.3 Cell Substrates (GT vector and vaccines)
 - WHO TRS 878 (cell substrates for biologicals)
- Extremely sensitive assay for detection of a wide range of retroviral RT activities in cell substrate supernatant, vaccines and gene therapy vectors. Sensitivity:
 <1000 retrovirus particles (500 particles per ml)
- □ FDA 2010 Vaccine guidance document states that the assay limit of detection should be comparable to published literature (specifically Lovatt *et al.*, 1999)
- Assay can sometimes be false positive from normal background cellular DNA polymerase activity → Retroviral infectivity testing (HEK 293 detector) are then used for confirmation
- Assay can also be performed using a Quantitative PERT (Q-PERT)

SGS

Assays used to detect Retroviruses

Infectivity Assays

- XC plaque: for the detection of ecotropic murine retrovirus
- S+L- focus forming: for the detection of xenotropic murine retrovirus
- Mus dunni: detects all types of murine leukaemia viruses
- Co-cultivation: test sample co-cultured with human or murine cell lines followed by specific end point analysis

RTase

- PCR-based RT assays (PBRT or PERT)
- Detects reverse transcriptase activity associated with virus particles
- Sensitivity: 1000-10,000 retrovirus particles

Quantitative PCR

- Specific nucleic acid amplification of viral genomes
- Detects infectious and non-infectious virus
- Specific, but only detects the specified virus

TEM

- Sensitivity: -10⁵ particles per ml (bulk harvest)
- Specific for identification of retrovirus type



APPROVAL OF A CELL BASED VACCINE

VITROLOGY

For Marketing Approval

- Safety
- Efficacy
- Consistency

Investigational Medicinal Product (IMP) for clinical trials

- Safety
- Efficacy
- Consistency

In summary each vaccine should have a case by case, regulated biosafety testing strategy and manufacturing controls at each stages from development

VITROLOGY IS NOW PART OF SGS, THE WORLD'S LEADING INSPECTION, VERIFICATION, TESTING AND CERTIFICATION COMPANY.

Biosafety Considerations for Cell Based Viral Vaccines

Appendix

Margaret Temple SGS Vitrology Site Director Glasgow - UK





SGS LIFE SCIENCE SERVICES VITROLOGY OVERVIEW



- Over 35 years experience
- 1,400 full time staff
- 28 facilities in 15 countries
- Leader with unique international analytical laboratory network
 - across America, Europe, Asia
 - with Centers of Excellence
- Global drug development partner from Molecule to Market
 - Wide-range of laboratory infrastructure, size and diverse testing capabilities matching Biopharmaceutical and Small Molecules needs
- Strong commitment to clinical and laboratory Quality and Operational Excellence
 - Harmonized QMS and Validation & Transfer methods, LIMS



VITROLOGY HISTORY

- Established in April 2007
 - 4 founding members now part of SGS Life Science Services
 all former senior employees with Q-One Biotech/BioReliance
 - Dedicated Partnerships with Moredun Foundation (animal),
 Beckman Coulter Genomics (MPS) and Takara Bio Inc. (Japan)
- 2010-12 successful re-inspections by MHRA for GLP and GMP compliance.
 - GLP/GMP compliant since 2008
- Acquired by SGS in May 2012
 - "Centre of Excellence for cell bank characterisation & virus testing"



VITROLOGY CORE SERVICES

- GMP Cell Bank & Virus Seeds Characterisation (Vaccines & Gene Therapies)
 - Cell line characterization & safety testing : sterility, mycoplasma, virus, retrovirus
 - Genetic stability studies from MCB, WCB, CAL: gene copy number
- GMP Bulk Harvest Release Testing
 - Sterility, in vitro assays, RVLPs quantification, (EM), QPCR
- Host Cell DNA for Non GLP (process development) & GMP testing by qPCR
 - 15 years experience in GMP, FDA, EP, ICH compliant validated assays
- GMP Final Product Batch Release Testing
 - HCD & HCP Endotoxin Ab.Toxicity process related impurity testing
- Regulatory Consultancy
 - Expert Report Custom Assay development & validation