

# Tangential Flow Filtration

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Methods For Successful Scale up

International Workshop on "Vaccine Quality Management"  
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# Presentation Overview

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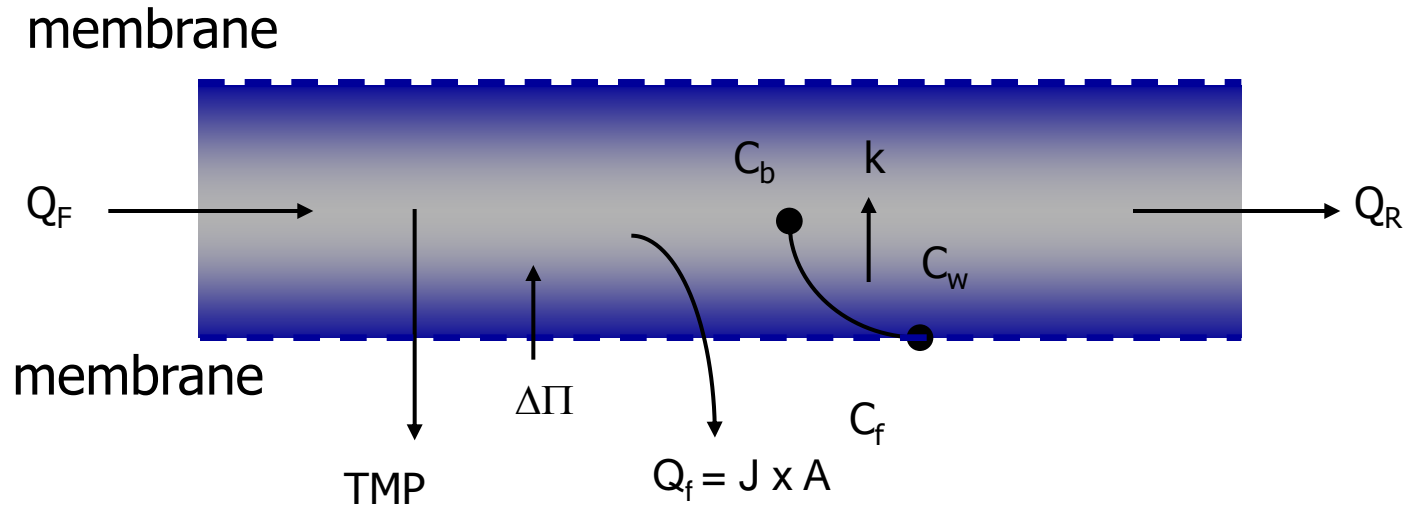
- Scale-up considerations in TFF
  - ◆ Scaling Rule and its Implementation
  - ◆ Various Practical Adaptations of the Scaling Rule
  - ◆ Considerations for Membrane/Module Scale-up
  
- TFF Scale-up
  - ◆ System Design Considerations
  - ◆ Operational Considerations

# TFF Scaling Rule

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- Ensure 'similar' concentration profiles at small and large scales
- What are these concentration profiles and how do we express them?

# Concentration Profiles in a TFF Process



$$\frac{C_{\text{wall/gel}}}{C_{\text{bulk}}} = 1 - R^{\text{Obs}} \left\{ 1 - \exp\left(\frac{J}{k}\right) \right\}$$

Polarization Profile

$$\frac{C_{\text{bulk}}^f}{C_{\text{bulk}}^o} = \exp \left\{ \underbrace{-(1 - R^{\text{Obs}})N}_{\text{DF}} + \underbrace{R^{\text{Obs}} \ln X}_{\text{UF}} \right\}$$

Operating Profile

DF

UF

# Polarization Profile

- Results from steady-state mass transfer considerations
  - ◆ Also called concentration polarization
- Determines
  - ◆ Stability of flux, polarization
- Affected by
  - ◆ Membrane retention
  - ◆ Flux
  - ◆ Cross flow rate
  - ◆ Module Geometry

Affect 'k'

$$\frac{C_{\text{wall/gel}}}{C_{\text{bulk}}} = 1 - R^{\text{Obs}} \left\{ 1 - \exp\left(\frac{J}{k}\right) \right\} \quad \text{where } k = \frac{D}{\delta}$$

Polarization profile is a vital characteristic of a TFF process

# Operating Profile

- Represents mass balance
  - ◆ Establishes perm, retentate volumes during the step
- Establishes concentration profile as a function of
  - ◆ Time
  - ◆ Area
  - ◆ Flux (processing velocity)

$$\frac{C_{\text{bulk}}^f}{C_{\text{bulk}}^o} = \exp \left\{ - \left( 1 - R^{\text{Obs}} \right) N + R^{\text{Obs}} \ln X \right\}$$

Note that X and N can be expressed as function of flux (J), processing time (t) and process loading (V/A)

$$X = \left( \frac{1}{1 - \frac{J_{1\text{avg}} A t_1}{V_0}} \right) \quad N = \frac{J_{2\text{avg}} A t_2}{V_0} \left( \frac{1}{1 - \frac{J_{1\text{avg}} A t_1}{V_0}} \right)$$

# 'Executing' the Scaling Rule for TFF Processes

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- For equivalent polarization profile  $C_{\text{wall/gel}}/C_{\text{bulk}}$ , maintain constant
  - ◆ J or production rate (liters/m<sup>2</sup>-hr)
  - ◆ K or mass transfer coefficient
- For equivalent operating profile, maintain constant
  - ◆ X, N
  - ◆ Order or sequence of X, N

# Production Rate → Area Requirements

- Constant production rate criterion helps establish membrane area requirements ('size') :

$$\begin{array}{ccc} \text{Large Scale (LS)} & = & \text{Experimental Scale (ES)} \\ \text{Production Rate} & & \text{Production Rate} \end{array}$$

$$\text{LS} \cdot \text{Area} = \text{ES} \cdot \text{Area} \times \frac{\text{LS} \cdot \text{Volume}_{\text{Filtered}}}{\text{ES} \cdot \text{Volume}_{\text{Filtered}}} \times \frac{\text{ES} \cdot \text{Time}}{\text{LS} \cdot \text{Time}} \leftarrow \mathbf{1/J_{ES}}$$



# Adaptations of Scaling Rule

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- Equivalent concentration profiles at different scales may be achieved by maintaining
  - ◆ Same polarization, same operating profile **Linear**
  - ◆ Same polarization, similar operating profile **Variation of Linear**
  - ◆ Similar average polarization & similar average operating profile **Serial**
  - ◆ Similar theoretical polarization & similar average operating profile **Theoretical**
- UF: 'Variation of linear' scaling ('lab' scale) followed by linear (pilot) scaling is most common
- MF: 'Serial module' scaling ('lab' scale) followed by linear scaling (pilot) is commonly used

# Linear Scaling

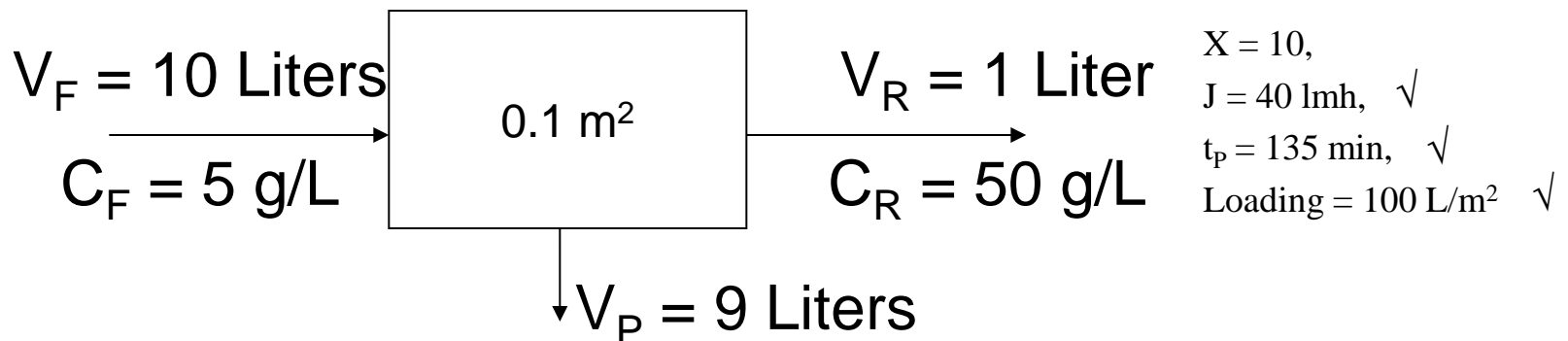
- Characteristics – same ‘everything’ (well ‘almost’)
  - ◆ Same membrane material, MWCO, device format
  - ◆ Same flux (Production Rate)
  - ◆ Same  $k$  (Cross Flow Rate, Module Geometry:  $h$ ,  $L$ )
  - ◆ Same processing time or Volume/Area loading
  - ◆ Only Module width,  $w$ , changes from small to large scale

} Same  
Polarization  
Same  
Operating  
Profile
- Need
  - ◆ Linearly Scale-down Device for process development
  - ◆ Large volume for process development



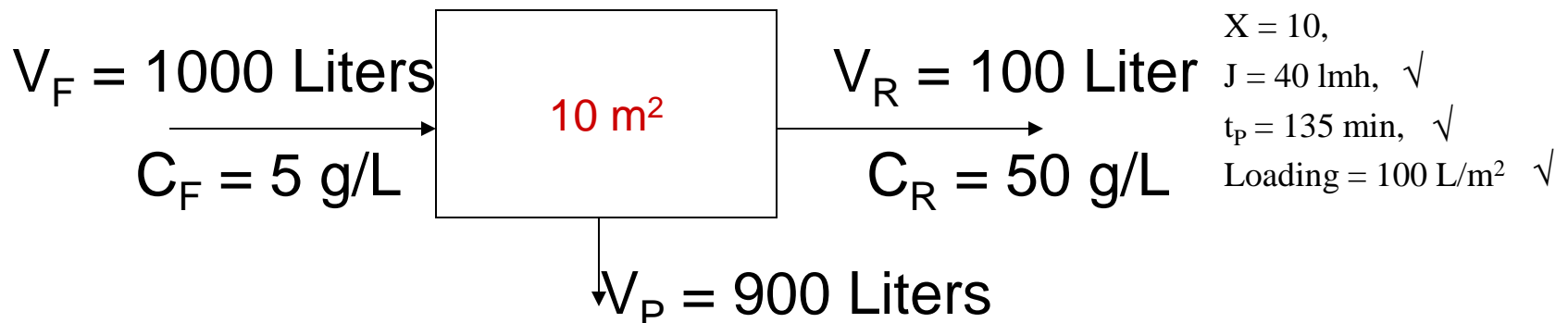
# Example of Linear Scaling

## UF Process

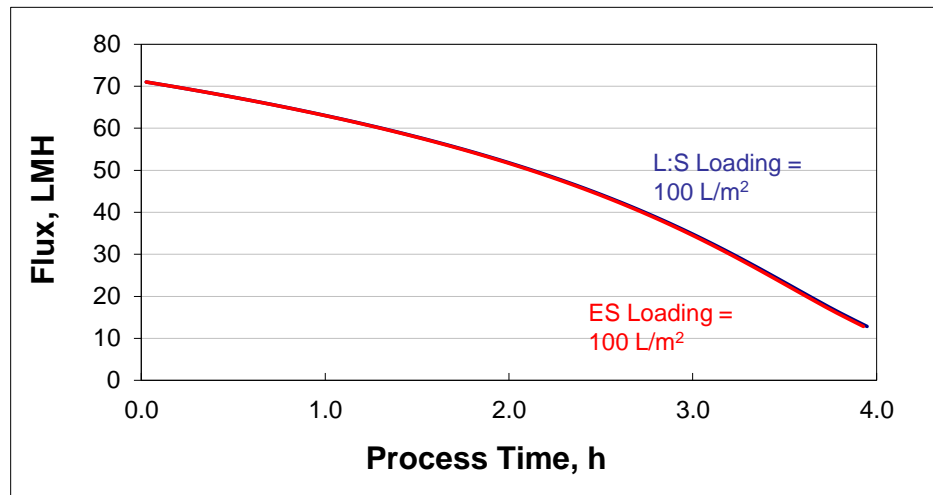
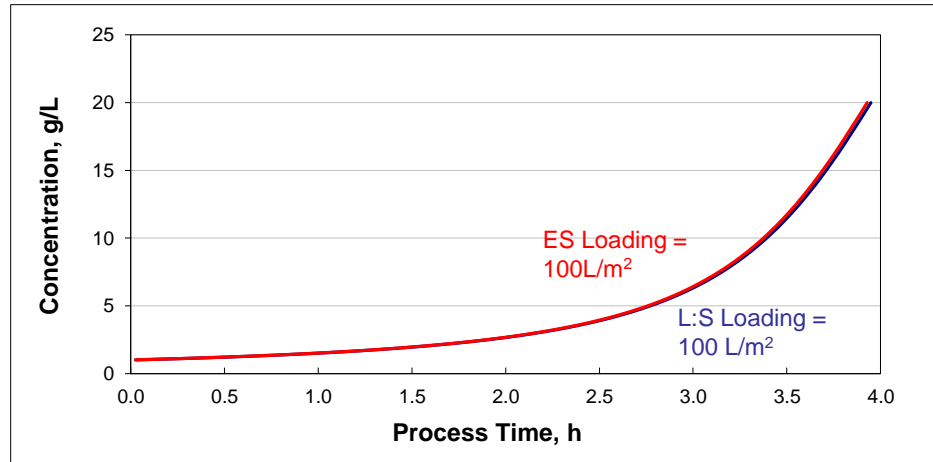


Same 'Fouling' risk

$$\text{LS} \cdot \text{Area} = 0.1 \times \frac{900}{9} \times \frac{135}{135}$$



# Concentration and Flux Profiles – Linear Scaling



# Variation of Linear Scaling

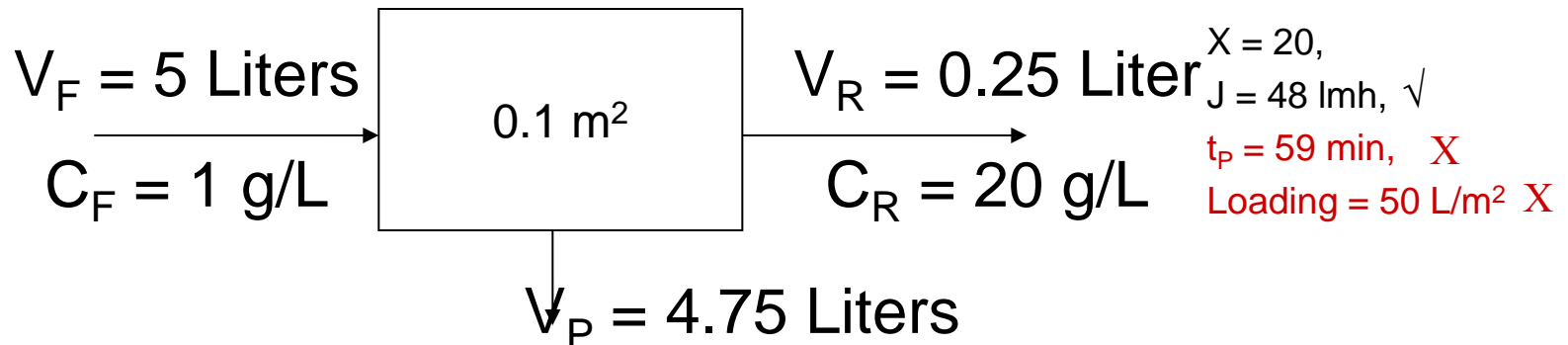
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## ■ Characteristics

- ◆ Meets all criteria for linear scaling **except processing time or volume/area loading** i.e.
  - Same membrane material, MWCO, device format
  - Same average flux (Production Rate)
  - Same  $k$  (Cross Flow Rate, Module Geometry:  $h$ ,  $L$ )
  - ~~Same processing time or Volume/Area loading~~
  - Module width,  $w$ , changes from small to large scale
- ◆ Works well for UF-DF processes within limits
  - Limit  $< 2$  fold process time extension
  - Requires considerably less volume for process development
  - Development time is short
- ◆ Should not be used for MF-DF processes

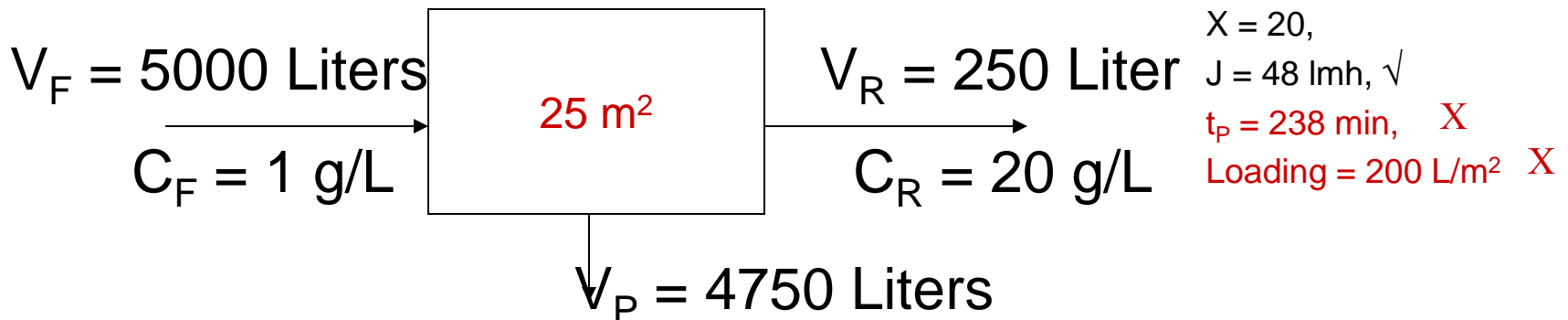
# Example: Variation of Linear Scaling

## UF Process

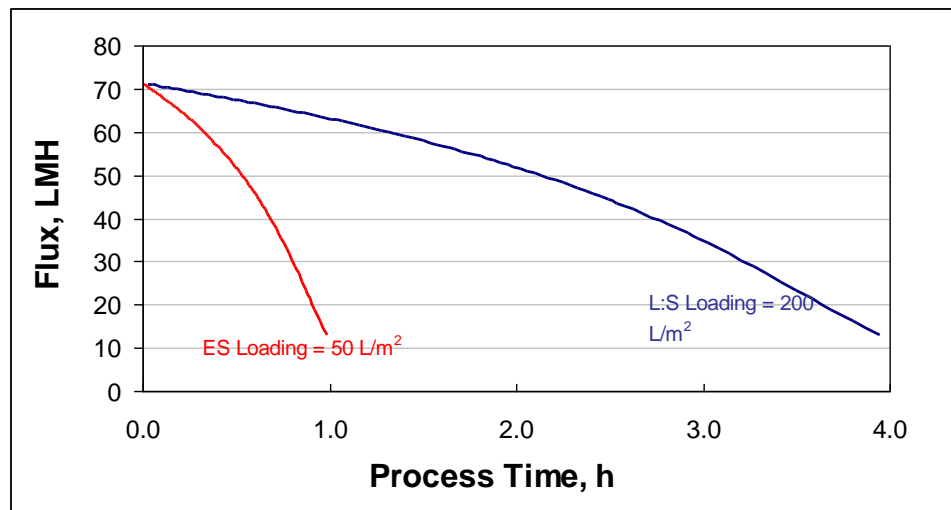
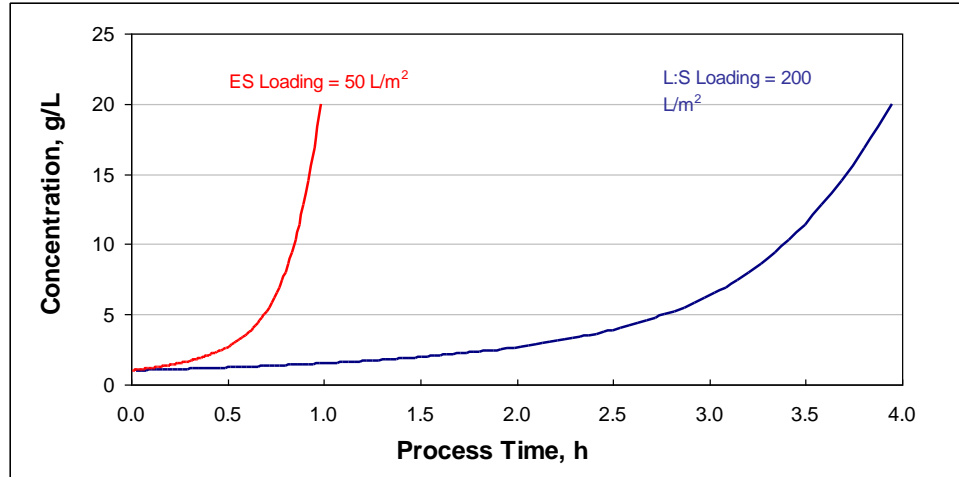


$$\text{LS} \cdot \text{Area} = 0.1 \times \frac{4750}{4.75} \times \frac{59}{238}$$

'Fouling' risk could be different due to different processing times



# Concentration And Flux Profiles – Variation of Linear Scaling



# Linear Scaling: Implications for Module Design

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- Feed Channel Length
  - ◆ Constant for all device sizes
- Entrance/Exit Effects
  - ◆ Consistent inlet/outlet porting between sizes
- Channel Compression
  - ◆ Stable channel height
  - ◆ Controlled manufacturing

*'Geometric Similarity' is essential to ensure equivalent velocity, concentration and temperature profiles!*



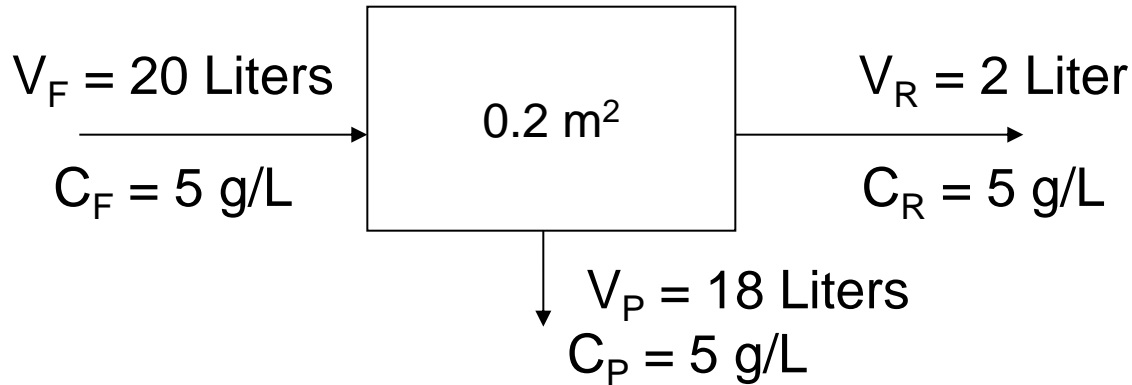
# Serial Module Scaling

## ■ Characteristics

- ◆ Stack multiple modules in series to reduce cross flow (pump size) and system size (piping size etc)
  - Fluid flow Path length,  $L_{\text{effective}}$ , changes
  - Cross-flow in a serial bank of modules is same as that for a single module
  - Widely used for MF processes with open channel devices: Prostack, Hollow Fibers
- ◆ Same membrane material, MWCO, device format
- ◆ ~~Same~~ **similar** average flux (Processing Rate)
- ◆ ~~Same~~ **similar**  $k$  (Cross Flow Rate, Module Geometry:  $h$ ,  $L$ )
  - $L_{\text{effective}}$  is changed
- ◆ Same processing time or Volume/Area loading
  - For MF process
  - Not so for UF process
- ◆ Module width,  $w$ , changes from small to large scale

# Example: Serial Module Scaling

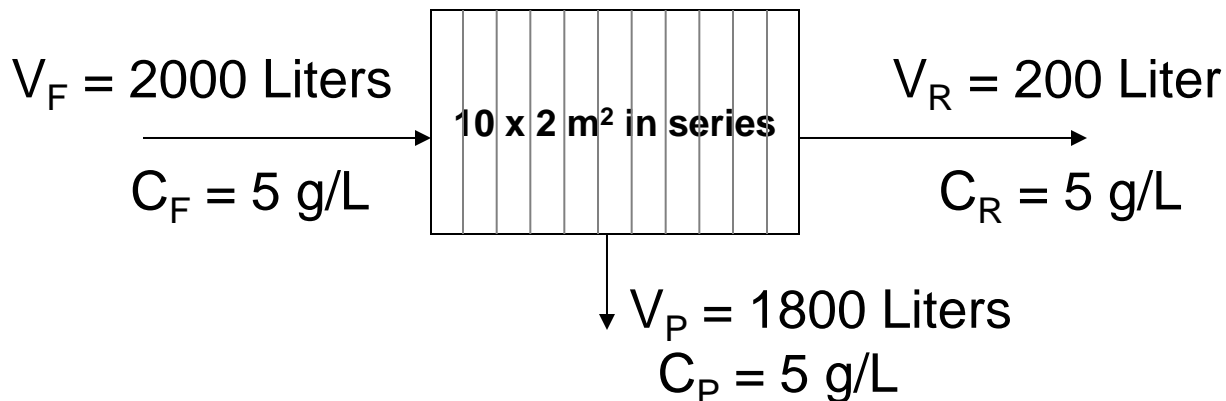
## MF Process



$X = 10,$   
 $J = 30 \text{ lmh}, \checkmark?$   
 $t_p = 180 \text{ min}, \checkmark$   
 $\text{Loading} = 90 \text{ L/m}^2 \checkmark$

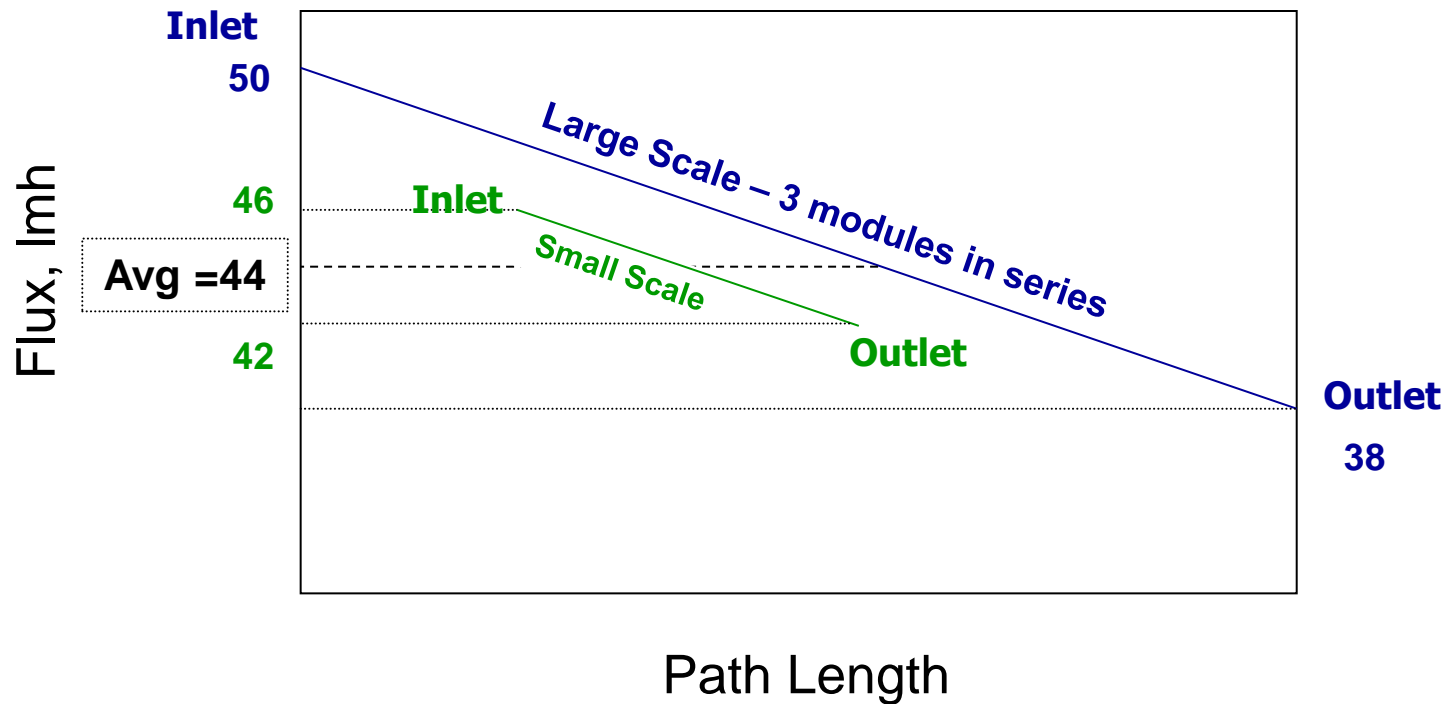
$$\text{LS} \cdot \text{Area} = 0.2 \times \frac{1800}{18} \times \frac{180}{180}$$

Different Fouling risk, DP,  
 Flux/TMP Profile → Must be accounted  
 For during Scale-up



$X = 10,$   
 $\text{Avg } J = 30 \text{ lmh}, \checkmark?$   
 $t_p = 180 \text{ min}, \checkmark$   
 $\text{Loading} = 90 \text{ L/m}^2 \checkmark$

# Potential Issues with Serial Module Scaling



- How much variation in Flux, TMP is considered acceptable?
  - ◆ Depends on the process, fluid, module construction – need good process characterization data
  - ◆ Up to 3-5 in-series for UF; Up to 10 for MF
    - Always test the serial configuration in a scale-down format for confirmation

# Non Linear/'Model-Based' Scaling

Uses predictions to estimate performance at different scales

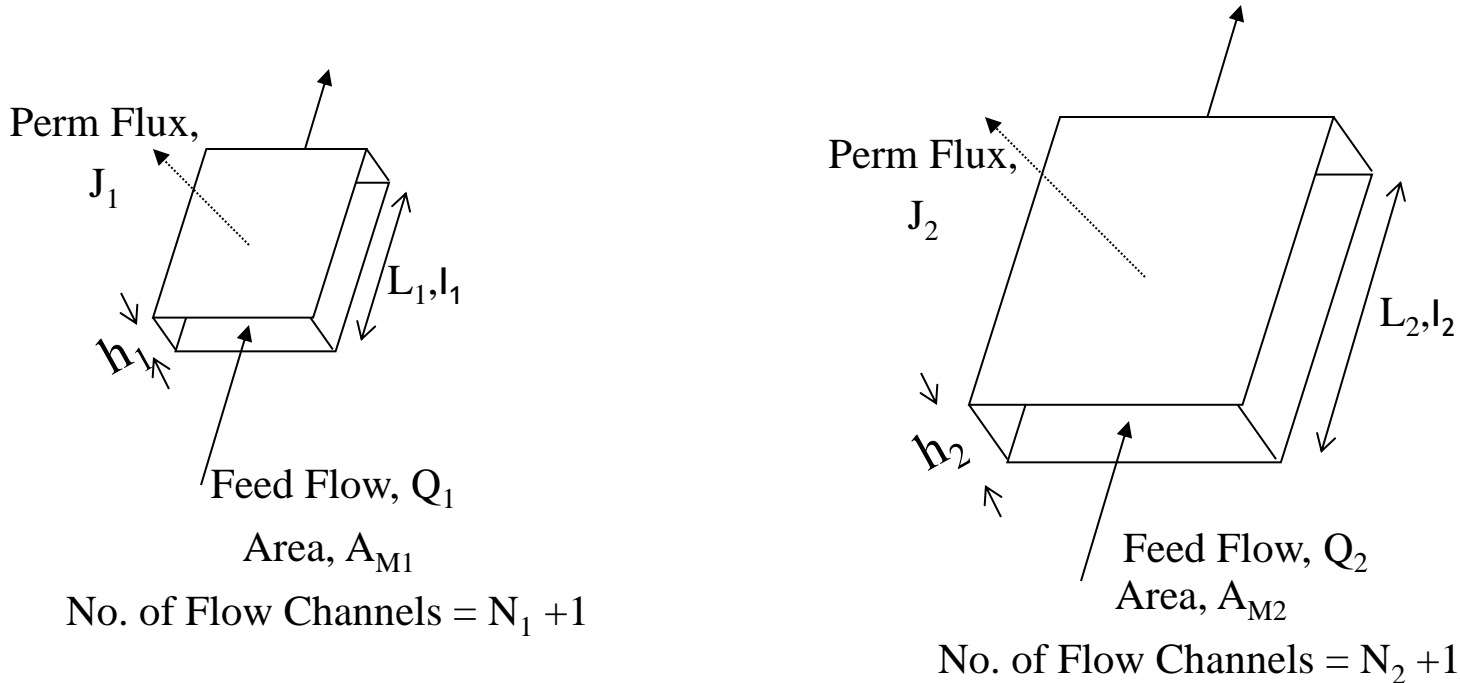
## ■ For Similar Geometry

- ◆ Modules are geometrically similar
  - Ex Hollow Fiber or flat sheet with  $d_1, L_1 \rightarrow d_2, L_2$
- ◆ Same membrane material, MWCO, device format
- ◆ Same average flux (Processing Rate)
- ◆ ~~Same  $k$  (Cross Flow Rate, Module Geometry:  $h, L$ )~~
  - Equivalent  $k$  is obtained using mass transfer scaling correlations
- ◆ ~~Same processing time or Volume/Area loading~~
- ◆ Module width,  $w$ , may change from small to large scale

## ■ For Dissimilar Geometry

- ◆ Ex. Flat sheets to Spirals scale-up/scale-down
- ◆ Not Recommended

# Example: Non-Linear Scaling with Similar Geometry



For Equivalent Polarization, maintain same  $J$  (Flux) and  $k$  (mass transfer coefficient):

$$\text{Use } J_1 = J_2 \text{ and } \frac{Q_2}{Q_1} = \left[ \frac{N_2 + 1}{N_1 + 1} \right] * \left[ \frac{N_1}{N_2} \right] * \frac{A_{M2}}{A_{M1}} * \left( \frac{h_2}{h_1} \right)^2 * \frac{L_2}{L_1} * \left[ \frac{l_1}{l_2} \right]$$

# TFF Scale-up:

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System Design Considerations

The 'Non-Linear' aspects of the Process

# Pump Effects

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- Issues:
  - ◆ Pump configuration and number of pump passes
  - ◆ Final working volume
- Pump configuration:
  - ◆ Avoid micro-cavitation (air-liquid interface)
  - ◆ Minimum pressure at inlet of pump should equal manufacturer's specification (NPSH requirements)
    - Inlet line to the pump need to be properly sized

Quality

# Pump Selection

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## ■ Pump Style

- ◆ Circumferential piston displacement style is most common in biotech for large-scale
  - Waukesha and Viking are the major players – single and dual-lobe
- ◆ Quattro-Flow is a relatively new option for smaller scale (4-chamber diaphragm)
  - Gentle protein processing

## ■ Rotor Clearance

- ◆ Don't specify heat-tolerant rotors unless required – greater slip
- ◆ Rotor clearance does not scale linearly with pump size
  - More protein damage likely in smaller pumps because clearance (slip) is a larger percentage of the total displacement



# Pump Selection

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- RPM and Sizing
  - ◆ Typical target is 400 – 500 rpm for protein processing, lower for cell processing
  - ◆ Design flowrate typically ~ 80% of pump maximum
- System Integration
  - ◆ Minimize distance and equipment from tank outlet to pump and use adequate ID's to avoid starving the pump
  - ◆ Target developing a cleaning protocol that uses the same feed flowrate as the process so that the pump (and piping) does not have to be oversized just to accommodate cleaning!

# Tank Design

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- Characteristics of a good tank design
  - ◆ Low Holdup Volume
  - ◆ Able to function at high (20-100 fold) volumetric reduction
    - No foaming or air entrainment
  - ◆ Good Mixing at low levels
    - Ensures homogeneous concentrations
    - Prevents retentate short circuit
  - ◆ No spray ball shadows
    - Efficient CIP

**Yield**

**Quality**

**Purity**

# Tank Design Guidelines

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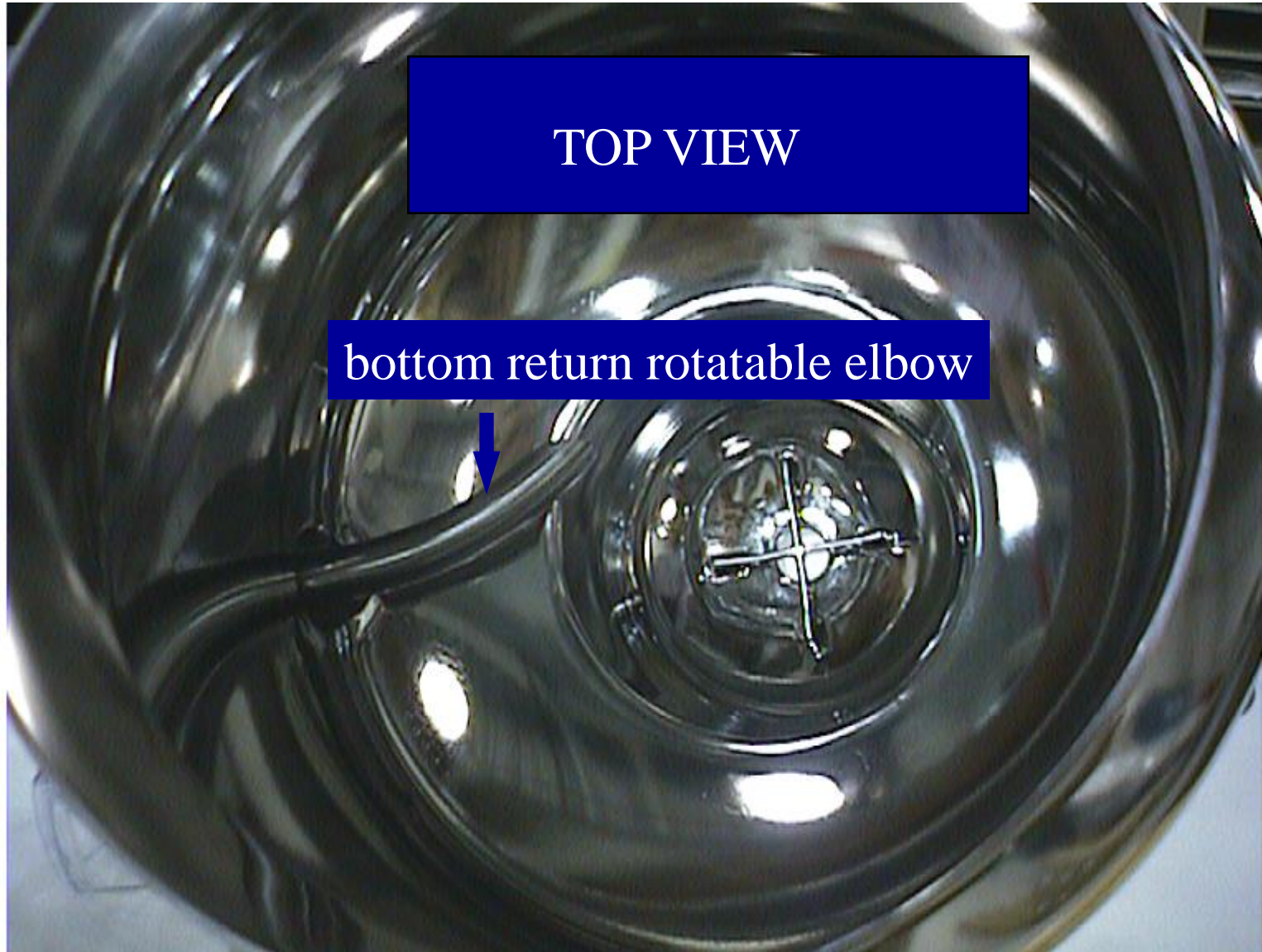
- Sizing
  - ◆ Maximum volume based on both a reasonable fed-batch ratio and on a reasonable cleaning solution volume
    - FBR ~ 4-5, cleaning flush ~ 10 L/m<sup>2</sup>
- Retentate entry
  - ◆ Very low on the tank sidewall to minimize retentate piping and also enable low tank volume operation
  - ◆ Design of dip tube is important to ensure good mixing and low turbulence
- Lots of experience with two key tank design types
  - ◆ Reduced bottom well (tulip style)
  - ◆ Conical bottom
  - ◆ Both can achieve low working volumes and good mixing
    - Conical may have an edge for cleanability, mixing, and level measurement

# Tank Designs



Retentate return

# Recirculation Tank

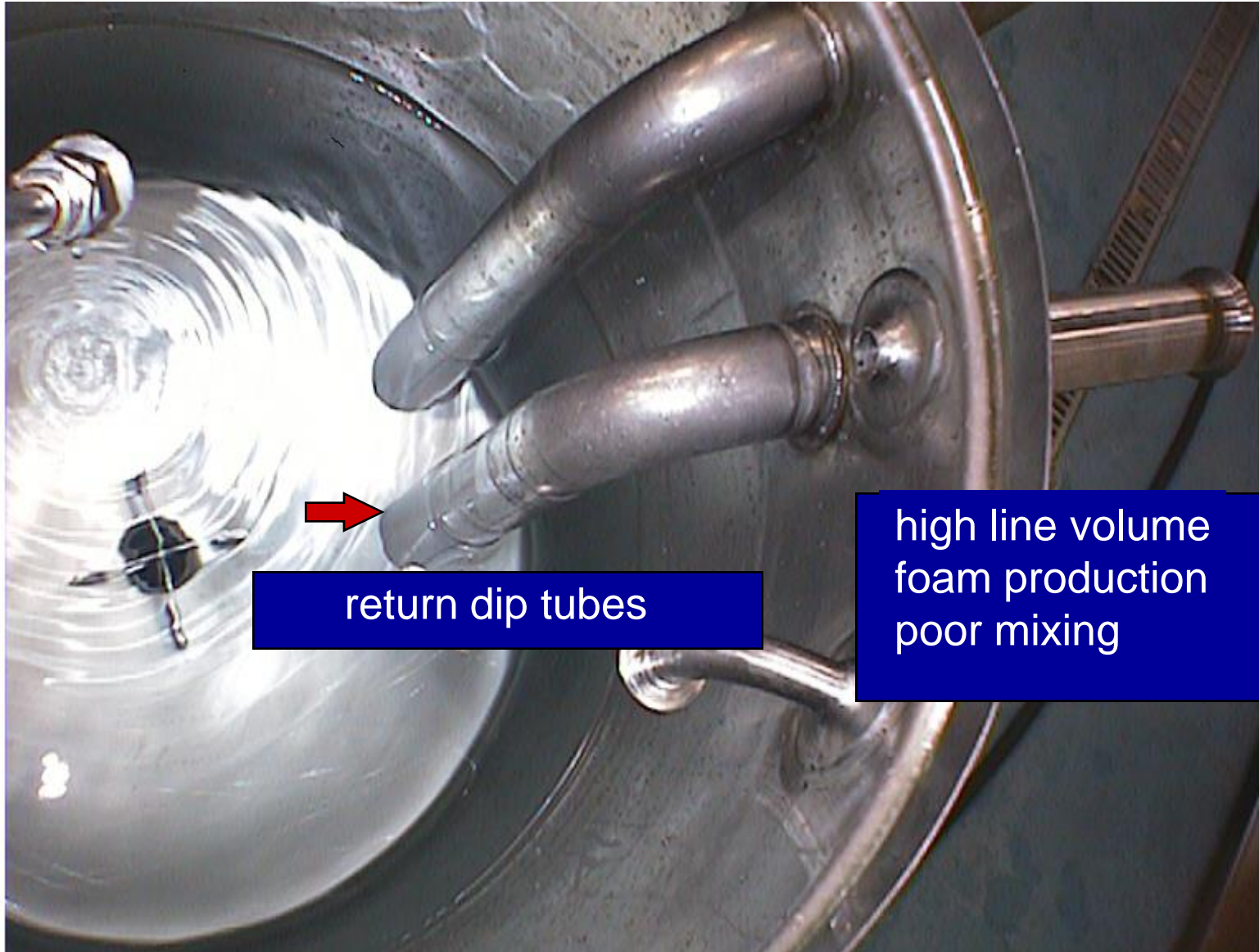


TOP VIEW

bottom return rotatable elbow



## Example of Improper Tank Design



return dip tubes

high line volume  
foam production  
poor mixing

## Integration of tank and recirc loop



# Other Considerations

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- Minimize hold-up
  - ◆ Minimize volume in piping
    - Fitting to fitting connections when possible
    - Instrument blocks incorporated onto membrane holders
    - Return dip tube into sidewall of recycle tank
  - ◆ Slope all lines to a low-point recovery port (1/16" per ft)
    - Tank on top of a vertically-mounted pump works well
    - Include a high-point air port for blowdown
- Avoid air/liquid interfaces
  - ◆ Very damaging to proteins
  - ◆ Vortexing in recycle tank (air entrainment)
    - design of diptube return and agitator placement/speed
- Ensure a well-mixed process stream
  - ◆ Agitator design and placement
  - ◆ Retentate dip-tube return and DF buffer stream entry
    - Ensure that DF buffer doesn't float at top of tank while retentate short-circuits into feed intake



# Other Consideration (Cont'd)

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- Minimize deadlegs and contamination points:
  - ◆ Use short pull T's, NA connects, and zerostatic valves
  - ◆ Avoid flex lines
  - ◆ Minimize clamp connections
  
- System Size Flexibility
  - ◆ Trying to be too flexible compromises the system design for all scales of operation
  - ◆ Recommend a maximum of approximately 4-fold membrane area or flow range on a single system (2-fold is better)
  
- Line size:
  - ◆ Balance between pressure drop and holdup considerations
  - ◆ Maintain turbulent flow (5-7 ft/sec during CIP)
  
- Temperature control:
  - ◆ Jacketed tank vs in-line heat exchanger

# Operational Considerations

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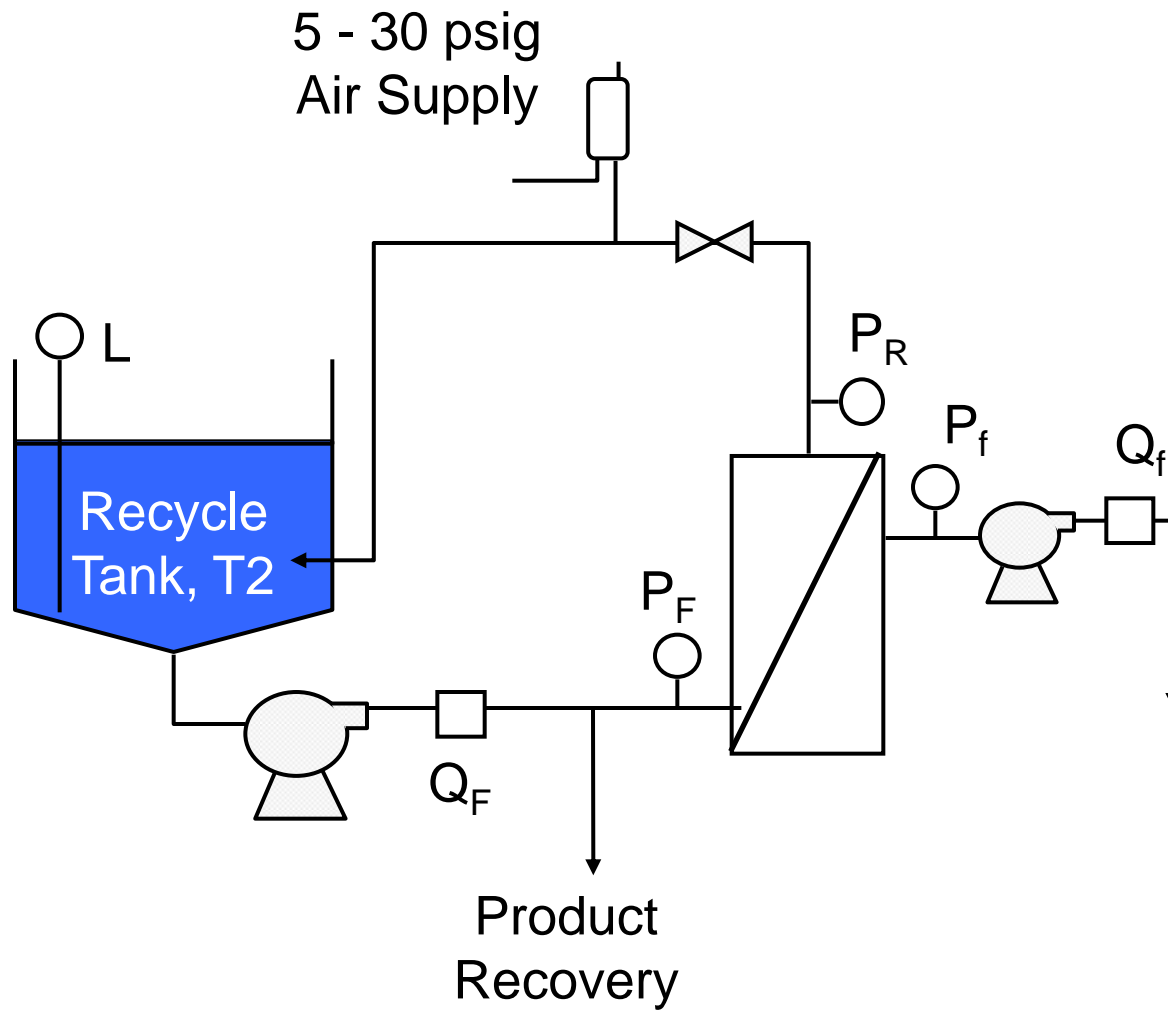


# Operational Considerations

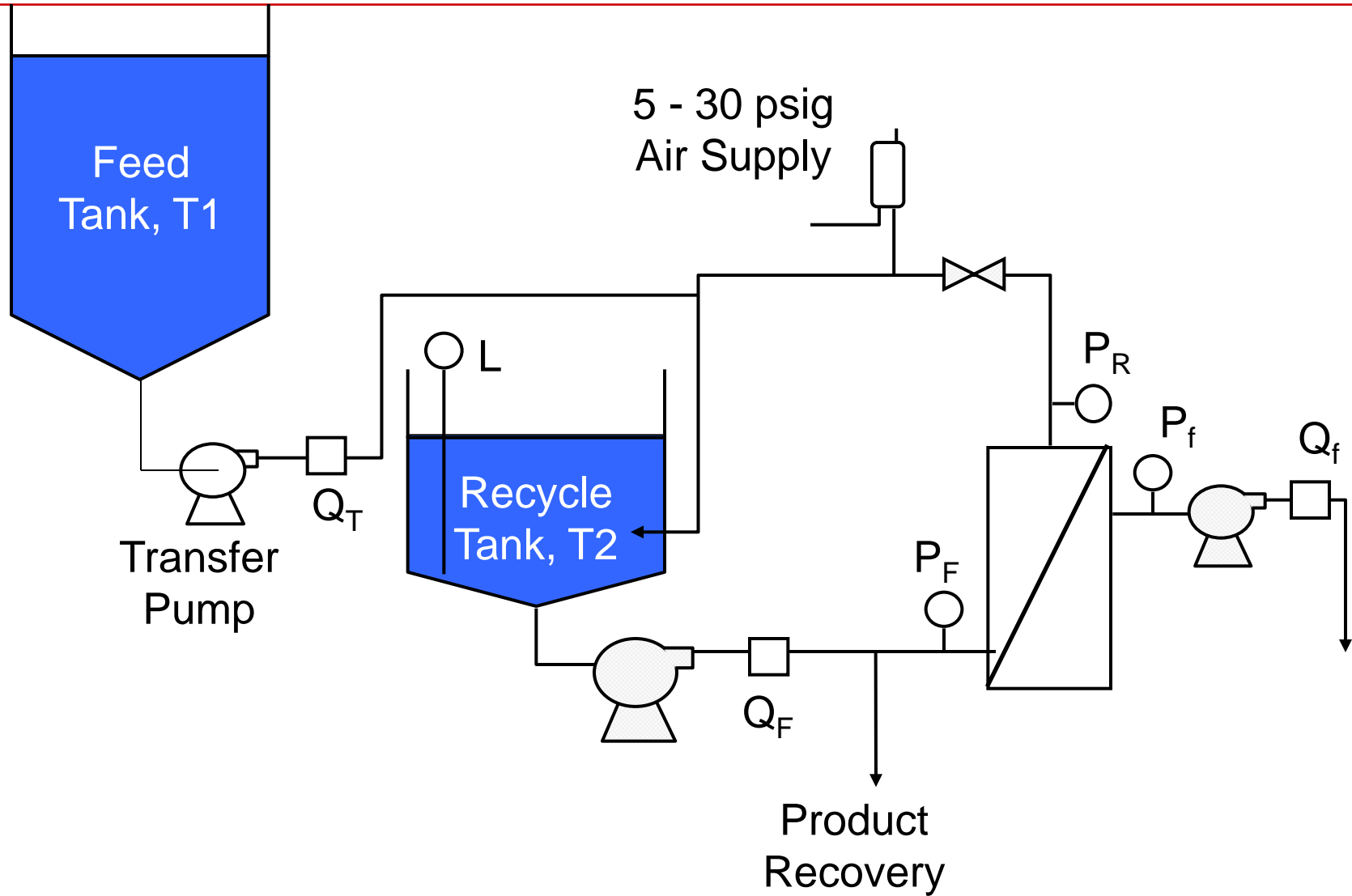
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- Batch Vs Fed Batch
- Scale-up of process steps:
  - ◆ Module Installation, flushing storage solution, Integrity testing, NWP testing, Buffer Equilibration, Product Recovery, Cleaning/storage, Reuse
- Process Control

# Batch



# Fed-Batch

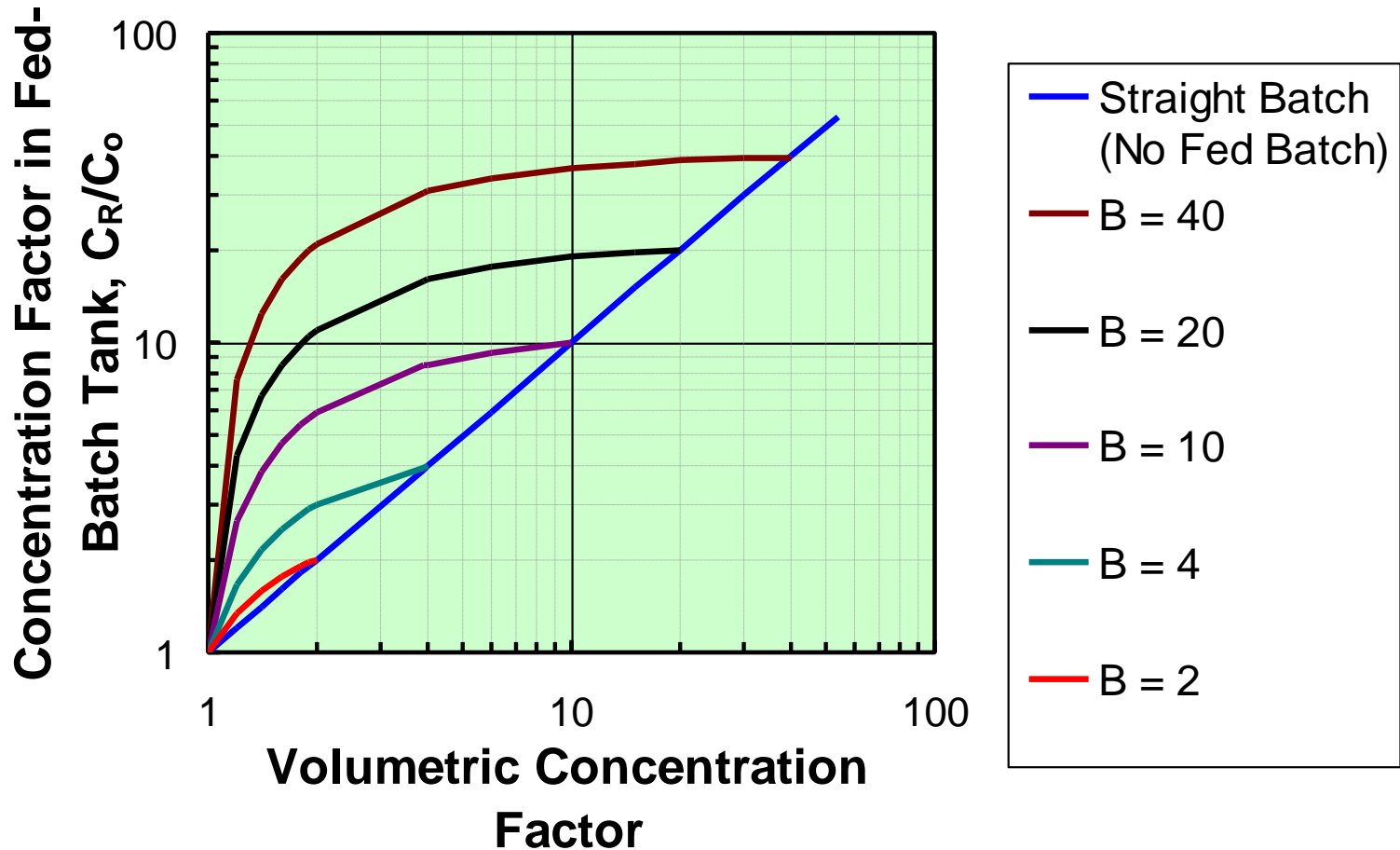


# Batch versus Fed-Batch UF

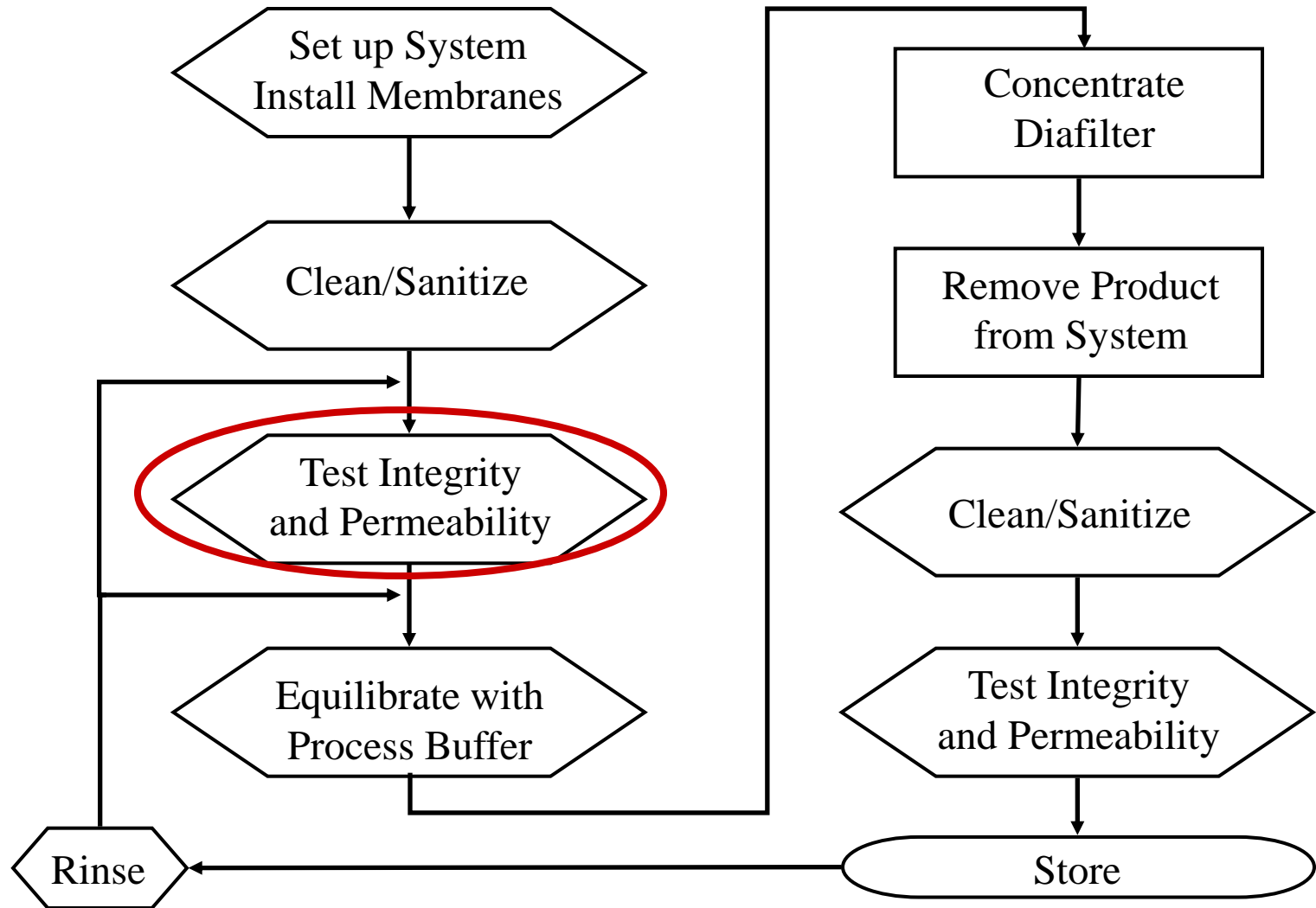
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- Batch UF is preferable whenever possible
  - ◆ Simplest system and control scheme
  - ◆ Feed is at lower  $C_{avg}$  → so less membrane area
  - ◆ Maximum achievable VCF ~ 20X
    - System or hardware limits: level measurement, mixing are reached
- Fed-Batch UF
  - ◆ Usually required for VCF > 10 - 15X
  - ◆ Minimize fed-batch ratio (= Total  $V_o$  / Recycle tank  $V$ ) as much as possible to minimize membrane area
  - ◆ Maximum achievable VCF ~ 50 - 100X
  - ◆ Test process performance using proposed fed-batch ratio

# Effect of Fed-Batch Ratio on Concentration in Recycle Tank



# A Typical Process Sequence



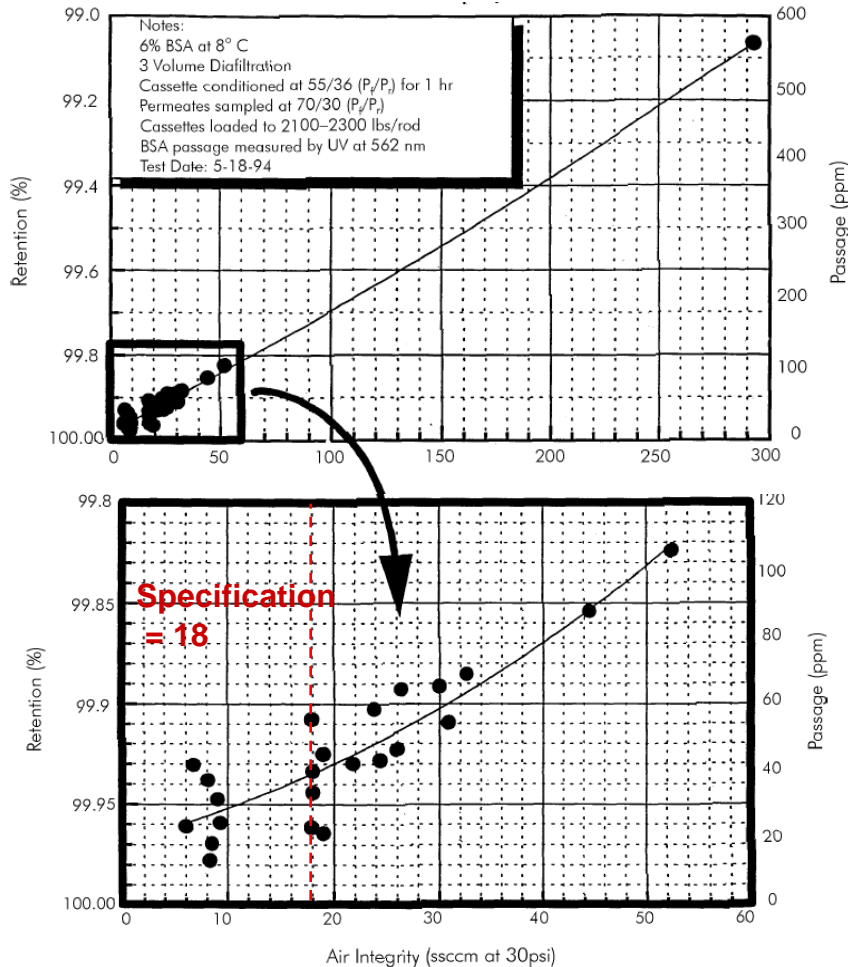


# Integrity Testing

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- Many UF/MF processes use Air-Diffusion ‘integrity’/leak test prior to processing
  - ◆ Convenient and non-destructive way to check/confirm
    - Proper module installation
    - Damage during shipping/handling etc
    - Provides a gross measure of system integrity
  - ◆ Issues:
    - False Integrity failures can occur due to
      - Incomplete Cleaning
      - Lack of adequate wetting

# Correlation to Protein Retention?



**Integral Modules:**

$R_{BSA} > 99.9\%$  at Air Flow  $< 25$  scc/min

**Non-Integral Modules:**

$R_{BSA} > 99\%$  at Air Flow = 270 scc/min

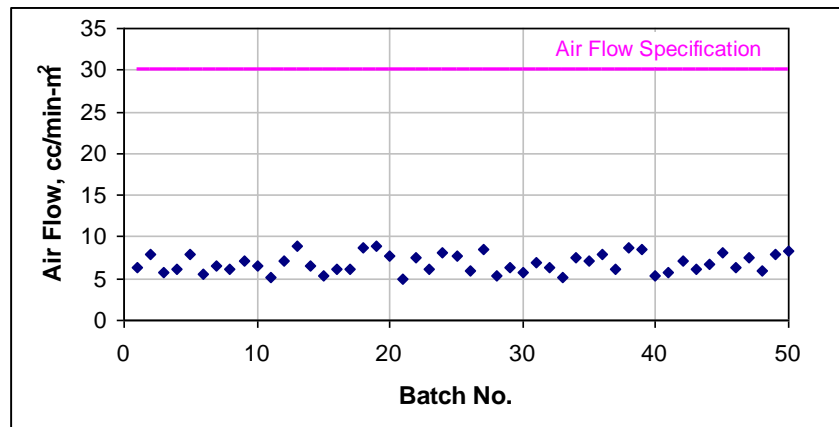
For  $X=20$ ,  $N=5$ , this represents an additional yield loss of 6.3%

Gross difference in air flow may indicate issues

# Integrity Specification

$$\text{Specification} = N_{\text{Modules}} \times \text{Specification}_{\text{Individual Module}}$$

- For large installations, risk of letting a non-integral module go 'undetected'
  - ◆ Reduce risk by trending diffusion values over batch runs
  - ◆ Alternate Spec. based on statistics
    - 'Limited benefit for added the effort'
    - Risk of false failures
  - ◆ Other complementary tests: permeate sampling for product etc



# Integrity Testing – Practical Aspects

- It is useful to have a single module holder in hand to individually test modules in the event of a failure
- Cassette devices rely on 'sealing force' to seal feed from permeate
  - ◆ Often a torque specification is provided by the manufacturer
    - The torque is a function of tie-rod type, thread, nut-size etc
    - The torque may be converted into sealing force as:
      - $F = T/(u \times d)$  where  $F$  = Force,  $T$  = Torque,  $u$  = friction factor,  $d$  = thread diameter
  - ◆ Many large scale systems have hydraulic closure mechanisms
    - Here, the sealing force is a function of the seal pressure resulting from the cylinders
      - Force = Effective Cylinder Area x Cylinder Pressure
  - ◆ Need to carefully review these factors when installing one set of filters into holders built by a different manufacturer

# Membrane Permeability Testing

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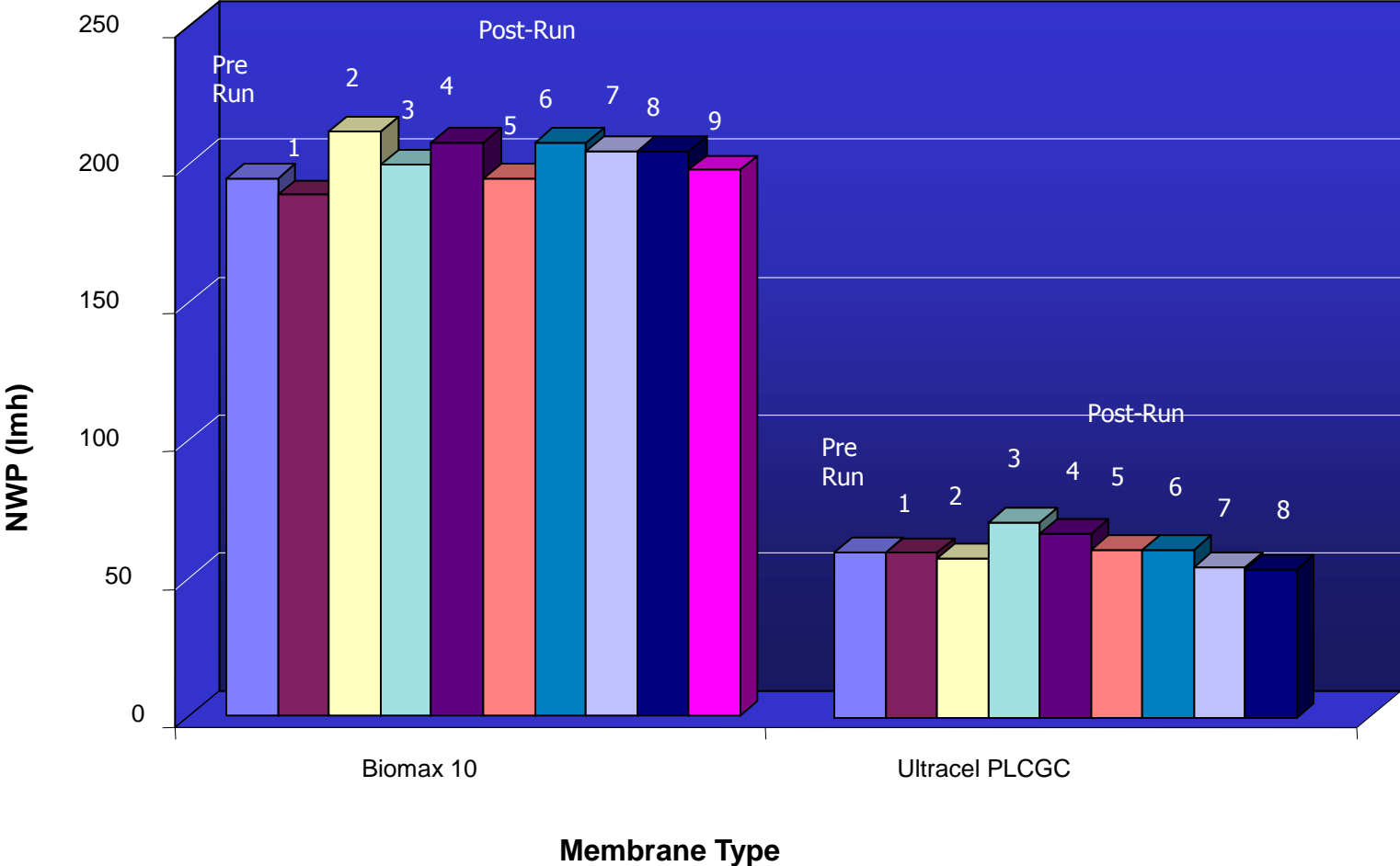
- Easy and nondestructive means of confirming the effectiveness of cleaning procedures and process consistency
- Strict definition is normal water permeability (NWP), but any solution is OK as long as you're consistent
  - ◆ Best if test solution is similar to water (viscosity)
  - ◆ Testing on storage solution is convenient
- Record temperature for accuracy
  - ◆ As well as flows and pressures

# NWP – Practical Aspects

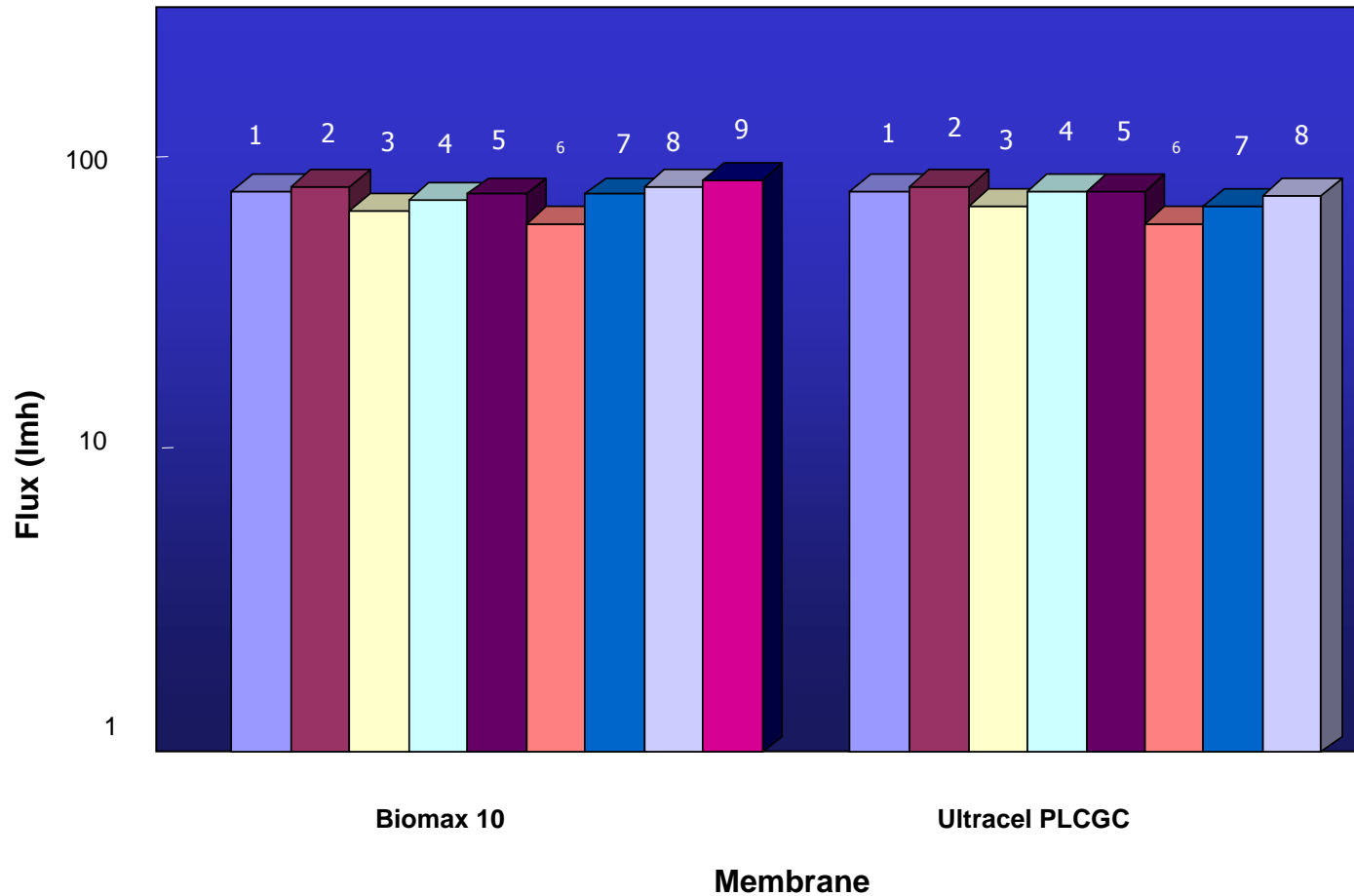
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- Must be correctly used to serve as a ‘cleanliness’ indicator
  - ◆ Reliable as an indicator for tighter UF membranes ( $\sim \leq 300$  kD)
  - ◆ For more open UF membranes & MF membranes, use NWP as a guideline rather than a specification:
    - Pressure gauge accuracy at low TMPs
    - Process permeability & TMP are stronger indicators of performance consistency
- Specification: Select NWP of the system after new filters are installed, flushed & cleaned to remove preservatives
  - ◆ Choose a  $\pm$  range for acceptance criteria (varies between 60 - 80%)
- For improved reliability:
  - ◆ Use measurements at multiple TMPs (5, 10, 15 psi)

# NWP Recovery after Cleaning



# Process Permeability versus Batch Cycle





# Product Recovery

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- Product recovery strategy is often ignored at the small scale
  - ◆ Manual recovery procedures are employed (may include unclamping the unit at critical points to recover product)
  - ◆ Protein retention close to 100% is deemed a sufficient indicator for yield
- For large scale process, product recovery strategy is critical to ensure high yield
  - ◆ Gravity drain & buffer flush are often used to recover product
  - ◆ Product may still be trapped in the 'unrecoverable' holdup in the system and may contribute to significant yield loss
  - ◆ Enhanced product recovery techniques may be employed to boost yield

# Product Recovery Techniques

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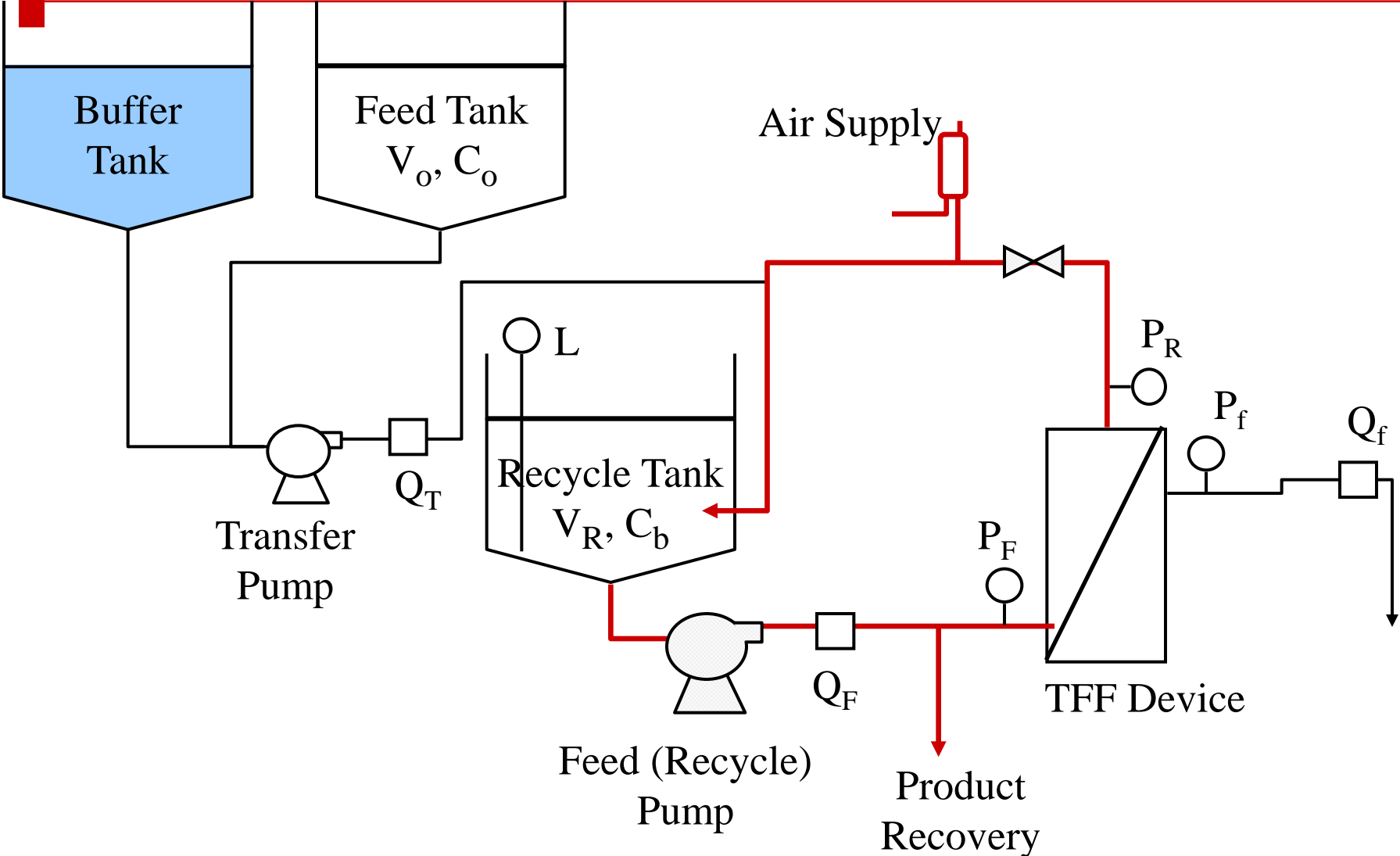
- First, release polarization layer and mix feed
- Then, five recovery options to consider
  - ◆ System drain
  - ◆ System blowdown
  - ◆ Buffer flush
  - ◆ Buffer recirculation

# Simple Product Recovery Options

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- System Drain
  - ◆ Simplest!
  - ◆ Most effective if system has low-point drain port
  - ◆ Usually used in combination with another option
  - ◆ Works better for some devices than others
  
- System Blowdown
  - ◆ Air-pressure assisted drain
  - ◆ Most effective if system has high-point air port
  - ◆ Most effective to use low (5 psig) air pressure
    - Once an air path exists, recovery is finished

# Drain and Blowdown Flowpath

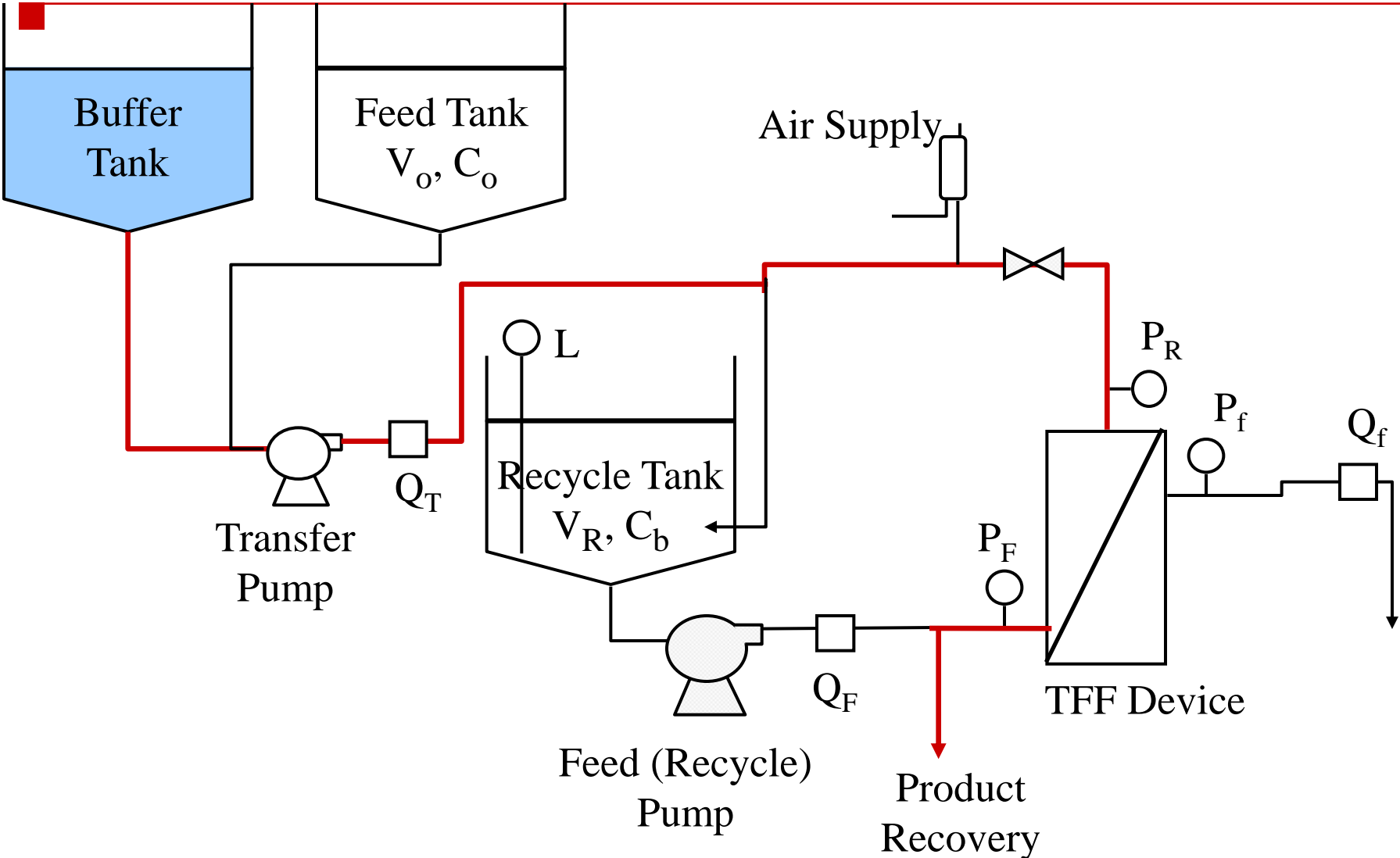


# Buffer Flush or Recirculation

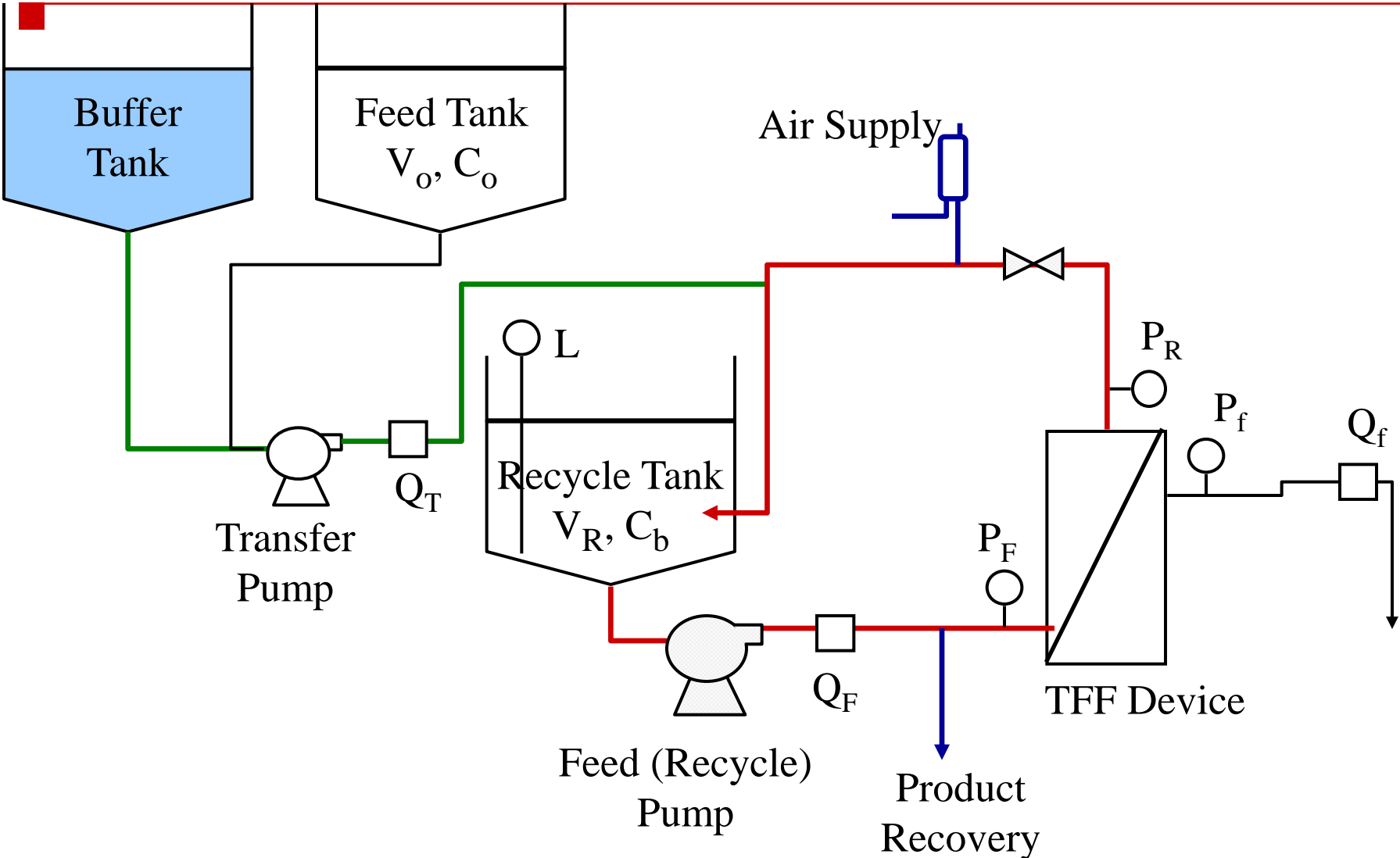
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- These are very commonly used
- Buffer Flush
  - ◆ Flush buffer in single pass, through drained system
  - ◆ Approximately one system volume of buffer is required
  - ◆ Entire volume of buffer is added to product, so dilution must be acceptable
- Buffer Recirculation
  - ◆ Add buffer to empty recycle tank, recirculate through empty system, then recover
  - ◆ same dilution concerns as with buffer flush

# Buffer Flush Flowpath



# Buffer Recirculation Flowpath



# Cleaning/Storage Considerations

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- Establish a robust cleaning procedure:
  - ◆ Cleaning solution make-up:
    - If dosing with a high concentration stock solution, select the dosing point carefully → Ex. downstream of retentate line to maximize the probability of thorough mixing.
  - ◆ Allow NO significant lag time between end-of-processing (conc/recovery) and cleaning → ideally < 1 hr
  - ◆ Ensure CIP chemicals are 'clean'
  - ◆ Ensure there are no dead legs, paths during the CIP cycle.
  - ◆ Provide for partial 'flush' with cleaning solution before recirculation
  - ◆ Allow adequate distribution of flow between retentate and permeate side → target 30% conversion
    - Permeate restriction for MF and open UF membranes
    - Adequate TMP for tight UF membranes



# Reuse Considerations

- Validation Requirements: Ensure

- ◆ Reproducible process
  - Demonstrate performance consistency
    - Flux, TMP, crossflow, Yield, NWP, Integrity test
- ◆ Contaminant clearance
  - Flush water residuals – measure TOC, sample for cleaning/sanitizing agents
  - Lot-to-lot product or contaminant carryover levels
- ◆ System is free of bioburden
  - Live bacteria, mold, spores, etc
  - Endotoxin levels (measured by LAL test) are below spec.

Full or partial data from small scale

Demonstrate at large scale to # Of reuses

- Economic Tradeoff

- ◆ Membrane cost per run decreases with increasing number of uses
- ◆ However, validation effort can become oppressive

# Process Control

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Objective: Key process parameters need to be consistently maintained at fixed setpoints

- Crossflow Control

- ◆ Creates the tangential flow (sweeping action)

- Retentate Pressure Valve Control

- ◆ Creates driving force through the membrane to cause flux

- Other Control

- ◆ Filtrate Control

- Required to moderate the flux on large pore size UF and all MF systems

- ◆ Level, Temperature

# Crossflow Control Options

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- Modulate the feed pump speed throughout a process to maintain:
  - ◆ Pressure Drop setpoint
  - ◆ Feed Rate setpoint

# Crossflow Control by Constant Pressure Drop

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- Simplest to implement (least expensive!)
  - ◆ Only pressure gauges required for control
- Actual crossflow is unknown
  - ◆ Less process information
  - ◆ Crossflow changes if viscosity changes significantly during run
- Lot-to-lot & within-run pumping requirements vary
  - ◆ Must have adequate high and low-end pumping capacity to span the potential range
- Scaleup / scaledown pump sizing more difficult

# Crossflow Control by Constant Feed Rate

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- Actual crossflow is known
  - ◆ More information on system performance
  - ◆ Crossflow is maintained if viscosity changes during run
- Lot-to-lot and within-run pumping req's are constant
  - ◆ Maximum pumping capacity need is known
- Scaleup / scaledown pump sizing is straightforward
  - ◆ Simple ratio of membrane area difference
- Requires feed flow measurement

# Retentate Pressure Valve Control Options

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- Modulate the retentate valve position throughout a process to maintain:
  - ◆ Pressure setpoint
    - Retentate pressure
    - TMP
  - ◆ Flux setpoint
    - Constant
    - Variable throughout run

# Retentate Valve Control Using a Pressure Setpoint

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- Retentate Pressure Setpoint
  - ◆ Simplest, robust control scheme for most UF processes - most widely used
  - ◆ Requires only retentate pressure measurement
- Transmembrane Pressure Setpoint
  - ◆ More complex than simple retentate pressure setpoint
  - ◆ For processes with constant  $\Delta P$ , no real advantage
  - ◆ Maintains more constant pressure during viscosity changes
  - ◆ On-line TMP calculation required for an automated system

# Retentate Valve Control Using a Flux Setpoint

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- Constant Flux setpoint
  - ◆ More typical for MF and Virus-TFF processes than UF (often via a filtrate pump)
- Variable Flux setpoint
  - ◆ Sophisticated UF control scheme to maintain constant protein concentration at membrane surface ( $C_{wall}$ ) regardless of bulk concentration or membrane permeability
  - ◆ Impossible to implement on manually-controlled system
- Both schemes require filtrate flow measurement



# Filtrate Control

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- Most UF processes (< 100kD)
  - ◆ free flow filtrate is the standard
  - ◆ no control and no restriction in filtrate line
  - ◆ permeability is low enough so that only a portion (<30%) of the feed flow converts to filtrate
- All MF processes and UF >100kD
  - ◆ filtrate flow must be regulated (restricted) to avoid most or all of the feed flow converting to filtrate
  - ◆ minimizes fouling, polarization, and over-concentration
  - ◆ results in pressure on the filtrate lines, reduced TMP
  - ◆ NEVER use with standard cellulose membrane!

# Filtrate Control Options

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- Regulate the filtrate flow to a fixed, robust setpoint throughout a process through the use of a:
  - Pump
  - Flow Control Valve
- Flux setpoint is most common
  - ◆ TMP setpoint is also possible but less robust, especially for MF

# Level Control

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- Maintain level in recycle tank constant throughout a fed-batch UF or a DF step
- Modulate either a transfer pump speed or a flow control valve position to maintain either:
  - ◆ Constant recycle tank level (+/- deadband)
  - ◆ Transfer flowrate equal to filtrate flowrate
- Considerations
  - ◆ Effects of pressurizing transfer tank contents
  - ◆ Degree of control required
  - ◆ Confidence in equivalency of 2 different flowmeters

# Useful Equations

| Process Step   | Concentration  | Yield   |
|--|--|---|
| <i>Concentration Step</i>                                    | $C_R = C_o \left( \frac{V_o}{V} \right)^r = C_o (X)^r$ | $Yield_{Retentate} = \frac{(V_R C_R)}{V_o C_o} = X^{-(1-r)}$                  |
| <i>Constant Volume Diafiltration Step</i>                    | $C_{RN} = C_R e^{-(1-r)N}$                             | $Yield_{Retentate} = \frac{(V_{RN} C_{RN})}{V_R C_R} = e^{-(1-r)N}$           |
| <i>Concentration plus constant Volume Diafiltration Step</i> | $C_{RN} = C_o e^{[-(1-r)N + r \ln X]}$                 | $Yield_{Retentate} = \frac{(V_{RN} C_{RN})}{V_o C_o} = e^{-(1-r)(N + \ln X)}$ |

*Membrane Area:*  $FS \cdot Area = ES \cdot Area \times \frac{FS \cdot Volume_{Filtered}}{ES \cdot Volume_{Filtered}} \times \frac{ES \cdot Time}{FS \cdot Time} = \frac{Avg \cdot FS \cdot production \cdot rate}{Avg \cdot ES \cdot flux}$

*Number of Pump Passes:*  $NPP = \frac{N + \ln X}{\left( \frac{Q_P}{Q_F} \right)}$